



Review Article

Bioactive Phytoconstituents and Medicinal Properties of Jamun (*Syzygium cumini*)



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Abstract

Natural products have been used effectively to treat different ailments since the advent of human history. Angiosperms contain numerous bioactive molecules that have been applied as medicines to treat various human diseases, including cancer. Jamun (*Syzygium cumini*) is an angiosperm belonging to the Myrtaceae family. This comprehensive review on Jamun includes information collected from Google Scholar, SciFinder, PubMed, ScienceDirect, and other websites on the internet, giving an account of its botanical profile, chemical composition, and medicinal properties. Ethnomedicinally, various parts of Jamun are used to treat various conditions and have been administered since ancient times in Ayurveda to treat arthritis, obesity, urinary diseases, asthma, bowel spasms, stomach pain, flatulence, diabetes, and dysentery. Several scientific studies also have demonstrated the pluripotent medicinal properties of Jamun, including anti-oxidant, anti-allergic, antiretinitis, antipyretic, antidiarrheal, antinociceptive, anticancer, antidiabetic, anti-obesity, antihyperlipidemic, anti-inflammatory, antimicrobial, diuretic, cardio-protective, chemopreventive, gastroprotective, immunomodulatory, hepatoprotective, wound healing, anthelmintic, and radioprotective. Jamun contains alkaloids, anthraquinones, catechins, cardiac glycosides, flavonoids, glycosides, steroids, phenols, tannins, and saponins. Numerous active phytochemicals have been isolated from its roots, stems, leaves, flowers, fruits, and seeds. Jamun increases glutathione, glutathione peroxidase, catalase, and superoxide dismutase expression and reduces lipid peroxidation levels to exert its beneficial effects on important organs and tissues. Jamun also protects against DNA damage induced by toxic agents including metals, chemicals and ionizing radiation. Jamun activates peroxisome proliferator-activated receptors alpha and gamma and increases fatty acid and glucose metabolism. Additionally, Jamun suppresses various genes at the molecular level. Thus, the scientific evaluation of Jamun is a step forward in validating its traditional use to treat various disorders and may pave the way for translational research for its medicinal use.

Introduction

Angiosperms are a diverse group of plants encompassing more than 300,000–400,000 species, which represent about 80% of all living green plants on Earth.^{1,2} The World Health Organization has identified 21,000 medicinal plants, and India is an abode for 2,500 important medicinal plants in the world.³ Angiosperms have the unique ability to synthesize numerous bioactive compounds that can be of medicinal and healthcare importance to humans.¹ The use of plants and natural products dates back at least 5,000

years, and the Atharva Veda lists at least 50 plants used for the treatment of different ailments.^{4,5} The Myrtaceae family consists of 121 genera and 5,800 species of perennial trees and shrubs that are distributed widely in the subtropical and tropical regions of the world. This family consists of plants that bear edible fruits, which are a berry type with pulp. The plants have a characteristic aroma and are of great agro-industrial importance.^{6,7}

Jamun is native to India and has 400–500 varieties; however, only a few varieties of Jamun produce edible fruits. Jamun can be found along the roadside in tropical and subtropical regions in the Indian subcontinent. It has a remarkable ability to adapt to a variety of climatic conditions, including alkaline soils (pH 10.5); therefore, it is widely planted in semi-arid regions. Jamun is grown as a minor commercial crop in different parts of India and throughout the world for its fruits/timber. Usually, two varieties of Jamun, namely Rama Jamun and Raja Jamun, are cultivated in northern India for their fruits and seeds. These varieties have small seeds and large quantities of pulp. Another seedless variety is commonly grown in Varanasi. The Jamun fruits have a sub-acidic spicy fla-

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vor and are used to prepare squashes, juices, jams, jellies, pickles, wines, and cookies. Jamun fruit squash is a highly refreshing drink to quench thirst during the summer. Jamun is a rich source of various nutrients like proteins and iron.⁸ The alkaloid jambosin, the glucoside jambolin, and antimalin from its seeds reduce the conversion of starch and sugars. Seeds (10–47% of the total mass) are one of the important byproducts left after the processing of Jamun. The seeds are a rich source of minerals, carbohydrates, proteins, lipids, and vitamins, and they serve as important ingredients for the food industry. Jamun syrup cures diarrhea, and Jamun vinegar has cooling, carminative, diuretic, stomachic, and digestive properties. The foliage of Jamun is used as feed for cattle, while Jamun wood is used in buildings, agricultural implements, and railway sleepers.^{9–11}

Natural products have been used to treat several human disorders since time immemorial. Trial-and-error experimentation for many centuries by traditional medicinal practitioners has established the medicinal importance of natural products in the treatment of human diseases and their nontoxic nature. The medicinal activities of plants and herbs are due to their ability to synthesize numerous secondary metabolites that are medicinally bioactive once they reach into the human body. This is reflected in the fact that 75% of modern marketed drugs have a natural origin and are used for the treatment of almost all human diseases, including infectious diseases, inflammatory conditions, cardiovascular disorders, and even cancer.^{12,13} Ayurveda is an ancient system of medicine that mainly uses natural products for the treatment of all diseases.^{14,15} The application of scientific research and new modern analytical methods, including genome profiling, molecular profiling, chemoprofiling, and metabolite fingerprinting, in natural products research has established the utility of natural products in the treatment of various human diseases. Furthermore, the advent of molecular biology techniques and their application in natural product research has provided new insights into the molecular mechanisms of action of natural products in various human disorders.^{16,17} This comprehensive review on Jamun includes information collected from Google, Google Scholar, SciFinder, PubMed, ScienceDirect, and other websites on the internet as well as several individual publications. Importantly, this review focuses on the chemical composition, pharmacology, and medicinal properties of Jamun (*Syzygium cumini*), which has been used to treat various human disorders in India since Vedic times.

Profile of Jamun

The scientific and colloquial names of Jamun in different languages of the world are listed in Table 1.^{18–21}

Botanical description

The scientific classification of Jamun is as follows: kingdom, Plantae; subkingdom, Viridiplantae; infrakingdom, Streptophyta; division, Tracheophyta; subdivision, Spermatophytina; infradivision, Angiospermae; class, Magnoliopsida; superorder, Rosanae; order, Myrtales; family, Myrtaceae; genus, *Syzygium*; and species, *cumini*. Jamun grows profusely in the Indian subcontinent, especially India, Ceylon, Bangladesh, Pakistan, Myanmar, and Madagascar. The well-drained, deep, and loamy soils are most suitable for the growth of Jamun. The USA, Israel, and the West Indies grow Jamun for its fruits and timber.^{21–23} It takes 40 years for the Jamun tree to become fully grown. Jamun reaches a height of 100 feet (30 m), and its canopy spreads up to 36 feet (11 m) with a trunk diameter of 2–3 feet (0.6–0.9 m) (Fig. 1). Jamun branches out at a

short distance from the ground, and its stem bark is discolored at the lower end and becomes smooth and light gray at higher levels. The stem bark of Jamun is rough, cracked, and flaking (Fig. 1). The leaves are 8–10 inches long, up to 4 inches wide, oblong, oval, or elliptically shaped, opposite, blunt, or tapering at the apex, and grow 5–10 inches long (5–25 cm) (Fig. 1). The leaves are pink colored when young and become leathery, glossy, and dark green above, and lighter beneath with yellowish midrib when fully mature with a turpentine smell. Jamun flowers are scented, occur in clusters of a few or 10–50 or more, and each cluster is 1–4 inches long (2.5–10 cm). The flowers are round to oblong, funnel-shaped, 1/2 inch (12.7 mm) wide, and 0.16 inch (4 mm) long, and they bloom during March–April. The flowers bear 4–5 petals, which are united as a small disk with a toothed calyx (Fig. 1). The flowers are greenish-white when young and become rose-pink later.^{10,21,24} Jamun begins to fruit in June–July, and the fruits ripen in the summer. The fruits are round to oblong, initially green colored but become light to dark purple or even black colored when fully ripened, and their size varies between 1/2 and 2 inches (1.2–5 cm) and 1–2.8 cm wide (Fig. 2).^{10,20,21,23,24} Jamun fruits are sweetish sour in taste, and eating Jamun fruits turns the tongue purple. The seeds of the Jamun are oblong in shape, whitish purple colored, and turn brown when dried (Fig. 2). The Hindus and Buddhists consider Jamun a holy tree, and it is commonly grown in the compounds of Hindu temples. The Jamun fruits and leaves are commonly offered to Lord Ganesha (Elephant God) during worship, and it is loved by Lord Krishna (*Jamboo phala saara bhakshitam*).²⁰

Phytochemical analysis

Different parts of Jamun, including the roots, stems, leaves, fruits, and seeds, are reported to synthesize several phytochemicals (Table 2).^{25–48} The water, ethanol, chloroform, ethyl acetate, hexane petroleum ether, and methanol extracts of Jamun leaves contain high-to-low amounts of anthraquinones, alkaloids, cardiac glycosides, flavonoids, glycosides, phenols, steroids, saponins, tannins, proteins, triterpenoids, phytosterols, mucilage, amino acids, carbohydrates, fixed oils, volatile oil, terpenoids, and fats. Additionally, Jamun leaves contain calcium, copper, iron, magnesium, manganese, nitrogen, phosphorus, potassium, sulfur, and zinc as micronutrients.^{25–32} Carbohydrates, flavonoids, terpenoids, and tannins have been reported to be present in the Jamun ethanol leaf extract; whereas only carbohydrates have been detected in the ethyl acetate, methanol, and chloroform extracts.³³ The acetone extract of Jamun leaf contains glycosides, phenols, resins, saponins, and proteins; while flavonoids and alkaloids have been detected in the acetone stem bark extract. Flavonoids, alkaloids, glycosides, phenols, saponins, resins, and proteins have been reported in the acetone root extract. Alkaloids, steroids, and proteins are present in the chloroform Jamun leaf and root extracts. The Jamun seed extract has shown the presence of phenols, alkaloids, tannins, carbohydrates, and proteins; while alkaloids and tannins have been detected in the chloroform stem bark extract. The Jamun leaf and stem bark methanol extracts contain glycosides, flavonoids, alkaloids, phenols, saponins, steroids, tannins, resins, and carbohydrates; while the root extract contains proteins in addition to all other phytochemicals. The stem bark extracted in *n*-hexane showed the presence of alkaloids, proteins, and tannins; but only alkaloids were detected in the leaf extract. The root extract has been reported to contain alkaloids, resins, and carbohydrates; whereas the seed extract contains proteins.³⁴ Flavonoids, glycosides, phenolics, saponins, tannins, triterpenoids, steroids, lipids,

Table 1. Names of Jamun (*Syzygium cumini*) in various languages¹⁸⁻²¹

S. No.	Language/country	Names
1	Scientific names	<i>Syzygium cumini</i> (L.) Skeels, <i>Syzygium jambolana</i> (Lam.) DC., <i>Calyptanthes oneillii</i> Lundell, <i>Eugenia cumini</i> Druce, <i>Syzygium jambolanum</i> DC, <i>Syzygium caryophyllifolium</i> (Lam.) DC., <i>Eugenia djouat</i> Perr. <i>Calyptanthes jambolana</i> Willd. <i>Eugenia caryophyllifolia</i> Lam., <i>Eugenia jambolana</i> Lam., and <i>Myrtus cumini</i> L.
2	English	Indian blackberry, black plum, jambolan, Java plum, purple plum, Malabar plum, jambul, jamblang, Damson plum, Duhat plum, Jambolan plum, rose-apple and Portuguese plum
3	Hindi	Jamun, jaman, duhat and jam
4	Sanskrit	Jambu, jambuphalam, phalendra, mahaskandha, raja-jambuh, or meghamodini
5	Prakrit	Jambu in Pali; jambulo, and jammulo
6	Assamese	Jamu and kala jamu
7	Bengali	Kala jam
8	Gujrati	Jambu, and jaambu
9	Kannada	Nerale hannu, jambunerale, jumnerale, nainerale, jambuva, naayinaerale and neeram
10	Manipuri	Gulamchat and jam
11	Malayalam	Gnaval, naga, naivil, palamper, perinnaralnjara, njaval, perin-njara, and naval-pazham
12	Mizo	Hmuipui and Lenhmui
13	Oriya	Jam, jaman, jambul, rajale, rajjambula and thorajambula in Marathi; Jamkoli
14	Punjabi	Jaman
15	Tamil	Areconitamaram, arugadam, arukatam, caccanam, cattuvalam, nampu, neretu, kavarkalimaram, turavam, and turkolum
16	Tangkhul	Chomshathei
17	Telegu	Goyya-pandu, jam-pandu, jamba, jambu, naredu and raacahnaeredu
18	Urdu	Jaman, jamun and poast jamun
19	Brazil	Azeitona, jambol, jambulao, jamelao, Murta and jalao
20	Cambodia	Pring bai
21	Khmer	Pring bai, Pring das krebey
22	Cook Islands	Paramu (Aitutaki); Damson plum in Jamaica; Pistati and ka'ika
23	France	Jamélongue, jambolanier, jamelongier, faux-pistachier and jamelon-guier
24	Germany	Jambolanapflaume, rosenapfel and wachsjambuse
25	Fiji	Duhat in Guam; Kavika ni India and jammun
26	Indonesia	Jambhool, Duwet, Djoowet, and jamblang
27	Bali	Jambul, Jambulan, Jambulana, Jumbul, Jiwat, Juwet
28	Italy	Pomo della Malesia and Aceituna dulce
29	Laos	Va
30	Japan	Janboran, Murasaki futo momo, Madan
31	Madagascar	Rotra
32	Malaysia	Jambolan, obah jambulana, and jiwat
33	Nepal	Jaamun, kaalo jaamun, phanir, jaambu and jamunaa
34	New Caledonia	Jamelonguier
35	Palau	Mesekerrak and mesigerak
36	Philippines	Lomboi and Duhat
37	Portugal	Jambleiro and jambolão

(continued)

Table 1. (continued)

S. No.	Language/country	Names
38	Russia	Sitsigiui dzhamboza
39	Spain	Guayabo pesgua and yambolana
40	Costa Rica	Ciruelo de Java, and Jambolan
41	Sri Lanka	Jambu, jambul, madan and naval
42	Surinam	Koeli, jamoen and druif
43	Sweden	Jambolanäpplein
44	Thailand	Thabyay-hypyoo in Myanmar; Hakhiphae, lukwa, ma-ha, and wa
45	Tibet	Dza mbu, dzam-bu, and ka ka dz mbu in
46	Kenya, Uganda and Tanzania	Msambarau and mzambarau
47	Venezuela	Guayabo pesjua and pesjua extranjera
48	Vietnam	Va in Laos, Trâm môt, and voi rung
49	West Indies	Indian blackberry in Jamaica; Jambol

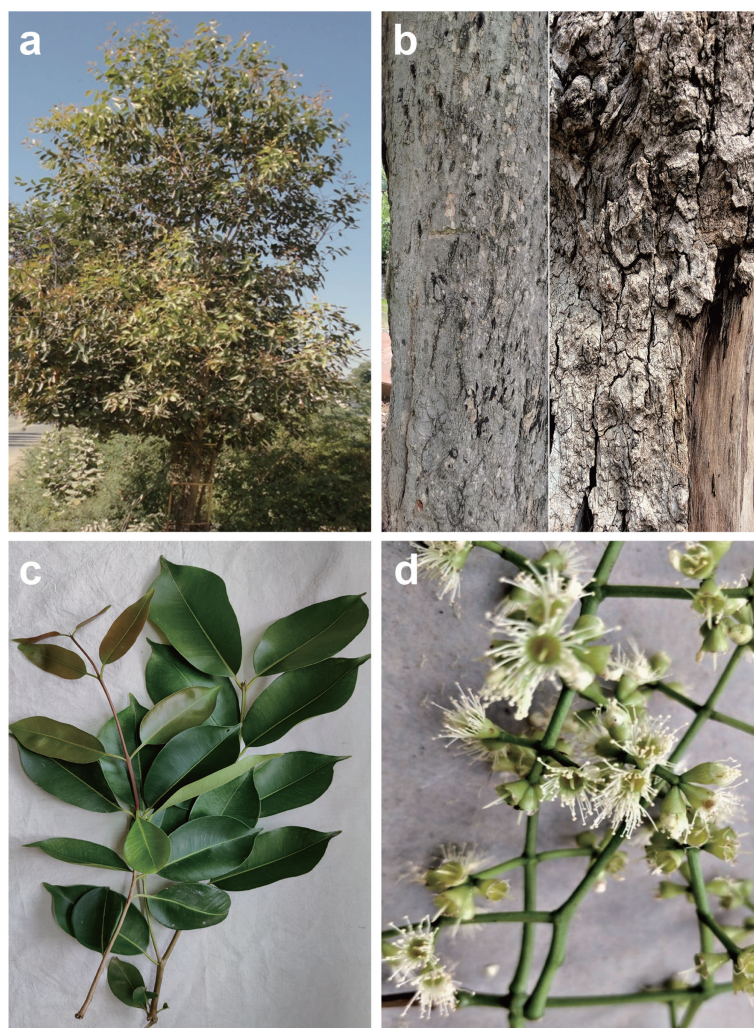


Fig. 1. Photos of Jamun (*Syzygium cumini*). (a) Tree in its natural habitat; (b) Matured stem and bark; (c) Leaves; and (d) Flowers.



Fig. 2. Photos of Jamun (*Syzygium cumini*) fruit. (a) Fruits in their natural form on the tree; (b) Ripened fruits; (c) Fresh seeds; and (d) Dried seeds.

and sugars were detected in the aqueous and alcoholic leaf extracts of Jamun.^{35,36} Catechins, flavonoids, phenols, and quinones are present in the aqueous Jamun stem bark extract.³⁷ The methanol extract of Jamun fruits and its subsequent extraction in *n*-hexane, chloroform, ethyl acetate, and *n*-butanol exhibited the presence of alkaloids, flavonoids, glycosides, steroids, phenols, saponins, tannins, fatty acids, carbohydrates, and reducing sugars.³⁸

Alkaloids, flavonoids, phytosterols, phenols, saponins, tannins, and amino acids have been found in the aqueous and methanol extracts of Jamun seeds.³⁹ The analysis of ethanol, methanol, and water extracts of Jamun seeds has shown the presence of glycosides, alkaloids, proanthocyanidins, flavonoids, terpenoids, phe-

nol, steroids, saponins, tannins, reducing sugars, and reducing monosaccharides. However, glycosides, alkaloids, steroids, and saponins have not been found in the chloroform extract. Proanthocyanidins, flavonoids, tannins, phenols, reducing sugars, and monosaccharides have been detected in the Jamun seed diethyl ether extract. Flavonoids and steroids have been detected in the *n*-hexane extract of Jamun seeds; whereas the benzene extract has shown the presence of terpenoids in addition to flavonoids.⁴⁰ Alkaloids, flavonoids, glycosides, saponins, steroids, triterpenoids, and tannins have been detected in the ethyl acetate and methanol extracts of Jamun seeds.⁴¹ Anthocyanins, alkaloids, flavonoids, cardiac glycosides, proanthocyanins, phenols, saponins, terpe-

Table 2. Phytochemical constituents of Jamun (*Syzygium cumini*)

S.No.	Parts used	Extract type	Phytochemicals	Reference
1	Leaves	Methanol, ethanol, aqueous, chloroform petroleum ether, acetone and hexane	Alkaloids, anthraquinones, flavonoids, glycosides, steroids, phenols, tannins, saponins, phenols, steroids triterpenoids, cardiac glycosides, phytosterols, resins	25–36,47,48
2	Stem bark	Aqueous, ethanol, methanol, hexane and chloroform	Terpenoids, flavonoids anthraquinone glycosides, alkaloids, catechins, phenols, quinones, saponins, phytosterols, quinones steroids tannins and amino acids and flavonoids,	34,35,37
3	Seed and fruit pulp	Ethyl acetate, methanol, ethanol, hydroalcoholic	Alkaloids, anthocyanins, tannins, cardiac glycosides, flavonoids, phenols, terpenoids, glycosides, steroids, saponins, reducing monosaccharides, reducing sugars and proanthocyanidins, proanthocyanins, saponins, steroids, tannins and phenolic, amino acid, and phytosterols	36–46
4	Root	Hexane, Aqueous	Alkaloids and resins and carbohydrates Volatile oil, alkaloids, flavonoids, glycosides, saponins, steroids, tannins, terpenoids, carbohydrates and mucilage	32,34

noids, tannins, steroids, phytosterols, and amino acids have been found in the ethanol extract of Jamun fruit pulp and seeds.⁴²⁻⁴⁴ Flavonoids, alkaloids, tannins, and steroids have been detected in the hydroalcoholic Jamun seed extract.⁴⁵ Glycosides, alkaloids, flavonoids, steroids, saponins, tannins, volatile oils, terpenoids, carbohydrates, and mucilage have been reported in the aqueous root extract of Jamun.³² The aqueous and methanol extracts of Jamun seeds showed the presence of alkaloids, flavonoids, phenols, saponins, tannins and terpenoids. However, the terpenoids were absent in the aqueous extract.⁴⁶ The leaves of Jamun have been found to contain alkaloids, flavonoids, glycosides, resins, tannins, saponins and protein.^{47,48}

Specific active phytochemicals

Jamun synthesizes numerous specific chemical molecules, which have been isolated from various parts including the leaf, stem, roots, flowers, fruits, and seeds (Table 3).^{32,45,49-87} The medicinal activity of Jamun may be due to these phytochemicals. The chemical structures of some important active bioactive secondary metabolites synthesized by Jamun are depicted in Figure 3.

Root

Flavonoid glycosides, isorhamnetin 3-*O*-rutinoside; myrecetine glycoside (5-((3,4-dihydroxy-5-(((3,4,5-trihydroxytetrahydro-2-furanyl)methoxy)methyl)tetrahydro-2-furanyl)methoxy)-3,7-dihydroxy-2-(3,4,5-trihydroxyphenyl)-4H-4-chromenone; myrecetine gentobioses (2-(3,4-dihydroxy-5-(((3,4,5-trihydroxy-6-(((3,4,5,6-tetrahydroxytetrahydro-2H-2-pyranyl) methoxy) methyl)tetrahydro-2H-2-pyranyl)methoxy)phenyl)-3,5,7-trihydroxy-4-oxo-4H-chromenium); 4-(2-amino-2-(2-(2-hydroxy-3 methylbutyl) octahydropyrrolo [1, 2-*a*]pyrazin-7-yl) ethyl)-2-ethyl-phenol; 9-((2-hydroxy-5-m-tolylpentan-2-yloxy)methyl)-2,10-dimethoxy-icosahydro-1*H*-phenanthro[2,1-*f*]chromene-1,9-diol; (*E*)-1-(3-aminophenyl)-7-hydroxy-6-methoxy-3-methyl-7-(1,3,11-trimethoxy-2,4,4,13,14-pentamethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthren-17-yl)oct-4-en-3-yl acetate; 6-(6-acetyl-2-(11-acetyl-8a-(1-amino-ethyl)-4,4,6a,6b,11-pentamethyl-14-oxo 1,2,3,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14,14a,14b-icosahydro-picen-3-yloxy)-4,5-dihydroxy tetrahydro-2*H*pyran-3-yloxy)-3,4,5-trihydroxytetrahydro-2*H*-pyran-2-carboxylic acid; 14-hydroxy-11-methoxy-10-(2-methoxypropoxy)-4,6a,6b, 12,14b-pentamethyl-8a-(methylamino)-4-(1-(2,4,8-trimethyl-2,5,6,7,8,8a-hexahydro-1*H*-pyrido [3,4-*d*1,3]oxazin-6-yloxy)ethyl)octadecahydro-1*H*-phenanthro[1,2-*h*]isochromen-3(4*H*)-one and 6-(17-(4,6-dihydroxy-5-methoxy-2-(methylperoxy)-tetrahydro-2*H*-pyran-3-yloxy)-1,12-di-methoxy-4,5,8,10,12,13,14,17-octahydro-1*H*-cyclopenta[*a*]phenanthren-3-yloxy)-2-(methylperoxy)-3,4-dihydro-2*H*-pyran-3,4,5-triol have been obtained from the Jamun roots.^{32,49}

Stem

Bergenins; eugenin; friedelin; epi-friedelinol; β -sitosterol, and fatty acid ester of epi-friedelinol have been extracted from the stem bark of Jamun.⁴⁵ In addition, the Jamun stem contains quercetin; kaempferol; myricetin; 11-*O*-galloylbergenin; ellagitannin; betulinic acid; ellagic acid and gallic acid.⁵⁰⁻⁵³ 2-butoxy-ethanol; cyclohexanone; 1,2,3,5-tetramethyl-benzene; cyclohexasiloxane, dodecamethyl; 2-butenic acid, 2-methyl-1,1a,1b,4,4a,5,7a,7b,8,9-decahydro-4a,7b-dihydroxy-3-(hydroxymethyl)-1,1,6,8-tetramethyl-5-oxo-9*H*-cyclopropa[3,4]- benz[1,2-*c*]azulene-9,9a-diyl ester, [1*a*R-[1*a*.alpha.,1*b*.beta.,4*a*.beta.,7*a*.alpha.,7*b*.alpha.,8*a*.alpha.,9*b*.beta. (*E*); 9*a*.alpha. (*E*)]]; 2,4-imidazolidinedione, 5-[3,4-bis(trimethylsilyl)oxy]phenyl]-3-methyl-

5-phenyl-1-(trimethylsilyl)-; psi.,psi.-carotene,3,3', 4,4'-tetrahydro-1,1',2,2'-tetrahydro-1-hydroxy-1'-methoxy-; 9,10-anthracenedione, 1-(methylamino)-4-[(4-methylphenyl)amino]; acetic acid, 1,1',4'-triacetoxy-5,5'-diisopropyl-6,7,6',7'-tetramethoxy-3,3'-dimethyl-2,2']binaphthalenyl-4-yl ester; 3,9.beta.;14,15-diepoxy-pregn-16-en-20-one,3,11.beta.,18-triacetoxy-; canthaxanthin; cephalontaxine, 11-(acetyloxy)-, acetate (ester), (11*a*.); 1*H*-cyclopent[*c*]isoxazole, 1-[2,3:5,6-bis-*O*-(1-methylethylidene)- α -*D*-mannofuranosyl]hexahydro-4,5,6-tris(phenylmethoxy)-, [3*a*R-(3*a*.alpha.,4*a*.alpha.; β : 9-15 desoxo-9-x-acetoxy-3,8,12-tri-*O*-acetyl-lingol; spiro[9,9']-difluorene, 2,2'-(2,5,8,11-tetraoxadodecane-1,12-diyl)-; 3,8,12-tri-*O*-acetyl-lingol 7-phenylacetate; 2*H*-1,4-benzo-diazepin-2-one,7-chloro-1,3-dihydro-1-methyl-5-[4-[(trimethylsilyl)oxy]phenyl]-; α -lumi colchicine; pregn-16-en-20-one,11,18-bis(acetyloxy)-3,9-epoxy-3-methoxy-, (3*a*.5. β .11*a*.); 3-hydroxybromoazepam, bis(trimethylsilyl)-deriv; 6,6'-diacetyl-7,7'-dihydroxy-2,2',4,4',5,5'-hexamethoxy-1,1'-binaphthalene; pregnane-11,20-dione, 3,17,21-tris[(trimethyl-silyl)oxy]-, 20-[*O*-(phenylmethyl)oxime], (3*a*,5*a*)- and silane, [(3. β .5*a*.11. β .20*S*)-pregnane-3,11,17,20,21-pentayl]pentakis(oxy)]pentakis(trimethyl)- have been extracted from the methanol stem bark extract of Jamun.⁵⁴

Leaves

Jamun leaves show the presence of myricetin; mycaminose; myricetin 3-*O*-(4"-acetyl)- α -*L*-rhamnopyranosides; *n*-nonacosane; noctacosanol; quercetin; *n*-dotricanol; *n*-hentriacontane; *n*-heptacosane; β -sitosterol; *n*-triacontano; betulinic acid; catechol (maslinic) acid; eicosane; octacosane; octadecane; quercetin 3-*O*-rutinoside; prenylbenzoic acid 4- β -*D*-glucoside; morolic acid 3-*O*-caffeate; 5,4'-dihydroxy, 7-methoxy, 6-methylflavone; 3,4,5-trihydroxybenzoic acid, isetin-7-*O*- β -*D*-glucopyranoside, and (4'-hydroxy-3'-methoxyphenol- β -*D*-[6-*O*-(4"-hydroxy-3",5"-dimethoxybenzoate)]glucopyranoside).⁵⁵⁻⁵⁸ Diferulic acid; butin; methyl gallate; cyanidanol; kaempferide, 4'-hydroxyflavan; taxifolin; palmitic acid; punicic acid; cedrol; caffeic acid; 3 (3hydroxyphenyl) propanoic acid; xanthoxylin; ferulic acid; quinic acid; astragalin; 6-*O*-feruloyl-*D*-glucose; gallic acid; isoquercetin, and 3,5,7,4'-tetra-hydroxy- 6-(3-hydroxy- 3-methylbutyl) flavone have been isolated from the aqueous Jamun leaf extract.⁵⁹

Essential oils

Jamun leaves contain several essential oils, including alloocimene; cineole; caryophyllene; caryophyllene oxide; *L*-limonene; eucarvone; geranyl acetone; α -myrtenal; pinocarvone; pinocarveol; α -terpeneol; myrtenol; muurolol; α -pinene; α -terpineol; α -bornyl acetate; 2- β -pinene; α -humulene, and α -terpineolene.⁶⁰⁻⁶² (*E*)-caryophyllene; α -humulene; α -zingiberene; hydroxytoluene butylated; caryophyllene alcohol; caryolan-8-ol; thujopsan-2- α -ol, and *n*-heneicosane are also present in Jamun leaves.⁶³ The presence of δ -cadinene; τ -cadinol; τ -muurolol; β -eudesmol; globulol; β -pinene; γ -cadinene; camphene; α -terpinenol; camphor; humulene 6,7-epoxide; cubeban-11-ol; α -muurolene; epicubenol; α -copaene; viridiflorene; β -guanine; β -bourbonene; terpinen-4-ol; endoborneol; levoverbenone isobornyl acetate, and 4-methylene-2,8,8-trimethyl-2-vinyl-bicyclo[5.2.0]nonane also has been confirmed in Jamun leaves.⁶⁴ In addition, Jamun leaves contain β -farnesene; caryophyllenol; β -myrcene; fenchol; cis- β -ocimene; 1,3,6-heptatriene; and 3,5-heptadienal, 2-ethylidene-6-methyl-⁶⁵

Flowers

Different phytochemicals, including kaempferol; isoquercetin; quercetin; myricetin-(quercetin-3-glucoside); isoquercetin

Table 3. Different active phytochemicals isolated from different parts of Jamun (*Syzygium cumini*)

S.No.	Parts	Isolated compounds	Reference
1	Roots	Flavonoid glycosides, isorhamnetin 3-O-rutinoside, myrecetone glycoside (5-(3,4-dihydroxy-5-((3,4,5-trihydroxytetrahydro-2-furanyl)methoxy)methyl) tetrahydro-2-furanyl)methoxy)-3,7-dihydroxy-2-(3,4,5-trihydroxyphenyl)-4H-4-chromenone); myrecetone gentobios (2-(3,4-dihydroxy-5-(3,4,5-trihydroxy-6-(((3,4,5,6-tetrahydroxy tetrahydro-2H-2-pyranyl)methoxy)methyl) tetrahydro-2H-2pyranyl) methoxy) phenyl)-3,5,7-trihydroxy-4-oxo-4H-chromenium) and 4-(2-amino-2-(2-(2-hydroxy-3-methyl butyl) octahydropyrrolo-[1,2-a] pyrazin-7-yl)-ethyl)-2-ethylphenol; 9-(2-hydroxy-5-m-tolylpentan-2-yloxy)methyl)-2,10-dimethoxy-icosahydro-1H-phen-antho[2,1-f]chromene-1,9-diol; (E)-1-(3-aminophenyl)-7-hydroxy-6-methoxy-3-methyl-7-(1,3,11-trimethoxy-2,4,4,13,14-pentamethyl-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthren-17-yl) oct-4-en-3-yl acetate; 6-(6-acetyl-2-(11-acetyl-8a-(1-amino-ethyl)-4,6a,6b,11-pentamethyl-14-oxo 1,2,3,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14a,14b-icosahydricen-3-yloxy)-4,5-dihydroxy tetrahydro-2H-pyran-3-yloxy)-3,4,5-trihydroxytetra-hydro-2H-pyran-2-carboxylic acid; 14-hydroxy-11-methoxy-10-(2-methoxypropoxy)-4,6a,6b, 12,14b-pentamethyl-8a-(methylamino)-4-(1-(2,4,8-trimethyl-2,5,6,7,8,8a-hexahydro-1H-pyrido [3,4-d]1,3,6-dioxazin-6-yloxy)ethyl)octadecahydro-1H-phenanthro[1,2-h]isochromen-3(4H)-one and 6-(17-(4,6-dihydroxy-5-methoxy-2-(methylperoxy)-tetrahydro-2H-pyran-3-yloxy)-1,12-di-methoxy-4,5,8,10,12,13,14,17-octahydro-1H-cyclopenta[a]-phenanthren-3-yloxy)-2-(methylper- oxy)-3,4-dihydro-2H-pyran-3,4,5-triol.	32,49
2	Stem	Bergenins; eugenin; friedelin; epi-friedelanol; fatty acid ester of epi-friedelanol; quercetin; β -sitosterol; kaempferol; myricetin; 11-O-galloylbergenin; ellagitannin; butulinic acid; ellagic acid; gallic acid; 2-butoxyethanol, cyclohexanone, 1,2,3,5-tetra-methyl-benzene, cyclohexasiloxane, dodecamethyl, 2-butenol, 2-methyl-, 1,1a,1b,4, 4a,5,7a,7b,8,9-decahydro-4a,7b-dihydro-oxy-3-(hydroxymethyl)-1,1,6,8-tetramethyl-5-oxo-9aH-cyclopropa [3,4]-benz[1,2-e]azulene-9,9a-divylester,[1aR-[1a. α , 1b. β , 4a. β , 7a. α , 7b. α , 8. α , 9. β , (E)9a. α (E)]]-2,4-imidazolidinedione, 5-[3,4-bis-[(trimethylsilyl) oxy]phenyl]-3-methyl-5-phenyl-1-(trimethylsilyl)-; psi,...psi-carotene,3',4,4'-tetradehydro-1',2,2'-tetrahydro-1-hydroxy-1'-methoxy-, 9,10-anthracenedione, 1-(methylamino)-4-[(4-methylphenyl)amino]; acetic acid, 1,1',4'-triacetoxy-5,5'-diisopropyl-6,7,6',7'-tetramethoxy-3,3'-dimethyl[2,2']binaphthal-enyl-4-yl ester, 3,9. β ;14,15-diepoxy-pregn-16-en-20-one,3,11. β ,1,8-triacetoxy; canthaxanthin; cephalotaxin, 11-(acetoyloxy)-acetate (ester), (11. α .); 1H-cyclo-pent[<i>c</i>]isoxazole, 1-[2,3:5,6-bis-O-(1-methyl-ethylidene)- α -d-mannofuranosyl]-hexa-hydro-4,5,6-tris-(phenylmethoxy)-[3aR-(3a. α ,4. α ,5. β ; 9-15des-oxo-9-x-acet-oxy-3,8,12-tri-O-acetyl-lingol, spiro[9,9]-difluorene, 2,2'-(2,5,8,11-tetraoxadodecane-1,12-diy); 3,8,12-tri-O-acetyl-lingol 7-phenyl-acetate; 2H-1,4-benzodiazepin-2-one, 7-chloro-1,3-dihydro-1-methyl-5-[4-[(trimethylsilyl)oxy] phenyl]; α -lumicol-chicine pregn-16-en-20-one,11,18-bis(acetyl-oxy)-3,9-epoxy-3-methoxy-, (3. α ,5. β ,11. α .); 3-hydroxybromo- azeepam, bis(tri-methylsilyl)-deriv; 6,6'-diacetyl-7,7'-dihydroxy-2,2',4,4',5,5'-hexa methoxy-1,1'-binaphthalene; pregnane-11,20-dione, 3,17,21-tris[[trimethyl-silyl]oxy]-, 20-[O-(phenylmethyl)-oxime], (3. α , 5. α .), and silane [[(3. β ,5. α ,11. β ,20S)-pregnane-3,11,17,20,21-pentayl]-pentakis(oxy)pentakis(trimethyl-	45,50-54
3	Leaves	Myricetin; mycaminose; myricetin 3-O-(4"-acetyl)- α -L- rhamno-pyranosides; n-nonacosane; noctacosanol; quercetin; n-dotricontanol; n-hentriacontane; n-hepatcosane; β -sitosterol; n-triacontanol; betulinic acid; strategolic (maslinic) acid; triterpenoids tannins eicosane; octacosane; octadecane; quercetin-3-O-rutinoside; prenylbenzoic acid 4- β -D-glucoside; morolic acid 3-O-cafate; 5,4'-dihydroxy, 7-methoxy, 6-methyl-flavone; 3,4,5-trihydroxybenzoic acid; isoetin-7-O- β -D-glucopyranoside and (4'-hydroxy-3'-methoxyphenol- β -D-[6-O-(4"-hydroxy-3",5"-dimethoxybenzoate)] glucopyranoside).	55-59
4	Essential oils	Alloocimene; α -cadinol; cineole; caryophyllene; caryophyllene oxide; L-limonene; eucarvone; geranyl acetone; α -myrtenal; pinocarvone; pinocarveol; myrtenol; muurolol; α -pinene; α -terpinol; α -bornyl acetate; 2- β -pinene; α -humulene; α -terpineolene; (E)-caryophyllene; α -humulene; α -zingiberene; hydroxytoluene butylated; caryophyllene alcohol; caryolan-8-ol; thujopsan-2- α -ol; n-heneicosane; τ -cadinol; τ -muurolol; globulol; 6-cadinene; β -eudesmol; β -pinene; γ -cadinene; camphene; α -terpinenol; camphor; humulene 6,7-epoxide; cubeban-11-ol; α -muurolene; epicubenol; α -copaene; viridiflorene; β -guanine; β -bourbonene; terpinen-4-ol; endoborneol; levoverbenone isobornyl acetate; 4-methylene-2,8,8-trimethyl-2-vinyl-bicyclo[5.2.0]nonane; β -farnesene; caryophyllenol; β -myrcene; fenchol; cis- β -ocimene; 1,3,6-heptatriene, and 3,5-heptadienal, 2-ethylidene-6-methyl-	60-65

(continued)

Table 3. (continued)

S.No.	Parts	Isolated compounds	Reference
5	Flowers	Kaempferol; isoquercetin; quercetin; myricetin-(quercetin-3-glucoside); isoquercetin (quercetin-3-glucoside); myricetin-3-L-arabinoside; quercetin-3-D-galactoside; ellagic acid; dihydromyricetin; oleanolic acid; acetyl oleanolic acid; eugenol-triterpenoid A and eugenol-triterpenoid B.	66
6	Fruits	Cyanidin 3-glucoside; cyanidin 3,5-diglucoside; delphinidin 3-glucoside; delphinidin 3,5-diglucoside; malvidin 3,5-diglucoside; malvidin 3-glucoside; petunidin 3,5-diglucoside; peonidin 3,5-diglucoside; delphinidin acetyl-diglucoside; petunidin 3-glucoside; dihydromyricetin diglucose ester; dihydro-quercetin diglucose; methyl-dihydromyricetin; gallic acid; dimethyl-dihydromyricetin diglucose; myricetin glucoside; myricetin pentoside; myricetin rhamnoside; myricetin acetyl-rhamnoside; myricetin; cyanidin-3,5-diglycoside; peonidin-3,5-diglycoside; delphinidin-3,5-diglycoside; petunidin-3,5-diglycoside; delphinidin-3,5-diglycoside; petunidin; peonidin; delphinidin di-glucoside; petunidin di-glucoside; malvidin di-glucoside; malvidin-3,5-diglycoside; cyanidin 3-glucoside; cyanidin 3-xyloside; cyanidin 3-rutinoside; cyanidin 3-galactoside; 3-dioxalylglucoside; cyanidin 3-malonyl-glucoside; quercetin 3-rutinoside; quercetin 3-glucoside; quercetin 3-galactoside; quercetin 3-glucosylpentoside; quercetin 3-glucuronide; lambertianin C isomer; quercetin 3-O-[6'-(3-hydroxy-3-methylglutaroyl)]-b-galactoside; quercetin 3-oxalylpentoside; quercetin 3-rhamnoside; quercetin; sanguin H-6 lambertianin A; galloyl-bis-HHDP glucose isomer; ellagic acid; peonidin-3,5-diglucoside; liquiritigenin; scopoletin; umbelliferon; catechin; quinic acid; chlorogenic acid; rosmanol, caffeic acid; glibb-3-ene-1,10-dicarboxylic acid; 2,4-dihydroxy-1-methyl-8-methylene-; 1,4-lactone, 10-methyl ester, (1a,2a,4a,4b,10a)-; 18,19-secoyohimban-19-oi acid,16,17,20,21- tetrahydro-16-(hydroxymethyl)-,methyl ester, (15a,16E)-; carda-5,20 (22)-dienolide, 3-[[6-deoxy-a-L manno pyranosyl)oxy]-14-hydroxy-, (3a)-; spirostan-9-ol,3-amino-, (3a,5a,25R)-; acetic acid,17-acetoxy-3-hydroxyimino-4,4,13-trimethyl- hexadecahydrocyclopenta(a)phenanthren-10-ylmethyl ester; aspidosermidin-17-ol, 1-acetyl-19,21-epoxy15,16-dimethoxy and cholestan-3-ol,2-methylene-, (3a,5a)-.	62,67-78
7	Seeds	Jamboline; 7-hydroxycalamenene; (6'methyl, 2'-1-hydroxyisopropene-1-yl)4,5,6 H pyran; jambosine; methyl-β-orsellinate; β-sitosterol; oleanolic acid; 3-hydroxy androstane [16,17-C]([6'methyl, 2'-1-hydroxy-isopropene-1-yl) 4,5,6 H pyran; hexahydrodiphenyl glucose; hexahydrodiphenic acid; 1-galloylglucose; brevifolin; δ-cadinene; 1,2,3-benzenetriol; bicyclogermacrene; (1a,3a,4a)-3,4-bis(dimethyl(4-methylphenyl)silyl)cyclopentane-1-yl acetate; shahamin B; ellagic acid; ellagitannins; oxirane; β caryophyllene; 2,3-dimethyl, caryophyllene, 5-(hydroxymethyl) 2-furan-carboxaldehyde; 3,7-dimethyl-1,3,6-octatriene; germacrene; 5,10-dichloro-5,10-dimethyl-tricyclo [7.1.0.0(4,6)]decanone; cadinene; 2-isopropenyl-5-isopropyl-7-methylbicyclo[4.1.0]hept-3-ene; 1-methyl-2-methylenecyclo- heptanol; bicyclo[2.2.1]heptan-2-one; 3-(3-butenyl)-2,2-dimethyl cyclopropane carboxylic acid; caryophyllene oxide, bicyclo(4.4.0)decanone; 4-methylene cyclohexane methanol; β-pinenoxide; 8,11,14-eicosatrienoic acid; capric acid; methylheptanoate; 3-thujanol; cis,cis-4,6-octadienol; 5, 9-dimethyl 1-decanol; 2-methylpentanal; hexadecanoic acid (methyl ester); tetradecanoic acid (methyl ester); 4-dodecen-1-ol acetate; 2-methyl pentanol; propyl spirpentane; 3-ethyl-2,2-dimethyl-oxirane; 1,10-decandiol; caryophyllene oxide; 4,11,11-trimethyl-8-methylene; isogeraniol; 3(2H)-furanone dihydroxy-2-methyl; 2-methyl-3-isobutenyl-4-penten-2-ol, 5(hydroxymethyl)-2-furaldehyde; decahydro-4A-methyl-1-methylene-7-(1-methylethenyl); 12-methyl-E-2,13-octadecadien-1-ol, nonadecanoic acid; guaioil; limonene oxide; 3-methyl-4-hexyn-3-ol; thujanol; tannic acid; ellagic acid; gallic acid; caffeic acid; catechin; epicatechin; quercetin; p-coumaric acid; cuscohygrine (alkaloid); naringin; rutin; myricetin (flavonoids); epoxy caryophyllanone (terpenoid); 5-acetamido-4,7-dioxo-4,7-di-hydrobenzofurazan; oxetane, 2,4-dimethyl-,trans-, ethanamine,2-methoxy-, (S)-2-hydroxy-propanoic acid; glycerin; 1-dimethyl- (pentafluorophenyl)silyloxy-cyclopentane; hydroperoxide, 1-methyl-pentyl; benzene-1,2,3-triol; phthalic acid; heptyl pentyl ester, 9,9-dimethoxybicyclo[3.3.1]nona-2,4-dione; 1,1-dodecanediol; hexa-tricotane, nonadecane, 1- chlorodotriacontane, 1-heptadecanamide, cyclohexane, 1-(,1,5-dimethyl-hexyl)-4-(4-methyl-pentyl)-N-hexa-decanoic acid; hexadecanoic acid ethyl ester; t-butyl cyclopentane-epoxy-carboxylate; 5-isopropyl-6-methyl-hepta-,35-dien-2-ol; 2,3-anhydro-D-mannosan; 2,3-anhydro-D-galactosan; α-cadinol; τ-muuroiol; ledene oxide (II); 1,1-dodecanediol diacetate; 1,6,3,4-dianhydro-2-deoxy-β-D-lyxo-hexopyranose; l-hexyl-2-nitrocyclohexane; oleic acid; eicosanoic acid; 1,6,2,3-dianhydro-4-O-acetylβ-D-allopyranose; α-cadinol; pregnan-3,11-di-ol-20-one; propanal,2,3-dihydroxy and diglycerol.	70,79-87

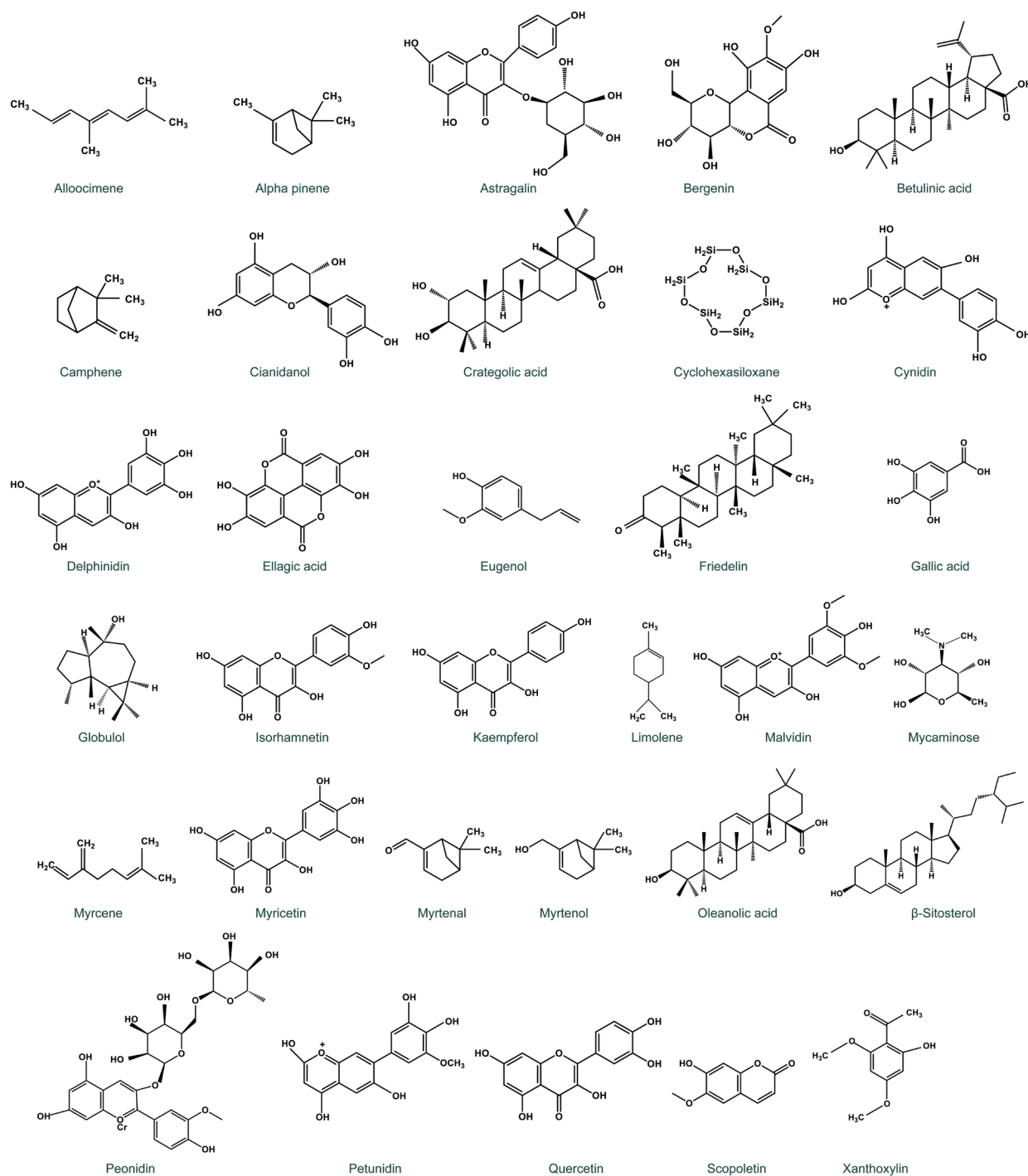


Fig. 3. Chemical structures of some important bioactive phytochemicals present in the different parts of Jamun (*Syzygium cumini*).

(quercetin-3-glucoside); myricetin-3-L-arabinoside; quercetin-3-D-galactoside; ellagic acids; dihydromyricetin; oleanolic acid; acetyl oleanolic acid; eugenol-triterpenoid A, and eugenol-triterpenoid B have been isolated from the flowers of Jamun.⁶⁶

Fruit

Anthocyanins including cyanidin 3,5-diglucoside; delphinidin;

cyanidin 3-glucoside; delphinidin di-glucoside; malvidin; delphinidin 3-glucoside; delphinidin 3,5-diglucoside; malvidin 3,5-diglucoside; malvidin 3-glucoside; malvidin di-glucoside; malvidin-3,5-glycoside; petunidin 3,5-diglucoside; peonidin 3,5-diglucoside; delphinidin acetyldiglucoside; petunidin 3-glucoside; petunidin diglucoside; petunidin-3,5-diglycoside; peonidin, and peonidin-3,5-diglycoside impart a peculiar color to

Jamun fruits. Jamun fruits also contain nonanthocyanin phenolic compounds, including dihydromyricetin diglucoside (flavanol); galloylglucose ester (phenolic acid); methyl-dihydromyricetin; dihydroquercetin diglucoside (flavanol); dimethyl-dihydromyricetin diglucoside (flavanol); gallic acid (phenolic acid); myricetin glucoside (flavonol); myricetin pentoside (flavonol); myricetin rhamnoside (flavonol); myricetin acetyl-rhamnoside (flavonol); and myricetin (flavonol). The fruit pulp also shows the presence of cyanidin 3-glucoside; cyanidin 3-xyloside; cyanidin 3-rutinoside; cyanidin 3-dioxyalylglucoside; cyanidin 3-malonylglucoside; quercetin 3-rutinoside; quercetin 3-glucoside; quercetin 3-galactoside; quercetin 3-glucosylpentoside; quercetin 3-glucuronide; lambertianin C isomer; quercetin 3-*O*-[6''-(3-hydroxy-3-methylglutaroyl)]- β -galactoside; quercetin 3-rhamnoside; quercetin 3-oxalyl-pentoside; quercetin; galloyl-bis-HHDP glucose isomer; sanguin H-6 lambertianin A, and ellagic acid.^{62,67-74} Peonidin-3,5-diglucoside; rosmanol; delphinidin-3-glucoside; myricetin; scopoletin; liquitigenin; umbelliferon; catechin; quinic acid; chlorogenic acid, and caffeic acid also have been detected in the methanol fruit extract.^{75,76} The aqueous fruit extract of Jamun contains gibb-3-ene-1,10-dicarboxylic acid, 2,4-adihydroxy-1-methyl-8-methylene-, 1,4-lactone, 10-methyl ester, (1a,2a,4aa,4b,10a)-, -; 18,19-secoyohimban-19-oi acid, 16,17,20,21-tetrahydro-16-(hydroxymethyl)-methyl ester, (15a,16E)-, -; carda-5,20 (22)-dienolide, 3-[(6-deoxy- α -l-manno-pyranosyl)oxy]-14-hydroxy-, (3a)-, -; spirostan-9-ol, 3-amino-, (3a,5a,25R)-, -; acetic acid, 17-acetoxy-3-hydroxyimino-4,4,13-trimethylhexadecahydro-cyclopenta(a) phenanthren-10-ylmethyl ester; aspidosermidin-17-ol, 1-acetyl-19,21-epoxy-15,16-dimethoxy, and cholestan-3-ol, 2-methylene-, (3a,5a)-. The acidified aqueous extract of Jamun fruit pulp and the peel extract show *p*-coumaric acid; ferulic acid; gallic acid; vanillic acid; caffeic acid; (phenolic acid); epicatechin; catechin; myricetin (flavonoids), 3,5-diglucosides of petunidin; malvidin, and delphinidin anthocyanins.^{77,78}

Seeds

Jamboline; 7-hydroxycalamenene, (6'methyl, 2'-1-hydroxyisopropene-1-yl)4,5,6-*H*-pyran, jambosine; methyl- β -orsellinate; β -sitosterol; oleanolic acid; 3-hydroxy androstane [16,17-*C*] (6'methyl, 2'-1-hydroxyisopropene-1-yl) 4,5,6-*H*-pyran; hexahydroxydiphenyl glucose; hexahydroxydiphenic acid; 1-galloylglucose and brevifolin have been separated from the ethanol Jamun seed extract.^{70,79,80} Jamun seed contains δ -cadinene; 1,2,3-benzenetriol; bicyclo-germacrene; (1a,3a,4a)-3,4-bis[dimethyl(4-methylphenyl)silyl]cyclopentan-1-yl acetate; ellagic acid; shahamin B and ellagitannins.^{70,81} The Jamun methanol seed extract shows the presence of oxirane; β caryophyllene; 2,3-dimethyl, caryophyllene; 5-(hydroxymethyl) 2-furan-carboxaldehyde; 3,7-dimethyl-1,3,6-octatriene; germacrene; 5,10-dichloro-5,10-dimethyl-tricyclo- [7.1.0.0(4,6)]-decane; cadinene; 2-isopropenyl-5-isopropyl-7,7methylbicyclo[4.1.0]hept-3-ene; 1-methyl-2-methylenecycloheptanol; bicyclo[2.2.1]heptan-2-one; 3-(3-butenyl)-2,2-dimethyl cyclopropane carboxylic acid; caryophyllene oxide; bicyclo(4.4.0)decane; 4-methylene cyclohexane methanol; β -pinenoxide; 8,11,14-eicosatrienoic acid; capric acid; methylheptanoate; 3-thujanol, *cis,cis*-4,6-octadienol; 5, 9-dimethyl 1-decanol; 2-methylpentanal; hexadecanoic acid (methyl ester); tetradecanoic acid (methyl ester); 4-dodecen-1-ol acetate; 2-methyl pentanol; propyl spiro-pentane; 3-ethyl-2,2-dimethyloxirane, and 1,10-decandiol. Caryophyllene oxide; 4,11,11-trimethyl-8-methylene; isogeraniol; 3(2*H*)-furanone dihydroxy-2-methyl; 2-methyl-3-isobutenyl-4-penten-2-ol; 5(hydroxymethyl)-2-fural-

dehyde; decahydro-4 A-methyl-1-methylene-7-(1-methylethenyl); 12-methyl-*E,E*-2,13-octadecadien-1-ol, nondecanoic acid, guaiol, limonene oxide, 3-methyl-4-hexyn-3-ol, and thujanol have been detected in the ethanol extract.⁸² The aqueous Jamun seed extract contains tannic acid; gallic acid; ellagic acid; caffeic acid; catechin; epicatechin; quercetin, and *p*-coumaric acid; whereas the ethyl acetate fraction of Jamun seed extract shows the presence of cuscohygrine (alkaloid); naringin; rutin; myricetin (flavonoids), and epoxycaryophyllanone (terpenoid).^{83,84} 5-Acetamido-4,7-dioxo-4,7-di-hydrobenzofurazan; oxetane; 2,4-dimethyl-trans-ethanamine; 2-methoxy-, (*S*)-2-hydroxy-propanoic acid; glycerin; 1-dimethyl-(penta-fluorophenyl)silyloxycyclopentane; hydroperoxide, 1-methylpentyl; benzene-1,2,3-triol; phthalic acid; heptyl pentyl ester, and 9,9-dimethoxy- bicyclo[3.3.1]nona-2,4-dione have been isolated from the methanol Jamun seed extract.⁸⁵ Gas chromatography-mass spectrometry analysis of the Jamun seed ethanol extract led to the isolation of 1,1-dodecanediol; hexatriacontane; nonadecane; 1-chloro-dotriacontane; 1-hepta-decanamide; cyclohexane; 1-(1,5-dimethyl-hexyl)-4-(4-methyl-pentyl); *N*-hexadecanoic acid; hexa-decanoic acid ethyl ester; *t*-butyl cyclopentane eperoxycarboxylate; α -cadinol; τ -muurolol; 5-isopropyl-6-methyl-hepta-,35-dien-2-ol; 2,3-anhydro-Dmanosan; 2,3-anhydro-D-galactosan; ledene oxide (II); 1,1-dodecanediol diacetate; 1,6;3,4-dianhydro-2-deoxy- β -D-lyxohexo-pyranose; 1-hexyl-2-nitrocyclohexane; eicosanoic acid; oleic acid; 1,6;2,3-dianhydro-4-*O*-acetyl β -D-allopyranose; α -cadinol; pregnan-3,1,1-diol-20-one; propanal, 2,3-dihydroxy, and diglycerol.^{86,87}

Nutritional profile

The Jamun fruit pulp has been analyzed for its nutritional value, which may vary depending on the region and season of analysis (Table 4).^{21,88-92} The moisture contents in the Jamun fruit pulp were found to be 79.21–86.12% and $1.03 \pm 0.08\%$, respectively. The Jamun fruit pulp has shown the presence of the following: total soluble sugars, 1.4 ± 0.15 (degree Brix); total dissolved solids, 217 ± 1.15 (ppm); total solids, $47.75 \pm 3.17\%$; total carbohydrates, $89.68 \pm 0.29\%$; total sugars, $5.54 \pm 0.69\%$; fats, $1.28 \pm 0.11\%$; pectin, $4.7 \pm 0.13\%$; proteins, $0.65 \pm 0.03\%$; fiber, $0.18 \pm 0.02\%$; antioxidant capacity (1,1-diphenyl-2-picrylhydrazyl; DPPH), $31.29 \pm 1.53\%$; tannins, 94.52 ± 9.19 (mg/100 g); anthocyanins, 195.58 ± 6.15 (mg/100 g); and polyphenols, 203.76 ± 9.84 (mg/g gallic acid equivalents). The minerals (mg/100 g) present in the Jamun fruit pulp include the following: sodium, 1.73 ± 1.70 – 8.75 ± 0.25 ; potassium, 172.4 ± 17.23 – 358.5 ± 5.0 ; calcium, 54.55 ± 4.78 – 81.4 ± 11.15 ; phosphorous, 152.65 ± 15.38 ; zinc, 0.46 ± 0.06 – 1.215 ± 0.1 ; iron, 4.66 ± 0.81 – 33.2 ± 1.0 ; magnesium, 27.13 ± 3.43 – 166.7 ± 7.10 ; manganese, 0.2 ± 0.007 ; copper, 1.8 ± 0.41 – 8.75 ± 0.25 ; lead, 0.33 ± 0.13 ; chromium, 1.06 ± 0.13 ; sulfur, 13, and chlorine, 8. The following vitamins (mg/100 g) have been detected in the Jamun fruit pulp: ascorbic acid, 49.78 ± 2.17 ; niacin, 8; choline, 7; folic acid, 3 μ g; riboflavin, 0.01–0.06; thiamine, 0.03–0.12; and vitamin A, 80 IU. The pH of Jamun fruit pulp has been measured as 3.87 ± 0.01 , with an acidity of 2.65 ± 0.21 .^{21,88-92}

The moisture and ash contents in Jamun seeds are reported to be $52.24 \pm 3.17\%$ and $3.13 \pm 0.16\%$, respectively. Jamun seeds have shown the presence of the following: total soluble sugars, 1.4 ± 0.15 (degree Brix); total dissolved solids, 217 ± 1.15 (ppm); total solids, $47.75 \pm 3.17\%$; total carbohydrates, $89.68 \pm 0.29\%$; total sugars, $5.54 \pm 0.69\%$; fats, $1.28 \pm 0.11\%$; proteins, $4.68 \pm 0.35\%$; fiber, $1.21 \pm 0.06\%$; antioxidant capacity (DPPH), $48.23 \pm 2.98\%$; tannins, 388.99 ± 7.34 (mg/100 g); anthocyanins, 18.47

Table 4. Nutritional profile of Jamun (*Syzygium cumini*) fruit pulp and seeds^{21,88-92}

S. No.	Parameters	Fruit pulp	Seeds
1	Moisture	79.21 to 86.12%	52.24 ± 3.17% 3.13 ± 0.16%
2	Ash	1.03 ± 0.08%	3.13 ± 0.16%
3	Acidity	2.65 ± 0.21	0.04 ± 0.001
4	pH	3.87 ± 0.01	2.5 ± 0.1
5	Total soluble sugars	14.86 ± 1.47	1.4 ± 0.15 (degree Brix)
6	Total dissolved solids	615 ± 0.21	217 ± 1.15 (ppm)
7	Total solids	20.33 ± 0.34	47.75 ± 3.17
8	Total carbohydrate	97.59 ± 0.09%	89.68 ± 0.29%
9	Total sugars	7.88 ± 0.41%	5.54 ± 0.69%
10	Fats	0.18 ± 0.02	1.28 ± 0.11
11	Pectin	4.7 ± 0.13%	–
12	Proteins	0.65 ± 0.03 %	4.68 ± 0.35%
13	Fiber	0.18 ± 0.02%	1.21 ± 0.06%
14	Antioxidants capacity (DPPH)	31.29 ± 1.53%	48.23 ± 2.98%
15	Tannins	94.52 ± 9.19 (mg/100g)	388.99 ± 7.34 (mg/100g)
16	Anthocyanins	195.58 ± 6.15 (mg/100g)	18.47 ± 1.99 (mg/100g)
17	Polyphenols (mg/g gallic acid equivalent)	203.76 ± 9.84	386.51 ± 10.25
	Minerals (mg/100 g)		
18	Sodium	8.75 ± 0.25- 1.73 ± 1.70	43.86 ± 12.09
19	Potassium	172.4 ± 17.23–358.5 ± 5.0	606.46 ± 69.37
20	Calcium	54.55 ± 4.78–81.4 ± 11.15	135.86 ± 26.81
21	Phosphorous	152.65 ± 15.38	–
22	Zinc	0.46 ± 0.06–1.215 ± 0.1	0.46 ± 0.17
23	Iron	4.66 ± 0.81–33.2 ± 1.0	4.2 ± 0.80
24	Magnesium	27.13 ± 3.43–166.7 ± 7.10	111.6 ± 18.06
25	Manganese	0.2 ± 0.007	0.4 ± 0.11
26	Copper	1.8 ± 0.41–8.75 ± 0.25	2.13 ± 0.86
27	Lead	0.33 ± 0.13	0.66 ± 0.06
28	Chromium	1.06 ± 0.13	1.4 ± 0.61
29	Sulphur	13	–
30	Chlorine	8	–
	Vitamins (mg/100 g)		
31	Ascorbic acid	49.78 ± 2.17	–
32	Niacin	8	–
33	Choline	7	–
34	Riboflavin	0.01 to 0.06	–
35	Thiamine	0.03–0.12	–
36	Folic acid	3 µg	–
37	Vitamin A	80 IU	–

± 1.99 (mg/100 g); and polyphenols, 386.51 ± 10.25 (mg/g gallic acid equivalents). The minerals (mg/100 g) present in Jamun seeds include the following: sodium, 43.86 ± 12.09 ; potassium, 606.46 ± 69.37 ; calcium, 135.86 ± 26.81 ; zinc, 0.46 ± 0.17 ; iron, 4.2 ± 0.80 ; magnesium, 111.6 ± 18.06 ; manganese, 0.4 ± 0.11 ; copper, 2.13 ± 0.86 ; lead, 0.66 ± 0.06 ; and chromium. The pH of Jamun fruit pulp was estimated to be 2.5 ± 0.1 , with an acidity of 0.04 ± 0.001 .⁸⁸⁻⁹⁰

Traditional medicinal uses

For centuries, Jamun has been used in diverse ethnomedicinal practices to treat a variety of human diseases.^{20,93} According to Verse 140 of Charak Sutrasthana, 27 Jamun is medicinally characterized as madhura (sweet), kashaya (slightly astringent), amla (sour) in taste, guru (heavy), ruksha (dry), vishtambhi (producing wind in the abdomen, causing bloating), sheetala (cooling), grahi (absorbent, bowel binding), vatakara (aggravates vata, i.e., subtle energy associated with movement), balances pitta (related to body's metabolism) and kapha (related to body structure), pramehagna (cures urinary diseases including diabetes), and medoroga (obesity).⁹⁴ Jamun purifies the blood and is a good general health tonic for humans. The Jamun stem bark is astringent, constipating, antibacterial, anthelmintic, digestive, carminative, diuretic, febrifuge, stomachic refrigerant, and sweet. In Ayurveda, Jamun is used to treat cold, anorexia, cough, diabetes, worm infestation, emaciation, diarrhea, dysentery, dental, digestive, liver, skin, erectile disorders, and wheezing difficulty. The patients are orally administered with 1–3 g of dried Jamun seed powder to treat diabetes in Ayurvedic medicine. In the traditional system of medicine, various parts of Jamun are used in the treatment of diabetes, dysentery, mouth blisters, digestive complaints, colic, stomachache, cancer, diarrhea, pimples, and piles.⁹⁵ The administration of 2.5–10 mL (half to two teaspoons) of ripe Jamun fruit juice thrice a day to patients cures diabetes.^{96,97} Traditionally, Jamun fruits and seeds are utilized to treat diabetes, bronchitis, asthma, and splenopathy.^{20,24,98-101} The application of the seed powder of Jamun removes blemishes left by acne as well as blackheads on the skin. Eating a mixture of fresh Jamun fruit pulp and honey keeps the body healthy. Enlarged spleen and urinary problems are treated by the use of Jamun fruit juice.^{20,101} Eating Jamun seed powder mixed with jaggery relieves dysentery and diarrhea.²⁰ The topical application of the poultice prepared from Jamun leaf juice is a good remedy to treat skin disorders and dysentery.^{102,103} The bleeding gums can be cured by topical application of Jamun leaf ash, which also keeps teeth healthy.^{20,91} One teaspoon of Jamun seed powder with water prevents bed wetting in children. Metrorrhagia in women can be treated by consuming 3 g of seed powder. The cataract can also be treated by consuming tablets prepared from Jamun seed powder and honey (1:1 ratio) with milk. The decoction prepared from Jamun leaves relieves conjunctivitis.⁹⁷

The Jamun is a liver tonic in Unani medicine, and the application of leaf paste heals wounds. Jamun strengthens teeth and gums, enriches the blood, and disinfects ringworm and head lice infection.^{20,104} Hemorrhoids can be treated by eating Jamun for 2–4 months, and gingivitis is treated with Jamun fruit pulp.²⁰ The decoction of Jamun stem bark, root bark, or dried seeds treats dyspepsia, diarrhea, and dysentery as well as acts as an enema.^{20,103} A mixture of dried stem bark powder and yogurt is used to treat menorrhagia.^{20,96} Jamun stem bark powder mixed with its fruit juice is a good remedy for cough and cold. Furthermore, a mixture of half a teaspoon of stem bark powder in one glass of Jamun fruit juice taken daily is useful against infections of the urinary tract and

urinary diseases. Strychnine poisoning is cured by giving Jamun seed powder in India.^{20,103} Asthma and bronchitis are treated by administering Jamun stem bark decoction.¹⁰⁵ Spongy gums, mouth ulcerations, and stomatitis are treated using Jamun stem bark decoction, which also acts as a good mouthwash.^{20,101,103} A mixture of Jamun stem bark ash and oil is applied to treat burns, and its mixture with water acts as an anti-inflammatory agent.^{20,101} The decoction prepared from Jamun seeds relieves strain and fatigue.²⁰ Looking into traditional uses, various investigators have scientifically investigated the application of different parts of Jamun to exploit its pleiotropic medicinal properties and substantiate the claims of traditional healers.

Toxicity evaluation

Safety evaluation of any drug is critical for human use. As such, Jamun has been reported to be safe in various preclinical models, which also justifies the traditional claim that it is a nontoxic remedy. Single intraperitoneal administration of 100, 200, 300, and 400 mg/kg body weight of 1:1 dichloromethane and methanol leaf extract of Jamun did not induce any changes in animal behavior or mortality; hence, a dose of 400 mg/kg is considered safe. The median lethal dose (LD₅₀) could not be determined because higher doses could not be administered.¹⁰⁶ Similarly, a dose of 1,000 mg/kg body weight was found to be safe in Swiss albino mice administered with 250, 500, 750, or 1,000 mg/kg body weight of 50% hydroethanolic Jamun seed extract as none of these doses could alter the behavior or induce mortality in mice in acute toxicity studies. It was not possible to determine the LD₅₀ as higher doses could not be administered.¹⁰⁷ The methanol leaf extract has been reported to be nontoxic up to 3.5 g in oral acute toxicity studies in mice, with an LD₅₀ of 3.873 mg/kg body weight, and oral administration of hydroalcoholic leaf extract did not induce toxicity up to 6 g/kg body weight in mice. The intraperitoneal administration of methanol leaf extract was toxic, with an LD₅₀ of 489 mg/kg body weight in mice.¹⁰⁸ The oral administration of aqueous extract of Jamun stem bark at doses of 300, 2,000, or 5,000 mg/kg body weight in rats was safe up to 5 g/kg body weight, and repeated oral administration of 300, 1,000, or 2,000 mg/kg body weight of aqueous extract of Jamun stem bark for 28 days did not produce any toxicity in the rats.¹⁰⁹ Likewise, 70% methanol stem bark extract of Jamun was nontoxic up to 5,000 mg given orally in mice, and an LD₅₀ of 3.873 mg/kg was reported for the leaf extract.¹¹⁰

The oral administration of a single dose (acute toxicity) of the ethanol leaf extract of Jamun was nontoxic up to 5 g/kg body weight in rats. Furthermore, the oral administration of 1,250, 2,500, or 5,000 mg/kg for 28 days also did not induce toxicity in rats.¹¹¹ The administration of aqueous Jamun seed extract and polymeric nanoparticles prepared from the seed extract given to Wistar rats was found to be nontoxic in acute toxicity studies as they did not induce mortality or alteration in the biochemical or hematological profiles after 14 days.¹¹² Similarly, the oral administration of Jamun leaves, stem bark, root, or seeds extracted in methanol (except seeds) was found to be nontoxic up to 2,000 mg/kg in rats, and the administration of 100 and 500 mg/kg body weight aqueous leaf extract for 30 days in rats in drinking water was nontoxic as no significant changes were observed in the organ or body weights of male Wistar rats.¹¹³ The 70% ethanol seed extract of Jamun was nontoxic up to 2 g/kg orally in acute toxicity studies in rats, whereas the daily oral administration of 250, 500, or 1,000 mg/kg of extract once daily for 28 days did not produce any toxic effects.¹¹⁴

Antioxidant activity

The increased oxidative stress by various reactive oxygen species is the major cause of numerous human diseases. Jamun has been reported to possess antioxidant activity, where different parts of Jamun passivated free radicals *in vitro*. Concentration-dependent inhibition of nitric oxide (NO[•]) free radicals has been reported for the dichloromethane, methanol, and 50% hydroalcoholic extracts of Jamun leaves and seeds.¹¹⁵ The methanol leaf extract and its ethyl acetate, chloroform, *n*-hexane, and water fractions scavenged DPPH radicals and increased the ferric reducing antioxidant power (FRAP).¹¹⁶ The ethanol extract of Jamun leaves scavenged DPPH radicals and increased the reducing power concentration dependently.¹¹⁷ The dichloromethane and methanol (1:1) extract of Jamun leaves scavenged the radicals of hydroxyl (•OH), superoxide (O₂^{•-}), DPPH, and 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid (ABTS^{•+}), depending on the concentration.¹¹⁸ The dichloromethane and methanol leaf extracts of Jamun passivated DPPH free radical generation and elevated FRAP.¹¹⁹ The generation of •OH, NO[•], and DPPH free radicals was inhibited in a concentration-dependent manner by the methanol and aqueous Jamun leaf extracts, which also increased FRAP in a similar manner.¹²⁰ Concentration-dependent passivation of •OH, NO[•], and ABTS^{•+} radicals has been detected with the acetone and aqueous leaf extracts of Jamun.¹²¹ The methanol extracts of Jamun leaves, fruit pulp, and seeds increased FRAP, and the leaf extract was more effective than the seed and fruit pulp extracts.¹²² The Jamun aqueous leaf and seed extracts reduced DPPH, NO[•] radicals, and H₂O₂ formation as well as increased FRAP, depending on their concentration. The leaf extract was more effective than the seed extract.¹²³ Ethyl acetate:methanol (3:1), ethyl acetate:methanol (1:1), ethyl acetate:methanol (1:3), and pure methanol extracts of Jamun leaves inhibited DPPH radical production.¹²⁴

The formation of •OH, O₂^{•-}, and DPPH free radicals was reduced by the aqueous extract of Jamun fruit skin.¹²⁵ The concentration-dependent alleviation has been detected in the generation of DPPH, •OH, and O₂^{•-} radicals by ethanol extracts of Jamun fruit pulp, kernels, and seed coat. The fruit pulp extract resulted in half-maximal inhibitory concentration (IC₅₀) values of 158 ± 5, 310 ± 10, and 1,703 ± 9 µg/mL for DPPH, •OH, and O₂^{•-} radicals, respectively. The IC₅₀ values for the kernel extract were 8.6 ± 3, 151 ± 5, and 85 ± 5 µg/mL; whereas the IC₅₀ values for the seed extract were 48 ± 9, 261 ± 4, and 759 ± 14,261 µg/mL for DPPH, •OH, and O₂^{•-} radicals, respectively.¹²⁶ The Jamun fruit extract rich in anthocyanin neutralized ABTS^{•+} and peroxy radicals efficiently.⁶⁸ The seed and fruit pulp of Jamun extracted in acidified ethanol inhibited the generation of DPPH and ABTS^{•+} free radicals and also exhibited iron-chelating activity.⁷¹ The methanol stem bark extract of Jamun increased the scavenging of DPPH and •OH radicals as well as the FRAP.¹²⁷ The acetone extract of Jamun seeds passivated the DPPH, NO[•], and ABTS^{•+} radicals as well as H₂O₂ and increased the antioxidant activity and FRAP concentration dependently.¹²⁸

Antibacterial and antifungal activity

Bacterial and fungal infections lead to numerous infectious diseases in humans, and Jamun can act against bacterial and fungal infections. The growth of *Bacillus sphaericus*, *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Salmonella typhimurium* has been reported to be inhibited by the essential oils from Jamun leaves.⁶¹ Antibiotic-resistant bacteria, including *Klebsiella pneumoniae*, *P. aeruginosa*, *S. aureus*, *Enterococcus faecalis*, *E. coli*, *Kocuria rhizophila*, and *Neisseria gonorrhoeae*,

and the fungus *Candida krusei* were effectively killed by the hydroalcoholic extract of Jamun leaves.¹²⁹ The 70% methanol leaf extract and leaf oil suppressed the growth of *E. coli*, *S. aureus*, *Salmonella typhi*, and *Bacillus subtilis* bacteria and *Candida albicans* fungi.¹¹⁰ The growth of *Vibrio cholerae* serogroups *Ogawa* and *Inaba* was suppressed by Jamun leaf ethanol extract.¹³⁰ The methanol and methylene chloride extracts and essential oils from the leaf of Jamun prevented the growth of *E. coli*, *P. aeruginosa*, *S. aureus*, *N. gonorrhoeae*, *B. subtilis*, *S. aureus*, and *E. faecalis*, and the methanol extract was more effective than the methylene chloride extract and essential leaf oils.¹¹⁹ The essential leaf oils were also active against the fungi *Aspergillus flavus*, *Aspergillus niger*, *Fusarium solani*, and *Rhizopus solani*.⁶⁶

The Jamun fruit ethanol extract attenuated the growth of *S. aureus* and *Staphylococcus epidermidis* (Gram-positive) as well as *P. aeruginosa* (Gram-negative) bacteria and *C. albicans*, *C. krusei*, *Candida parapsilosis*, and *Cryptococcus neoformans* fungi, indicating its antibacterial and antifungal potential.¹³¹ Jamun leaf, stem bark, pulp, and seeds extracted in 70% ethanol restricted the growth of *Bacillus cereus*, *B. subtilis*, *S. aureus* (Gram-positive), *P. aeruginosa*, *Shigella flexneri*, and *V. cholera* (Gram-negative) bacteria. The leaf and bark extracts of Jamun were more effective than the pulp and seed extracts.⁴³ *B. subtilis*, *E. coli*, *P. aeruginosa*, and *S. aureus* growth was suppressed by Jamun seed ethanol extract.¹³² The diethyl ether, methanol, and aqueous extracts of Jamun fruit arrested the growth of *S. epidermidis*, *B. cereus*, *Micrococcus luteus*, and *Salmonella typhi*, respectively.⁴⁸

The aqueous extract of the stem and leaves of Jamun was active against *S. aureus*, *E. coli*, *Staphylococcus saprophyticus*, *Proteus vulgaris*, and *P. aeruginosa*, and the fruit extract effectively suppressed the growth of *P. aeruginosa* and the fungi *Penicillium chrysogenum* and *C. albicans*.¹³³ The growth of the Gram-positive *Bacillus amyloliquefaciens* and the Gram-negative *S. aureus* was restrained by the methanol extract of Jamun stem bark; whereas the ethanol extract of Jamun roots was active against *E. coli*, *Streptococcus suis*, *S. aureus*, *S. epidermidis*, *Salmonella* spp., and *Corynebacterium diphtheriae*, and greater growth inhibition was seen for the Gram-positive than the Gram-negative bacteria.¹³⁴ The growth of *E. coli*, *S. aureus*, and *C. albicans* was inhibited by the chloroform and ethyl acetate leaf extracts of Jamun.³⁶ The growth inhibitory activity of aqueous Jamun seed extract and polymeric nanoparticles formulated from seed extract has been reported against *C. krusei*, *C. albicans*, *Candida haemulonii*, *Candida guilliermondii*, and *Cryptococcus* sp.¹³⁵ In addition, the fruit juice of Jamun effectively arrested the growth of *Salmonella typhimurium*, *S. aureus*, *S. flexneri*, and enterotoxigenic *E. coli*; whereas it did not have any effect on *Lactobacillus acidophilus* or *Lactobacillus bulgaricus*.¹³⁶

The ethanol and methanol extracts of Jamun stem bark killed *B. amyloliquefaciens*, *S. aureus*, *E. coli*, and *P. aeruginosa*; whereas the aqueous extract was ineffective against these bacteria.¹³⁷ The Jamun seed extracted in ethanol exhibited antibacterial activity against *Streptococcus agalactiae*, *S. aureus*, *B. cereus*, *E. faecalis*, *Clostridium perfringens*, *Listeria monocytogenes*, *S. typhimurium*, *E. coli*, *P. aeruginosa*, *K. pneumoniae*, and *Aeromonas hydrophila*; however, it was most effective against *B. cereus*.⁴⁴ The ethyl acetate extract of Jamun fruits acted against the Gram-negative bacterial strains *E. coli*, *Salmonella*, *P. aeruginosa*, and *S. typhi* as well as the two Gram-positive strains *S. aureus* and *Streptococcus*.¹³⁸ The aqueous, 80% ethanol, methanol, acetone, and hexane extracts of Jamun leaves arrested the growth of methicillin-resistant *S. aureus* and inducible clindamycin-resistant *S. aureus*, depending on the

concentration; the aqueous extract was the most effective, whereas the hexane extract was ineffective.⁴⁴ The methanol extracts of ripe fruit pulp, ripe fruit seeds, unripe fruit pulp, and unripe fruit seeds attenuated the growth of *E. coli*, *B. subtilis*, and *S. typhi* bacteria as well as *A. niger*, *Penicillium notatum*, and *Alternaria alternata* fungi, where 0.1% was effective against all bacterial and fungal species.¹³⁹ The aqueous and methanol seed extract inhibited the growth of *Salmonella enteritidis*.⁴⁷

Anti-inflammatory activity

Inflammation is related to several diseases, including cancer, and Jamun has been experimentally shown to be active against acute and chronic inflammation. The chloroform and ethanol extracts of Jamun seeds arrested carrageenan (acute) as well as kaolin-carrageenan (subacute)-induced paw edema in rats; moreover, they arrested exudation of proteins, dye leakage in peritoneal inflammation, and leukocyte migration.¹⁴⁰ The anti-inflammatory activity of aqueous, methanol, and ethyl acetate Jamun seed extracts has been reported in human neutrophils and carrageenan-induced rat paw edema, and the extracts also reduced ectonucleotidase, adenosine deaminase (ADA), acetylcholinesterase, and dipeptidyl peptidase IV activities and NO formation.^{141,142} The Jamun stem bark ethanol extract arrested the carrageenan (acute), kaolin-carrageenan (subacute), and formaldehyde (subacute)-induced rat paw edema and chronic cotton pellet granuloma.¹⁴³ Carrageenan-induced paw edema also was attenuated in Wistar rats by the methanol, ethyl acetate, and aqueous extracts of Jamun seeds, with the aqueous extract being more potent than the methanol extract.^{142,144} The Jamun leaves extracted in ethyl acetate and methanol alleviated carrageenan-induced paw edema in Wistar rats.¹⁴⁵

Acute and chronic inflammation was arrested by Jamun leaf methanol extract in histamine, carrageenan, and serotonin-induced rat paw edema as well as cotton pellet-induced granuloma.¹⁴⁶ In addition, the migration of rat eosinophils was attenuated by essential oils from Jamun leaves, indicating their anti-inflammatory potential.¹⁴⁷ The aqueous leaf extract of Jamun decreased the indomethacin-induced inflammatory changes by decreasing the expression of cyclooxygenase 1 and 2 (COX1 and COX2), tumor necrosis factor- α (TNF- α), and inducible nitric oxide synthase (iNOS) in mice.¹²¹ Furthermore, Jamun leaf essential oils reduced chronic granulomatous inflammation in BALB/c mice induced by an intravenous injection of *Mycobacterium bovis* and *Bacillo calmet* Guerin.¹⁴⁸ Importantly, hepatitis B vaccine antigen-induced inflammation was arrested in human neutrophils, lymphocytes, and monocytes by the Jamun leaf flavonoid fraction.¹⁴⁹ Likewise, Jamun root aqueous and ethanol extracts depleted interleukin 6 (IL6) production in RAW 264.7 macrophages, indicating their anti-inflammatory activity.¹³⁴

The inoculation of *C. albicans* into Wistar rats increased the inflammatory response by increasing the expression of IL1, IL6, TNF- α , and interferon gamma (IFN- γ) and decreasing the expression of IL10, an anti-inflammatory cytokine. The treatment of rats with aqueous Jamun seed extract and polymeric nanoparticles formulated from the seed extract administered once daily for 21 subsequent days significantly attenuated the inflammatory cytokines TNF- α , interferon gamma, dipeptidyl peptidase IV, IL1, and IL6 and elevated the anti-inflammatory cytokine IL10. The inoculation of *C. albicans* in rats led to an increase in the hydrolysis of adenosine diphosphate (ADP), adenosine triphosphate (ATP), and adenosine monophosphate (AMP) and raised the ADA activity in serum and platelets; whereas the administration of the aqueous

Jamun seed extract and polymeric nanoparticles prepared from the seed extract for 21 days significantly depleted ATP, ADP, and AMP and ADA activity.¹³⁵ The methanol extract of Jamun leaves suppressed the carrageenan-induced paw edema in the hind legs of rabbits in a dose-dependent manner.³¹ The methanol extract of Jamun fruits inhibited the formaldehyde, carrageenan, and prostaglandin E2-induced paw edema in mice as well as *in vitro*, whereas 50% methanol and dichloromethane extract showed weak anti-inflammatory activity. Similarly, the Jamun fruit methanol extract attenuated glutamate and formaldehyde-induced paw licking in Swiss albino mice.⁷⁶ The aqueous Jamun leaf extract inhibited carrageenan-induced paw edema in mice.¹⁵⁰

Anti-allergic activity

The hypersensitive response elicited by the immune system against certain substances leads to allergic reactions, and different extracts of Jamun have been found to act against allergic reactions. The aqueous leaf extract alleviated mast-cell degranulator C48/80 or OVA-induced anaphylaxis edema in mice and also hindered the accumulation of eosinophils in the pleural cavity, indicating its anti-allergic potential. The rats given 1 μ g/mL Jamun leaf extract before C48/80 treatment attenuated histamine delivery into the peritoneal mast cells as a result of attrition of the allergic reaction.¹⁵¹ The aqueous methanol and methanol fractions of the aqueous extract of Jamun roots suppressed the catalepsy induced by clonidine in mice by preventing the histamine release triggered by mast cell degranulation. The milk-induced eosinophilia was also alleviated in mice by the above extracts.¹⁵²

Hepatoprotective activity

Treatment of albino rats with aqueous leaf extract of Jamun for 7 days before carbon tetrachloride (CCl₄) treatment reduced aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels, indicating its hepatoprotective activity.¹⁵³ In addition, the anthocyanin-rich ethanol extract and Jamun fruit peel aqueous extract have been reported to protect cultured rat hepatocytes against CCl₄-induced toxicity and elevate reduced glutathione (GSH) and glutathione peroxidase (GPx) levels accompanied by a subsequent decline in lipid peroxidation and lactate dehydrogenase, in which the former was more effective than the latter.¹⁵⁴ The ethanol extract of Jamun fruit pulp, when administered for eight consecutive days before paracetamol treatment, attenuated AST, ALT, alkaline phosphatase (ALP), total bilirubin, total proteins, and albumin levels in the rat liver. It also protected the rat liver against paracetamol-induced pathological changes.¹⁵⁵ The methanol Jamun seed extract protected rats against the CCl₄-induced hepatotoxicity by alleviating the acid phosphatase (ACP), ALP, AST, and ALT activities and reduced bilirubin dose dependently.¹⁵⁶ The methanol Jamun fruit extract reduced damage to hepatocytes, decreased hepatic fibrosis and macrophage infiltration induced after bile duct ligation in C57BL/6 male mice, and also decreased lipid peroxidation. The extract attenuated mRNA expression of intercellular adhesion molecule 1 (ICAM1) and chemokine (C-X-C motif) ligand (CXCL2) genes. The methanol extract also reduced IL6, IL1 β , iNOS, and nuclear factor kappa B (NF- κ B) transcriptional activation in the mouse liver.¹⁵⁷ The ethanol extract of Jamun leaves protects rat liver against CCl₄-induced hepatotoxicity by lowering AST, ALT, ALP, and bilirubin and gradually bringing them to normal levels.¹¹⁷

The administration of aqueous seed extract protected the liver of alloxan-induced diabetic mice by lowering the AST, ALT, and

bilirubin levels and histological damage.¹⁵⁸ The administration of 100 or 200 mg/kg body weight ethanol extract of Jamun seeds depleted serum gamma-glutamyl transferase (GGT) and AST in rats. Similarly, the water extract of Jamun seeds protected liver damage by decreasing AST, ALT, ALP, and GGT activities in streptozotocin-induced diabetic rats and also restored the liver histology to normal.¹⁵⁹ The rats administered with the methanol Jamun seed extract for 14 days before CCl₄ administration protected the rat liver by decreasing the AST, ALT, and ALP activities and reducing histological damage in the liver.¹⁶⁰ The methanol extract of Jamun leaves protected against CCl₄-induced hepatotoxicity in Sprague-Dawley rats by decreasing AST, ALT, total bilirubin, total cholesterol, and total triglycerides. The extract increased the GSH concentration and superoxide dismutase (SOD) activity and decreased lipid peroxidation in the Sprague-Dawley rat liver.¹⁶¹ The 90% ethanol Jamun seed extract protected against cypermethrin-induced liver hypertrophy, hyperchromatic nuclei, foamy cytoplasm with a few vacuoles, and mitochondrial swelling in Wistar rats. The extract also protected against the dilated sinusoids filled with glycogen at a few places, degeneration of hepatocytes, focal necrosis, and formation of binucleated cells in the livers of cypermethrin-treated Wistar rats.¹⁶² The methanol, ethanol, and aqueous seed extracts of Jamun protected against arsenic-induced hepatotoxicity by reducing AST, ALT, and ALP levels in the rat serum.¹⁶³ Aqueous fruit extract of Jamun protected the mice liver against chromium-induced steatosis, fibrosis, dehydration, and atrophy.¹⁶⁴

Gastroprotective activity

Different parts of Jamun have been found to relieve gastric disorders. The tannins obtained from Jamun stem bark suppressed ethanol and HCl-induced gastric ulcers in Sprague-Dawley rats by reducing gastric mucosal damage, free radicals, and ulceration of the gastric mucosa.¹⁶⁵ The hydroalcoholic extract of Jamun fruit has been reported to prevent acetylcholine, serotonin, histamine, and calcium-induced rabbit ileum contraction.¹⁶⁶ In addition, the oral administration of 95% ethanol seed extract once daily for 10 days inhibited the aspirin, cold restrain (2 h), and ethanol-induced ulcer index, gastric ulcers, acid-pepsin secretion, and cell shedding and increased mucin and mucosal glycoprotein expression.¹⁶⁷ The Jamun seed ethanol extract reduced peptic ulcers and the acid-pepsin output in diabetic rats. It also reduced indomethacin and ethanol-induced peptic ulcers in streptozotocin-induced diabetic rats.^{168,169} The indomethacin-induced gastric ulcers in mice were suppressed by the aqueous leaf extract of Jamun, which also attenuated TNF- α , NF- κ B, iNOS, COX1, and COX2 expression. The aqueous leaf extract of Jamun elevated GPx activity and reduced oxidized glutathione and NO levels in the gastric mucosa, whereas lipid peroxidation decreased in both the gastric mucosa and serum.¹²¹

Cardioprotective activity

Cardiac ailments rank number one in terms of human mortality, and Jamun has been found to protect against cardiac diseases in various preclinical studies. The oral administration of 500 mg/kg methanol extract of Jamun seeds daily for 30 days protected against isoproterenol-induced myocardial damage in rats.¹⁷⁰ In addition, the daily administration of ethanol extract of Jamun seed powder 1 h before doxorubicin treatment for 15 days protected Wistar rats against the cardiotoxic effect of doxorubicin. Moreover, the ethanol seed extract decreased the activity of creatinine

phosphokinase, ALT, AST, and lactate dehydrogenase in the rat heart.¹⁷¹ Furthermore, H9C2 cardio myoblast cells were protected against glucose-induced stress by the methanol, ethanol, and aqueous seed extracts of Jamun.^{172,173}

The hydroalcoholic extract of Jamun fruit pulp protected rats against isoproterenol-induced cardiotoxicity by decreasing the serum levels of AST, creatine kinase-myocardial band (CK-MB), cardiac troponin I, TNF- α , IL6, and C-reactive protein as well as increasing the SOD and GSH levels. The extract also increased B-cell lymphoma 2 (Bcl2) and reduced Bcl2-associated X (Bax) in the rat heart tissue, along with decreased myonecrosis, contraction of myofibrils, vacuolization, and inflammation in myocytes.¹⁷⁴ In addition, the administration of 0.5 g of hydroalcoholic extract of Jamun leaves daily orally for 8 weeks to hypertensive rats decreased hypertension.¹⁷⁵ Meanwhile, the orogastric administration of 90% ethanol Jamun fruit extract reduced spontaneous hypertension in rats.¹⁷⁶ Jamun seed powder also reduced blood pressure in a randomized double-blind placebo control clinical trial when given orally before meals twice daily for 90 days.¹⁷⁷ The freeze-dried fruit extract of Jamun induced hypotension, bradycardia, and vasorelaxation in Wistar rats by activation of potassium channels.¹⁷⁸ Recently, methanol Jamun seed extract was found to reduce glucose-induced (25 mM) cardiomyopathy in cultured H9C2 cells by inhibiting gelatinase activity. The Jamun extract suppressed the expression of gelatinase B mRNA, TNF- α , IL6, and translocation of NF- κ B.¹⁷⁹

Renoprotective activity

The different extracts of Jamun have been found to reduce kidney toxicity induced by various agents. The ethanol extract of fruits and aqueous extract of seeds of Jamun reduced urea nitrogen, urinary protein, serum creatinine, serum total proteins, and lipid peroxidation in cisplatin-treated rats. These extracts also attenuated paracetamol-induced ALT, AST, ACP, ALP, urea, and creatinine levels. The administration of albino rats with ethanol extract of fruits and aqueous extract of seeds of Jamun restored the catalase and GPx activities as well as reduced histological damage in the kidneys.^{180–182}

The ethanol stem bark of Jamun protected the kidneys against calcium oxalate stone-induced urolithiasis.¹⁸³ Sprague Dawley rats treated with betulinic acid, which is found in Jamun leaves, protected kidneys against anti-Fx1A antiserum-induced passive Heymann nephritis. Betulinic acid also decreased renal dysfunction, histopathological alterations, and immune complex deposition in the rat kidneys. The protection of the kidney by betulinic acid is mediated at the molecular level by the suppressed expression of NF- κ B, iNOS, and TNF- α and upregulation of nuclear factor-erythroid factor 2-related factor 2 (Nrf2), heme oxygenase 1, and NAD[P]H quinone oxidoreductase 1 at the mRNA level.¹⁸⁴

Antidiabetic activity

The number of diabetic patients has reached 537 million globally, and it is the world's ninth-leading disease causing death.¹⁸⁵ Diabetes results in numerous health complications, and India stands second to China in the occurrence of diabetes.¹⁸⁶ Ayurveda gives a detailed account to reduce high blood sugar levels in diabetes with Jamun seed powder, even though diabetes was not prevalent in ancient times in India. Western countries have conducted more than 130 clinical trials to control blood sugar levels in diabetes using Jamun, but the results have been inconclusive due to mixed respons-

es of patients against Jamun therapy. In these trials, many patients responded well to the treatment, but others did not.^{20,24,100,186,187}

Preclinical studies

There are a couple of reports in which Jamun was found to be ineffective in treating diabetes in preclinical animal models. Alloxan-induced diabetic rats fed with Jamun seed extract did not show a reduction in blood sugar levels.¹⁸⁸ Similarly, lyophilized Jamun fruit pulp extract administered to streptozotocin-induced diabetes rats did not show any decline in their raised blood sugar levels in a Brazilian study.¹⁸⁹ However, studies reporting that Jamun reduces blood sugar levels in preclinical and clinical conditions outnumber the studies that found Jamun to be ineffective in controlling diabetes.^{24,100,186} The rabbits administered with aqueous Jamun seed extract did exhibit a lowering of the blood glucose level.^{190,191} In addition, the blood glucose levels declined significantly in the spontaneous diabetic rats fed with Jamun ethanol stem bark extract.^{192,193} Moreover, Jamun seed powder decreased the blood glucose levels after 3 h in rabbits orally fed with 1, 2, 4, and 6 g/kg body weight aqueous suspension, and the oral administration of Jamun seed powder at 4 g/kg was found to be the most effective dose. A 42.64% alleviation in the blood glucose level was reported in rats orally given an aqueous Jamun seed powder suspension at 4 g/kg.¹⁹⁴ The alloxan-induced diabetic rats fed with 2.5 and 5 g/kg body weight Jamun seed aqueous extract daily for 6 weeks showed a decline in the blood glucose level.¹⁹⁵ Similarly, the blood glucose levels were reduced in the diabetic rats fed with Jamun fruit pulp within 30 min, whereas the hypoglycemic effect of the seed extract was evident by 24 h.¹⁹⁶

The ethanol and aqueous extracts of Jamun seeds depleted blood glucose and increased insulin levels in the rats fed with a high-fructose diet.¹⁹⁷ A decline in the serum blood glucose levels was detected in streptozotocin-induced diabetic mice administered with aqueous Jamun seed extract.¹⁹⁸ Feeding of 15% unextracted, 15% defatted Jamun seed extract, and 6% water-soluble gummy fiber diets to alloxan-induced diabetic rats for 21 days effectively reduced the blood glucose level and increased the glucose tolerance.¹⁹⁹ The streptozotocin-induced Wistar rats fed with 250, 500, and 1,000 mg/kg of Jamun seed powder for 15 days decreased the fasting blood glucose level, especially at 500 and 1,000 mg/kg.²⁰⁰ Attrition in the fasting blood glucose level has been reported in alloxan-induced mild and severe diabetic rabbits given 100 mg/kg ethanol extract of Jamun seeds/kernel daily for 15 days.²⁰¹ Streptozotocin-induced diabetic rats administered with 100 mg/kg Jamun seed kernel ethanol extract for 30 days exhibited a significant reduction in the blood glucose, urea, AST, ALT, and cholesterol levels. It also brought the SOD, catalase, GPx, and GSH levels to normal and decreased lipid peroxidation in the kidney and liver of streptozotocin-induced diabetic rats.²⁰²⁻²⁰⁴ Sprague Dawley rats given 100 and 200 mg/kg of ethanol Jamun seed extract once daily for 8 weeks reduced blood sugar level and glycated hemoglobin at 4 and 8 weeks significantly.²⁰⁵ The administration of 200 or 400 mg/kg ethyl acetate and methanol extracts of Jamun seeds or 50 mg/kg mycaminose (an isolated compound from seeds) for 15 days lowered the blood glucose levels in the streptozotocin-induced diabetic rats.²⁰⁶

A decline in serum blood sugar has been reported in streptozotocin-induced type-2 diabetic Long Evans rats orally given 1.25 g/kg Jamun seed powder and ethanol extract for 21 days.²⁰⁷ A decline in the serum glucose levels also was found in the alloxan-induced mild and severe diabetes rats fed for 21 days with 10 mg/kg body weight active principles isolated from the ethanol fraction of

seed extract of Jamun.^{208,209} The oral administration of 100, 200, and 400 mg/kg aqueous seed extract of Jamun for 21 days lowered the blood glucose level to near normal in streptozotocin-induced type-II diabetic rats and raised peroxisome proliferator-activated receptor (PPAR) alpha and gamma gene expression in the streptozotocin-induced type-II diabetic rat liver.²¹⁰ The oral administration of 50, 100, and 200 mg/kg body weight of aqueous and ethanol extracts of Jamun fruit pulp or 25 mg of partially purified water extract to alloxan-induced diabetic rabbits lowered the blood glucose levels, and the aqueous extract was more effective than the ethanol extract.²¹¹ The aqueous extract of the Jamun fruit pulp administered at a dose of 200 mg/kg daily for 15 days depleted the glucose level in the serum of streptozotocin-induced diabetes female Wistar rats, and its combination with the stem bark extract of *Cinnamom zeylanicum* in the ratio of 1:1 was more effective than either treatment alone.²¹²

Feeding 50 or 100 mg/kg aqueous and methanol extracts of Jamun leaves, stem bark, root, and seeds for 21 days to alloxan-induced diabetic male Sprague Dawley rats decreased the serum glucose level. The maximum decline in serum glucose was detected for the leaf extract, which also restored the normal architecture of the islets of Langerhans.²¹³ Intraperitoneal administration of 150 and 250 mg/kg body weight of aqueous Jamun seed extract in alloxan-induced diabetic mice for 21 days significantly depleted the serum glucose levels.²¹⁴ The methanol extract of Jamun fruit pulp, seed, and kernel from different locations of Gujarat at a concentration of 0.1–300 µg inhibited porcine pancreatic α -amylase activity *in vitro*, and maximum inhibition of α -amylase activity was recorded for the kernel extract *in vitro*.²¹⁵ Sprague Dawley rats fed with a diet containing 3% aqueous ethanol extract (50%) of Jamun fruits and seeds for 60 days showed a decrease in the serum glucose levels and an increase in the insulin levels.²¹⁶ The alloxan-induced Wistar albino diabetic rats fed with different doses of methanol extract of Jamun seeds for 14 days reduced the serum glucose level significantly on days 9, 11, and 14.²¹⁷

The streptozotocin-induced diabetic Wistar albino rats fed with 100, 200, and 400 mg/kg/day aqueous Jamun seed extract once daily for 21 consecutive days showed a decline in the serum insulin, glucose, Homeostatic Model Assessment of Insulin Resistance (HOMA-IR), and TNF- α levels ensued by a rise in PPAR γ and PPAR α expression in the liver. The aqueous seed extract also elevated catalase, SOD, and GPx activities and reduced streptozotocin-induced pancreatic β -cell damage and lipid peroxidation in the rat pancreas.²¹⁸ Feeding of 100 mg/kg body weight Jamun fruit extract rich in triterpenoids once daily for ten days to streptozotocin-induced diabetic C57BL/6 mice resulted in a decline in the fasting serum glucose as well as damage to the pancreas and increased insulin levels and HOMA- β index. The triterpenoid-enriched Jamun fruit extract-treated diabetic mouse liver showed increased phosphorylation of serine/threonine kinase (AKT) and p-AKT/AKT and decreased Forkhead box protein 1, PPAR γ coactivator 1-alpha, glucose-6-phosphatase, and phosphoenolpyruvate carboxykinase.²¹⁹ Streptozotocin-induced diabetic rats administered with 200 and 400 mg/kg of aqueous Jamun root extract or its isolated compound 4-(2-amino-2-(2-(2-hydroxy-3 methyl butyl) octahydropyrrolo[1,2-a] pyrazin-7-yl) ethyl)-2-ethylphenol daily for 15 days exhibited a decline in the serum glucose level.³² The examination of *in-vitro* α -amylase and α -glucosidase activities against the *n*-hexane, ethyl acetate, and ethanol extracts of Jamun pulp and seed, respectively, showed IC₅₀ values of 75.85 mg/mL and 74.72 mg/mL (for α -amylase activity) and 55.79 mg/mL and 59.85 mg/mL (for α -glucosidase activity), indicating the antidia-

betic action of Jamun.²²⁰ Feeding of streptozotocin-induced diabetic rats with 5 and 10% Jamun fruit in diet for 2 months reduced fasting blood glucose levels at 15, 30, 45 and 60 days.²²¹ The aqueous extract of Jamun fruit exerted antidiabetic action by inhibiting α -amylase and α -glucosidase. The extract also inhibited lipase and increased SOD activity.²²²

Clinical studies

The serum ADA activity and glucose levels declined in diabetic patients treated with aqueous Jamun leaf extract.²²³ In another study, attrition in ADA, 5'-nucleotidase, glucose, triglycerides, and lipid peroxidation levels followed by a rise in the catalase activity was observed in diabetic patients given 80% ethanol leaf extract.²²⁴ In addition, human diabetic patients fed with Jamun seed powder for 30 days had reduced fasting and postprandial blood glucose levels in their serum.²²⁵ A decline in serum glucose was found in 99 diabetic patients given 5 g of seed powder before meals for 90 days twice daily in a randomized double-blind placebo-controlled clinical trial.²²⁶ Similarly, 2 g of roasted Jamun seed powder given for 60 days to type-II diabetic patients caused a decline in their serum glucose level.²²⁷ A similar effect was observed in another double-blind clinical study carried out between March 2018 and March 2019.¹⁷⁷ The feeding of 6 g of Jamun seeds to prediabetic individuals of both sexes reduced fasting blood glucose levels.²²⁸ The preclinical and clinical studies described above indicate that Jamun is an effective treatment to control diabetes.

Antihyperlipidemic activity

Diabetes is usually linked with hyperlipidemia and is the major cause of cardiovascular morbidity, and different parts of Jamun are useful in bringing down the serum cholesterol levels. The ethanol extract of Jamun seed has been reported to decrease the total serum cholesterol/high-density lipoprotein cholesterol (HDL-c) ratio, serum low-density lipoprotein cholesterol (LDL-c), and 3-hydroxy-3 methyl glutaryl CoA (HMG-CoA) reductase activity in alloxan-induced diabetic rabbits.²⁰¹ The streptozotocin-induced diabetic rats administered with Jamun seed kernel ethanol extract exhibited a decline in the LDL-c and very-low-density lipoprotein cholesterol followed by an increase in HDL-c levels.²⁰⁸ A decrease in serum triglycerides and total cholesterol as well as increased HDL-c levels has been detected in streptozotocin-induced diabetic rats intraperitoneally administered with 200 mg/kg aqueous Jamun fruit pulp extract.²¹²

Attrition in the triglycerides and total cholesterol as well as a rise in the HDL-c levels was observed in alloxan-induced diabetic rabbits given 10 mg/kg of active compounds separated from the ethanol seed extract fraction of Jamun for 21 days.²⁰⁹ Alloxan-induced diabetic Swiss albino mice administered with 150 and 200 mg/kg body weight of Jamun seed aqueous extract reduced hyperlipidemia by restoring triglycerides and total cholesterol to normal and increased the HDL-c level.²¹⁴ A depletion in triglycerides and LDL-c followed by an increase in HDL-c was observed in rats fed with a high-cholesterol diet for 30 days and administered with 3% ethanol extract of seeds and fruits of Jamun for 60 days continuously.²²⁹ The streptozotocin-induced diabetic C57BL/6 mice treated with 100 mg/kg body weight of Jamun fruit extract enriched with triterpenoid once daily for 10 days resulted in a decline in the triglyceride and free fatty acid levels in the serum and liver of diabetic mice accompanied by a decline in the mRNA levels of acetyl-CoA carboxylase 1 (ACC1), cluster of differentiation 36 (CD36), stearoyl-CoA desaturase-1 (Scd1), and fatty acid synthase

as well as ACC1 expression at the protein level. Jamun fruit extract raised p-ACC1 protein expression and normalized extracellular signal-regulated kinase (Erk1/2) and p-Erk1/2 expression in diabetic mice.²¹⁹

The diabetic patients given 2 g of Jamun seed powder daily for 60 days showed a significant depletion in serum cholesterol, very-low-density lipoprotein, LDL, cholesterol, and triglycerides.²²⁷ Hyperglycemic patients treated with 4.5 g of encapsulated Jamun seed powder once daily for 84 days showed a significant decline in the cholesterol and LDL-c levels.²³⁰

Anti-obesity activity

Obesity (body mass index > 30 kg/m²) is one of the major problems in the world today, and there has been a 39% increase in the obese population throughout the world between 1975 and 2020; approximately 764 million individuals were obese in 2020, and the number is constantly on the rise and will reach to more than 1 billion by 2030 (<https://www.worldobesity.org/about/about-obesity/prevalence-of-obesity>). The various parts of Jamun have been found to reduce obesity in several preclinical studies. The high-carbohydrate and high-fat diet-fed obese Wistar rats given 2.5% Jamun seed powder for 56 days showed a significant decrease in body weight gain, accumulation of white adipose tissue, blood glucose, serum insulin, total cholesterol, LDL-c, triglycerides, and lipid peroxidation in the plasma. It normalized the AST, ALT, and ALP levels in obese rats. In addition, the feeding of Jamun seed powder restored the levels of HDL-c, GSH, SOD, and catalase in the plasma of obese rats, followed by a decline in the advanced protein oxidation products, myeloperoxidase, NO generation, lipid peroxidation, and elevated catalase, GSH, and SOD levels in the obese rat liver.²³¹

The C57BL/6 obese mice fed with both ethanol and acetone Jamun fruit extracts returned the ratio of *Firmicutes* to *Bacteroidetes* in the gut to normal as well as inhibited body weight gain and white adipose tissue accumulation in epididymal, visceral, and subcutaneous tissues. The extracts also decreased the serum glucose and insulin levels in obese mice. The oral feeding of both ethanol and acetone Jamun fruit extracts led to normalization of the expression of p-AKT and phosphoinositide 3-kinase (p85) protein and a reduction in the phosphorylation of insulin receptor substrate 1 (p-IRS1) at Ser318, mRNA expression of ACC1, sterol regulatory element-binding protein 1c, FAS, PPAR γ , and CD36 in the liver of obese mice. A decline in cholesterol, triglycerides, and free fatty acid levels was found in the liver and plasma of obese mice fed with ethanol and acetone Jamun fruit extracts; however, the plasma cholesterol level remained unaltered.²³²

Oral feeding of 0.5 or 1.0 g/kg/day of hydroalcoholic Jamun leaf extract daily for 8 weeks decreased the weight gain and white adipose tissue storage in the retroperitoneal and periepididymal regions of monosodium L-glutamate-induced obesity in newborn Wistar rats. Attrition in the serum glucose and free fatty acid levels and restoration of the total cholesterol and triglyceride levels to normal was evident in obese rats fed with the Jamun leaf extract. A regression in nonalcoholic fatty liver disease and a decline in AST activity were detected in the obese rats treated with hydroalcoholic Jamun leaf extract. The livers of obese rats fed with hydroalcoholic extract suppressed endoplasmic reticulum stress by arresting the G protein-coupled receptor 98 (GPR98) expression, translation of the endoplasmic reticulum protein retention receptor (KDEL) chaperone, and GPR78 expression marginally. The Jamun leaf hydroalcoholic extract also attenuated spliced microsomal triglyc-

eride transfer protein (MTP), X-box-binding protein (XBP) 1s, unspliced XBP1u, and protein disulfide isomerase (PDI) expression in the hepatocytes of the obese rats.²³³

Antipyretic activity

The antipyretic potential of Jamun also has been explored, with the chloroform fraction from the methanol seed extract and the ethanol extract of Jamun leaves reducing brewer's yeast-induced pyrexia in rats and bringing the body temperature to normal.^{140,234,235}

Antidiarrheal activity

Diarrhea is a common occurrence, and everyone suffers from this disorder at some time in their life. Whatever the cause, diarrhea leads to discomfort to the individual suffering from it. Jamun has been used to control diarrhea for a long time. The oral administration of 125, 250, and 500 mg/kg body weight Jamun aqueous seed extract, given 30 min before castor oil treatment, reduced castor oil-induced diarrhea in mice.²³⁶ In addition, the stem bark of Jamun extracted in ethanol controlled the castor oil-induced diarrhea in mice and rats.^{237,238} Inhibition of castor oil-induced diarrhea has been observed in mice treated with the ethanol, chloroform, and petroleum ether fractions from the crude methanol extracts of Jamun leaves.^{239,240}

Anti-Leishmania activity

Leishmaniasis is a tropical disease, and Kala-azar is the most serious form caused by the protozoan *Leishmania*, which is transmitted in humans by an infected female phlebotomine sandfly bite. Exposure of promastigotes of *Leishmania amazonensis* to the essential oil of Jamun for 24, 48, and 72 h led to their killing, and the 24-h exposure time was most effective, with an IC₅₀ of 36 mg/L.²⁴¹ The essential oil and its constituent α -pinene from the Jamun leaves were effective against *L. amazonensis* promastigotes, with an IC₅₀ of 19.7 μ g/mL. α -Pinene was more effective against axenic and intracellular amastigotes, with IC₅₀ values of 16.1 μ g/mL and 15.6 μ g/mL, respectively, whereas the essential oil had IC₅₀ values of 43.9 μ g/mL and 38.1 μ g/mL against axenic and intracellular amastigotes, respectively. Increased phagocytic, lysosomal activity, and NO levels are responsible for this effect.²⁴²

Antispasmodic activity

Treatment of rat uterine smooth muscles with 936.55 μ g/mL Jamun seed ethanol extract reduced potassium chloride-induced muscle contraction *in vitro*.²⁴³ The exposure of rat uterus and jejunum to 27, 81, 243 and 729 μ g/mL hydroalcoholic extract of Jamun fruit alleviated oxytocin or carbachol-induced muscle contraction. Similarly, it also reduced muscle contraction induced by potassium chloride and carbachol.⁴⁶ In addition, isolated guinea pig trachea exposed to 2.5, 5, 7.5, and 10 mg/mL macerated aqueous Jamun leaf extract showed an attenuated potassium chloride-induced contraction and caused tracheal relaxation concentration dependently.²⁴⁴ A dose-dependent inhibition has been reported in acetylcholine, histamine, serotonin, and calcium-induced contraction in isolated rabbit ileum samples exposed to aqueous and ethanol extracts of Jamun fruits, whereby 3 mg/mL and 5 mg/mL extracts were more effective than 1 mg/mL and 2 mg/mL.²⁴⁵ Hydroalcoholic (70%) ethanol extract of Jamun leaves attenuated potassium chloride, acetylcholine, and calcium chloride-induced jejunum

contraction, depending on the concentration, and this was due to a reduction in the Ca⁺⁺ level.²³⁹ Exposure of the uterus and jejunum of Wistar rats to 70% ethanol extract of Jamun seeds inhibited the oxytocin and carbachol-induced contraction dose dependently.²⁴⁶

Antiretinitis activity

The mutagenic alteration in the X-linked retinitis pigmentosa gene is linked to loss of vision in humans, and there is no treatment for this disease presently. The antiretinitis effect of five anthocyanin compounds like cyanidin 3,5 diglucoside, delphinidin 3,5-diglucoside, petunidin 3,7-diglucoside, peonidin 3,5-diglucoside, and malvidin 3,5-diglucoside from Jamun fruit peel was evaluated on the X-linked retinitis pigmentosa gene by molecular docking studies, and cyanidin 3,5 diglucoside was found to be the most effective inhibitor of retinitis pigmentosa in humans compared to the other anthocyanin compounds.²⁴⁷

Anti-Alzheimer's disease (AD) activity

The methanol extract of Jamun fruits has been reported to protect against scopolamine-induced amnesia by reducing the activity of acetylcholine esterase and lipid peroxidation accompanied by an elevation in the activities of catalase and SOD in the rat brain.²⁴⁸ The administration of Jamun seed methanol extract daily for 12 weeks to amyloid β 1-40 induced AD in Wistar rats significantly increased their memory and reduced amyloid plaque formation in the cortex and hippocampus of the rat brain. The Jamun extract also reduced lipid peroxidation in the cortex and hippocampus of the brain and plasma of AD rats. A decline in TNF- α and a rise in brain-derived neurotrophic factor, postsynaptic density protein 95, tropomyosin receptor kinase B, and synaptosomal-associated protein 25 have been reported in the cortex and hippocampus of AD rats treated with Jamun extract.²⁴⁹ A significant reduction in acetylcholinesterase and butyrylcholinesterase activities, hydroperoxides, and lipid peroxidation accompanied by a rise in the SOD, catalase, and glutathione-S-transferase activities and the GSH concentration were observed in the brain of alloxan-induced diabetic rats treated with polyphenols extracted from Jamun leaves and 80% acetone leaf extract of Jamun.²⁵⁰

Effect on the central nervous system

The methanol seed extract of Jamun acts as a sedative in mice, whereby it changes the overall behavior pattern and reduces the exploratory behavior pattern, spontaneous motility, hypothermia, pentobarbitone-induced hypnosis, analgesia, spontaneous locomotor activity, electric shock-induced fighting, muscle relaxant action, aggressive behavior, and exploratory behavior patterns. The methanol seed extract inhibited pentylenetetrazol and strychnine-induced convulsions and lethality. Furthermore, the Jamun seed extract decreased amphetamine-induced toxicity in mice, depending on the dose.²⁵¹ The methanol and ethyl acetate extracts of Jamun seeds reduced diazepam and chlorpromazine-induced central nervous system activity in mice.²⁵² The oral administration of Jamun seed extract for 12 weeks improved the memory and learning behavior in senile rats by decreasing lipid peroxidation in the hippocampus and cerebral cortex.²⁵³ Moreover, the aqueous extract of Jamun leaves reduced the ADA activity in the cerebral cortex of Wistar rats.²⁵⁴ The aqueous and ethanolic stem bark extracts of Jamun also protected rat pheochromocytoma PC-12 cells against 6-hydroxydopamine-induced toxicity *in vitro*.¹³⁷

Antinociceptive activity

Pain is an unpleasant perception caused by numerous stimuli and tissue damage, and everyone experiences pain due to some or the other reason. The different parts of Jamun possess antinociceptive action and relieve pain. The hydroalcoholic extract of Jamun leaves exerted antinociceptive activity in a dose-dependent manner, as indicated by hot plate and formalin tests in rats, and also suppressed the release of excitatory amino acids, including prostaglandin E₂, kinins, glutamate, protons, and NO.²⁵⁵ The methanol extract of Jamun seed extract attenuated acetic acid-induced writhing in mice dose dependently.²⁵⁶ In addition, the methanol extract of Jamun roots exerted analgesic action in the rats in the tail immersion test.²⁵⁷ The petroleum ether, ethanol and chloroform root bark extracts of Jamun also exhibited a dose-dependent antinociceptive activity in mice as indicated by alleviation of acetic acid-induced writhing and formalin-induced pain.²⁵⁸ Moreover, the administration of 50 and 100 mg/kg body weight methanol extract of Jamun leaves exerted analgesic action in rabbits.³¹ The petroleum ether and chloroform fractions of the methanol extract of Jamun leaf exerted an analgesic effect in mice in a dose-dependent manner.²⁴⁰ Furthermore, the methanol extract of Jamun fruit pulp exerted antinociceptive activity in a dose-dependent manner in mice against the formalin and glutamate-induced paw licking test, and the maximum effect was observed at 200 mg/kg Jamun extract.⁷⁷

Anti-arthritic activity

Arthritis is tenderness and swelling of joints causing pain that increases with age. Its incidence has been increasing globally. Arthritis is categorized as osteoarthritis and rheumatoid arthritis, which is an autoimmune disorder. Jamun has been reported to be active against rheumatoid arthritis in preclinical models. The methanol seed extract of Jamun helped to attenuate Freund's complete adjuvant-induced arthritis in rats administered with 250 mg/kg and 500 mg/kg of the extract.²⁵⁹ The oral administration of 50, 500, 750, and 1,000 mg/kg of petroleum ether extract of Jamun stem bark also attenuated Freund's complete adjuvant-induced arthritis in rats. The extract reduced the expression of rheumatoid factor and the arthritic index as well as increased the body weight dose dependently.²⁶⁰

Diuretic activity

The oral administration of chloroform, methanol, and aqueous extracts of stem bark of Jamun increased the urine output in Wistar albino rats, with the methanol extract being the most effective among all three extracts, whereas the chloroform extract was the least active.²⁶¹ The oral administration of the methanol seed extract of Jamun elevated the urine volume and the excretion of chlorine, sodium, and potassium electrolytes at 24 h compared to 5 h in Wistar rats.²⁶²

Anthelmintic activity

The exposure of *Pheretima posthuma* to 50–100 mg/mL benzene, methanol, chloroform, and *n*-hexane extracts of Jamun leaves killed the worms in a dose- and time-dependent manner, and the benzene extract was the least effective. The anthelmintic activity of the methanol, chloroform, and *n*-hexane extracts was better than the standard albendazole.²⁶³ The aqueous and methanol extracts of the stem bark of Jamun paralyzed *P. posthuma* within 36.58 min

after exposure to these extracts, and 100 mg/mL methanol extract killed all worms within 70.58 min after treatment.^{264,265} The exposure of *P. posthuma* to 50–100 mg/mL 95% ethanol seed extract of Jamun paralyzed and killed worms dose and time dependently, and initiation of the effect was early for 100 mg/mL.²⁶⁶ The oral feeding of Jamun leaf powder to sheep led to a 76.67% reduction in the eggs/g of the intestinal worm *Haemonchus contortus*, indicating its anthelmintic activity *in vivo*.²⁶⁷ The treatment of *Lumbriacus terrestris* (earthworms), *Monnizia expansa*, (tapeworms), and *H. contortus* (red worms) with 6.25, 12.5, 25, 50, and 100 mg/mL methanol extract of Jamun leaves paralyzed and killed all the worms in a concentration and time-dependent manner. The higher the concentration, the less time was needed to paralyze and kill the worms with the methanol leaf extract.²⁶⁸

Immunomodulatory activity

The oral administration of the methanol seed extract of Jamun demonstrated immunomodulatory activity in mice and rats orally administered at doses of 100, 200, 300, 400, and 500 mg/kg. The methanol seed extract enhanced the carbon clearance and hemagglutination titer in mice and the delayed-type hypersensitivity reaction in rats in a dose-dependent manner. The seed extract elevated the white blood cell and lymphocyte counts significantly.²⁶⁹

Wound healing activity

Wounds are a common occurrence, and most individuals suffer from this type of injury during their life. Jamun has been found to heal wound injuries. The oral administration and topical application of the ethanol seed extract of Jamun healed punch wounds in streptozotocin-induced diabetic Sprague Dawley rats.²⁷⁰ The topical application of Jamun honey on excision wounds healed wounds of normal and streptozotocin-induced diabetic rats by increasing re-epithelialization and collagen deposition as well as the expression of collagen I, III, hypoxia-inducible factor 1 α , vascular endothelial growth factor (VEGF), and VEGF receptor II. Jamun honey also healed scratch wounds created on cultured primary fibroblasts of both normal and diabetic rats as well as increased the expression of collagen type I and II and alpha-smooth muscle actin.²⁷¹

Metal toxicity protective activity

Metals like mercury and arsenic are present in the environment, and they cause health hazards in humans. The leaf and seed extracts of Jamun have been applied to reduce metal toxicity. The aqueous Jamun seed extract protected two-day-old neonate rats against mercury toxicity by lowering the methyl mercury-induced *N*-acetyl- β -D-glucosaminidase activity in the kidney and urine, lipid peroxidation levels in the liver, and ADA activity in the hippocampus, kidney, and liver.²⁷² The treatment of mice with the ethanol extract of Jamun leaves for 12 weeks protected mice against the arsenic-induced decline in the body weight and the organ-to-body weight ratio of the spleen, kidney, and liver. It also reduced the arsenic-induced rise in the ALT, ALP, lactate dehydrogenase, uric acid, and glucose levels in the mouse serum.²⁷³ The oral administration of methanol, ethanol, and aqueous seed extracts of Jamun seeds for 60 days protected Wistar rats against arsenic-induced DNA damage and increased the serum AST, ALT, and ALP levels.¹⁶³ Treatment of rats with 70 ethanol extract of Jamun bark

reduced the arsenic induced rise in the glucose, albumin, blood urea, nitrogen, creatinine and total protein in plasma and AST and ALT in the serum.²⁷⁴

Hair growth stimulation

The topical application of Jamun fruit pulp and seed ethanol extract ointment induced hair growth in Swiss albino mice earlier than the control. The length and number of hairs were increased, and the time of appearance of hairs was also advanced in mice significantly. The Jamun fruit pulp extract was superior to the seed extract in accelerating hair growth in mice.²⁷⁵

Antimutagenic and antigenotoxic activities

Aqueous and ethanolic extracts of Jamun seeds protected pBR322 DNA against hydroxyl radical-induced strand breaks. Swiss albino mice administered with 1 or 1.5 g/kg of Jamun seed extract significantly protected mouse bone marrow cells against urethane and 7,12-dimethylbenz(a)anthracene (DMBA)-induced micronuclei formation and chromosomal aberrations. The seed extract also elevated the GSH concentration and the catalase, SOD, and glutathione-S-transferase activities as well as reduced lipid peroxidation.²⁷⁶ The ethyl acetate Jamun seed extract (100 µg/mL) attenuated the sodium azide and methyl methanesulfonate-induced increase in the revertant's frequency in *S. typhimurium* tester strains. The extract also reduced methyl methanesulfonate-mediated DNA fragmentation and oxidation in lymphocytes as well as oxidative damage in pBR322 plasmid DNA.⁸⁵ The 70% ethanol seed extract of Jamun also showed an antimutagenic effect against *S. typhimurium* (TA 98 and TA 100) strains.¹¹⁴

Radioprotective activity

Cosmic radiation, air and space travel, background radiation, radiodiagnosis, and/or radiotherapy procedures are the main sources of human radiation exposure in their daily life. Ionizing radiation inflicts various types of deleterious effects and is known to cause liver, cardiovascular, kidney, pulmonary, and reproductive disorders in addition to cancers of all organs. The deleterious effects of ionizing radiation can be reduced by pharmacological intervention. The Jamun leaf extracted in 1:1 dichloromethane/methanol protected against micronuclei formation in a concentration-dependent manner in human lymphocytes exposed to 3 Gy of γ -radiation.²⁷⁷ The administration of 5, 10, 20, 30, 40, 50, 60, and 80 mg/kg body weight of dichloromethane/methanol leaf extract protected mice against whole-body γ -irradiation-induced radiation sickness and mortality, with optimum protection at a dose of 30 mg/kg.¹⁰⁶ In addition, the hydroalcoholic Jamun seed extract protected mice against radiation-induced sickness and mortality, with optimum protection at a dose of 80 mg/kg body weight and a dose reduction factor of 1.24.¹⁰⁷

The effect of 5, 10, 20, 30, 40, 50, 60, and 80 mg/kg body weight of dichloromethane/ methanol leaf extract alleviated radiation-induced damage in the intestine of mice exposed to different doses of γ -radiation, as indicated by the increased villus height, higher number of regenerating crypts, and reduction in the number of goblet cells and dead cells.²⁷⁸ The mice administered intraperitoneally with 50 mg/kg body weight dichloromethane/methanol leaf extract had reduced micronuclei formation in an irradiation dose-dependent manner in the cytokinesis-blocked cultured splenocytes extracted from irradiated mice exposed to 0, 0.5, 1, 2, 3,

and 4 Gy whole-body γ -radiation.¹¹⁸ Ionizing radiation reduced the anti-oxidant status, and the intraperitoneal administration of 50 mg/kg body weight of dichloromethane/methanol Jamun leaf extract before exposure to 0, 0.5, 1, 2, 3, and 4 Gy whole-body γ -radiation elevated the GSH concentration and catalase and SOD activities as well as decreased radiation-induced lipid peroxidation in the mouse liver.²⁷⁹

Anticancer activity

Cancer is a noncommunicable disease, and it is the second-largest disease of the human population leading to death after cardiovascular diseases. Cancer is treated by surgery, radiotherapy, chemotherapy, or their various combinations. Chemotherapy is the only remedy to treat cancer in advanced stages; hence, it has emerged as one of the most important modalities for cancer treatment. Most chemotherapeutic drugs are of natural origin or their semisynthetic derivatives.^{12,280} Different parts of Jamun have been investigated for their cytotoxic action in different cell lines *in vitro*. The methanol crude extract of Jamun fruit skin was cytotoxic to HeLa (human papillomavirus-18 positive) and SiHa (human papillomavirus-16 positive) cells by a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay, and it was more cytotoxic to the former than the latter. The cytotoxicity of the extract was due to the induction of greater apoptosis in HeLa cells than in the SiHa cells.²⁸¹ Moreover, freeze-dried Jamun fruit pulp extract reduced the cell proliferation and growth of MCF-7 and MDA-MB-231 breast cancer cells in a concentration- and time-dependent manner, and the extract was less effective against nontumorigenic MCF-10A cells. The Jamun fruit pulp extract stimulated apoptotic cell death in both MCF-7 cells and MDA-MB-231 breast cancer cells, whereas it did not induce apoptosis in untransformed MCF-10A breast cancer cells.²⁸² Treatment of acute myeloid leukemia cells (immature monocytes) collected from the patients with 25, 50, and 100 µL of hexane, chloroform, ether, ethyl acetate, ethanol, and water extracts of Jamun fruits killed leukemia cells in a concentration-dependent manner, whereby the ethyl acetate and ethanol extracts were more cytotoxic than the other extracts.²⁸³

Treatment of MCF-7 cells with 62.5, 125, 250, 500, and 1,000 µg/mL methanol fruit pulp extract of Jamun triggered cytotoxicity, depending on the concentration, and 1,000 µg/mL pulp extract was the most cytotoxic, with an IC₅₀ of 266.8 µg/mL.²⁸⁴ The Jamun fruit acidified (0.1% HCl) methanol extract also has been reported to induce cytotoxicity in a concentration-dependent manner in HCT-116 colon cancer cells by the MTT assay and reduced colony formation in colon cancer stem cells, depending on the concentration. The Jamun fruit extract also induced apoptosis in HCT-116 and colon cancer stem cells by triggering DNA fragmentation and elevating the caspase 3 and caspase 7 activities.²⁸⁵ The methanol extract of Jamun fruits triggered cytotoxicity by decreasing cell proliferation in H460 lung cancer cells, depending on the concentration, with an IC₅₀ of 35.2 µg/mL.²⁸⁶ Treatment of A459 lung cancer cells with 50, 100, 150, 200, and 250 µg/mL Jamun pulp and seed extracted in 75% ethanol also inhibited cell proliferation in a concentration-dependent manner as determined by an MTT assay.⁷¹

Exposure of CHO cells to 10–1,000 µg/mL aqueous ethanol Jamun fruit extract induced cytotoxicity, depending on the concentration, by an MTT assay, with an IC₅₀ of 400 µg/mL.¹³² Treatment of HT29 cells with 1, 5, 10, 50, 100, 250, 500, and 1,000 µg/mL freeze-dried fruit peel extract of Jamun increased the cytotoxicity in a concentration-dependent manner and decreased the

G₂+M phase cells, especially at a concentration of 1,000 µg/mL.²⁸⁷ Non-small cell lung cancer cells (CP-H460) and human embryonic kidney cells (HEK-293) were treated with 0.01, 0.1, 1, and 2 µg/mL aqueous fruit extract, which increased the cytotoxic effect in a concentration-dependent manner, and 2 µg/mL was cytotoxic to 89% CP-H460 cells, which were more sensitive than HEK-293 cells.²⁸⁸ Treatment of HT-29 cells with 62.5, 152, 250, 500, and 1,000 µg/mL 80% methanol fruit extract increased the cytotoxic effect, depending on the concentration, with an IC₅₀ of 267.5 µg/mL, and also induced DNA damage. The fruit extract also inhibited cell migration as indicated by delayed healing of scratch wounds created on HT-29 cell cultures. The methanol fruit extract of Jamun suppressed the expression of anti-apoptotic Bcl2 mRNA and stimulated the expression of Bax mRNA in HT-29 cells.²⁸⁹

The ethyl acetate and methanol extracts of Jamun seeds reduced cell survival and increased the cytotoxicity in MCF-7 cells in a concentration-dependent manner, and the ethyl acetate extract was slightly better than the methanol extract. Almost similar results have been reported for DNA fragmentation, an indicator of apoptosis.²⁹⁰ Oral squamous cancer cells from the tongue (SCC-25) exposed to 1, 3, 10, 30, 100, and 300 µg/mL methanol seed extract of Jamun showed a concentration-dependent increase in the cytotoxicity, with an IC₅₀ of <50 µg/mL. The seed extract induced reactive oxygen species in SCC-25 cells that were detected by a dichlorodihydrofluorescein diacetate assay. The Jamun seed extract induced apoptosis as determined microscopically and by flow cytometry with a fluorescein isothiocyanate-conjugated annexin V binding assay. The seed extract increased the expression of cadherin 1 in SCC25 cells concentration dependently.²⁹¹ Additionally, the ethanol Jamun seed extract was cytotoxic in a brine shrimp assay, with an IC₅₀ of 61.50 ± 7.17 µg/mL.²⁹² The ethanol extract of Jamun seeds at concentrations of 10, 100, and 1,000 µg/mL exerted a cytotoxic effect on MCF-7, A2780 (ovarian adenocarcinoma), PC-3 (prostate carcinoma), and H460 (non-small cell lung carcinoma) cells, depending on the concentration. The A2780 cells were most sensitive, with an IC₅₀ of 49 µg/mL; whereas H460 cells were the least sensitive, with an IC₅₀ of 165 µg/mL. The MCF-7 and PC-3 cells showed an intermediate sensitivity, with IC₅₀ values of 110 µg/mL and 140 µg/mL, respectively.²⁹³

Exposure of Hep2 (human laryngeal epithelioma) cells to 8, 15.6, 31.25, 62.5, 125, 250, 500, and 1,000 µg/mL acetone, methanol, and ethanol seed extracts of Jamun resulted in a concentration-dependent rise in the cytotoxicity as studied by an MTT assay, and 50% of cell killing was detected at 125 µg/mL.²⁹⁴ Treatment of MCF-7, T-47D (breast), SF-295 (central nervous system), HCT-116 (colon), A-549 (lung), MDA-MB-435 (melanoma), OVCAR-5 (ovary), PC-3 (prostate), and A-498 (renal) cells with 100 µg/mL methanol seed extract of Jamun showed the following growth inhibition percentages: 93% (SF-295 cells), 75% (A-498 cells), 74% (HCT-116 cells), 72% (A-549 and PC-3 cells), 71% (MDA-MB-435 cells), 65% (OVCAR-5 cells), 61% (MCF-7 cells), and 60% (T-47D cells). Exposure of A459 cells to 1, 10, 30, and 50 µg/mL seed extract was cytotoxic in a concentration-dependent manner, with an IC₅₀ of 10 µg/mL.²⁹⁵ Treatment of MCF-7 cells with 5, 10, 20, 40, and 80 µg/mL aqueous extract of Jamun leaves also induced cytotoxicity in a concentration-dependent manner as well as increased the reactive oxygen species production.²⁹⁶

T47D breast cancer cells treated with 100 µg/mL hexane:50% ethyl acetate, ethyl acetate, and ethyl acetate:25% methanol extracts of Jamun leaves were cytotoxic, and the hexane:50% ethyl acetate extracts showed 68% cytotoxicity compared to the ethyl acetate extract. The ethyl acetate and ethyl acetate:25% methanol

extracts had cytotoxicity percentages of 56% and 50%, respectively.¹²⁴ The HT-29 cells treated with 20, 40, 60, 80, and 100 µg/mL ethanol extract of Jamun leaves resulted in a concentration-dependent rise in the cytotoxicity, with an IC₅₀ of 90.42 µg/mL.²⁹⁷ A concentration-dependent decline in cell proliferation was observed in HepG2, Caco2, and PC3 cells treated with different concentrations of methanol extract of ripe fruit pulp and seeds as well as unripe fruit pulp and seeds of Jamun. The IC₅₀ values for the ripe fruit pulp, ripe fruit seeds, unripe fruit pulp, and unripe fruit seeds were 27.78, 89.1, 77.33, and 75.2 µg/mL for HepG2 cells, respectively. The IC₅₀ values for Caco2 cells were 90.48, 39.48, 40.21, and 30.93 µg/mL, respectively, for ripe fruit pulp, ripe fruit seeds, unripe fruit pulp, and unripe fruit seeds. Similarly, IC₅₀ values of 50.21, 38.3, 53.71, and 43.21 µg/mL were recorded for ripe fruit pulp, ripe fruit seeds, unripe fruit pulp, and unripe fruit seeds in PC3 cells, indicating that the cells respond differentially to the different fruit extracts of Jamun.¹³⁰ The exposure of HepG2 cells to 8, 15.6, 31.25, 62.5, 125, 250, 500, and 1,000 µg/mL acetone, ethanol, and methanol extracts of Jamun seeds inhibited cell proliferation, depending on the concentration, and the methanol seed extract was most active, with 50% of cells being killed at 125 µg/mL.²⁹⁸ Treatment of MCF-7, MDA MB-231, and HCT 116 cells with 0, 0.1, 1, 10, 100, and 1,000 µg/mL Jamun leaf, fruit, seed, and flower extracted in methanol resulted in a concentration-dependent inhibition of cell proliferation. IC₅₀ values of 1.24 ± 0.09 mg/mL and 1.42 ± 0.34 mg/mL were recorded for the seed and leaf extracts, respectively, in HCT-116 cells; whereas MDA-MB-231 cells showed IC₅₀ values of 5.86 ± 0.63 mg/mL and 6.97 ± 0.68 mg/mL for leaf and flower extracts, respectively.²⁹⁹ Treatment of HT29 cells with 7.8, 15.6, 31.2, 62.5, 125, 250, 500, and 1,000 µg/mL fresh and dehydrated fruits of Jamun extracted in acetone resulted in a concentration-dependent rise in the cytotoxicity by an MTT assay, with growth inhibition percentages of 51.42% for the fresh fruit extracts (31.2 µg/mL) and 50.10% for the dehydrated fruit extracts (15.6 µg/mL). Similarly, hybrid fresh and hybrid dehydrated fruit acetone extracts resulted in 50.98% and 48.13% killing of HT29 cells at 125 µg/mL and 62.5 µg/mL, respectively. The acetone extract of dehydrated Jamun fruit was more effective than the other extracts.³⁰⁰

The anticancer activity of the Jamun stem bark methanol extract was evaluated in Ehrlich ascites carcinoma tumorized Swiss albino mice intraperitoneally administered with the extract at 25, 50, and 75 mg/kg per day. The methanol extract inhibited the tumor growth dose dependently, and 75 mg/kg extract caused maximum growth inhibition and increased the mean survival time of mice by approximately 12 days when compared to no extract-treated tumor-bearing mice. The methanol Jamun bark extract caused DNA fragmentation, induced apoptosis, increased p53 and proapoptotic Bax expression, and subsequently downregulated Bcl2 and NF-κB expression in Ehrlich ascites carcinoma cells.⁵⁴

Chemopreventive activity

The Jamun fruit pulp and seed extracts have been found to inhibit chemically induced skin and breast carcinogenesis in preclinical studies. The mice orally administered with the hydroalcoholic extract of Jamun seeds at 125 and 250 mg/kg body weight until the termination of the experiment was found to reduce the tumor incidence, the tumor burden, and the average number of tumors, followed by an increase in the latency period for tumor formation in the DMBA-treated two-stage skin carcinogenesis model. The administration of 250 mg/kg hydroalcoholic extract reduced lipid

peroxidation and elevated GSH, vitamin C, and activities of SOD and catalase in the liver and skin of the DMBA-treated mice.^{301,302} The feeding of a 5% Jamun fruit pulp powder diet to female ACI rats before 2 weeks of breast tumor induction by 17- β -estradiol delayed the appearance of breast tumors by 21 days and also reduced the tumor incidence, tumor burden, and tumor multiplicity in female rats. Jamun extract significantly reduced the progesterone levels as well as the expression of cyclin D1, estrogen receptor alpha, and proliferating cell nuclear antigen. Jamun also downregulated the mRNA levels of cyclin D1, cyclin D3, Fox1, Bcl2, and cyclin-dependent kinase 4 (Cdk4) in mammary tumors.³⁰³

Other activities

In addition to the above-listed activities, Jamun has been shown to possess anticoagulant, anti-aging, antimalarial and antiasthmatic properties.³⁰⁴⁻³⁰⁷

Adverse effects

Despite several salubrious effects, Jamun is known to cause a few adverse effects in humans because of its ability to lower blood sugar levels. Jamun fruits should not be eaten within one week before and a minimum of two weeks after surgery to avoid complications. Eating Jamun should be avoided on an empty stomach and immediately after drinking milk. Additionally, Jamun should not be eaten by breastfeeding mothers or pregnant women to avoid complications. Eating too large of a quantity of Jamun fruit causes cough, body ache, sputum accumulation in the lungs, or even fever in humans. Other side effects of Jamun are delayed digestion, flatulence, emphysema, and inflamed larynx and lungs. Common salt and *Piper nigrum* may be used as a corrective measure to minimize the side effects of Jamun.²⁰

Discussion

It is clear from the above studies that different parts of Jamun, including the root, stem, leaf, flower, fruit, and seed, show potential in the treatment of infection, inflammation, and oxidative stress-related diseases like diabetes, arthritis, neurodegeneration, cardiovascular disease, intestinal disorders, fibrosis, aging, obesity, carcinogenesis, and cancer. Jamun also protects against helminthic, bacterial, viral, fungal, and Leishmania infections, immunity-related disorders, metal-induced toxicity, kidney diseases, malaria, mutagenesis, pain, wound injury, and radiation-induced damage.¹¹³⁻³⁰³ The seeds of Jamun are clinically used to treat diabetes, allergies, inflammation, gastric ulcers, and viral infections.⁸ The medicinal properties of Jamun are due to its ability to produce various secondary metabolites that can be developed as future medicines with no adverse effects or few side effects, unlike modern synthetic drugs that have numerous side effects leading to secondary ailments.³²⁻⁸⁷ The root, stem, leaf, flowers, fruits, and seeds of Jamun have the potential as future medicines to treat numerous diseases, as indicated above, but more systematic research on each disease state is needed.

Mechanism of action

The mechanism of protection against various disorders by Jamun is not well understood. The generation of free radicals in the human body is linked to numerous diseases, including arthritis, dia-

betes, autoimmune disorders, neurodegenerative diseases, cardiovascular diseases, fibrosis, aging, and cancer. The action of Jamun depends on the conditions of the target tissues. The scavenging of free radicals by Jamun and its active phytochemicals, including myricetin, quercetin, cyanidin, imperatorin, ellagic acid, and kaempferol, seems to be one of the important mechanisms of action to exert conducive effects.^{115-122,125,128} The beneficial effects of Jamun on various organs may be due to its ability to activate Nrf2, which stimulates heme oxygenase and NAD[P]H quinone oxidoreductase 1 as well as increase the GSH, GPx, catalase, SOD, and glutathione-S-transferase levels and reduce lipid peroxidation (Fig. 4).^{154,161,171,184,235} At the molecular level, Jamun and its active biomolecules myricetin and quercetin activate PPAR α and PPAR γ , which play a crucial role in fatty acid and glucose metabolism and reduce inflammation by suppressing NF- κ B, IL6, COX1, and COX2, and subsequent events seem to protect various tissues and augment wound healing by increasing collagen type I, III, alpha-smooth muscle actin, hypoxia-inducible factor 1 α , VEGF, and VEGF receptor II (Fig. 5).^{121,158,180,185,210,219,230,271} The inhibition of ICAM1, CXCL2, iNOS, NF- κ B, COX1, COX2, TNF- α , and Bax by Jamun and a consequent rise in Bcl2 seems to play a crucial role in apoptosis evasion, leading to its protective effect on various tissues. The reduction in AST, ALT, ALP, ACP, GGT, and bilirubin may be an important mechanism to protect the intestine, heart, liver, kidneys, and other tissues.^{111,121,156-159,153,181} Jamun downregulates the mRNA expression of ACC1, Scd1, CD36, and FAS at the protein level, accompanied by a rise in the expression of p-ACC1 protein and normalized expression of Erk1/2 and p-Erk1/2 to reduce hyperlipidemia and diabetes.^{219,232} The anti-obesity effect of Jamun seems to be due to the normalization of pAKT and phosphoinositide 3-kinase (p85) protein expression as well as attrition in the phosphorylation of p-IRS1, the mRNA levels of sterol regulatory element-binding protein 1c, ACC1, FAS, PPAR γ , CD36, and the expression of KDEL, GPR78, XBP1s, XBP1u, MTP, and PDI.^{232,233}

The cytotoxic and chemopreventive actions of Jamun and its active components like myricetin, quercetin, cyanidin, and imperatorin are mediated by the arrest of cells in the G₂+M phase, downregulation of cyclin D1, cyclin D3, Cdk4, Fox1 (Fig. 6), NF- κ B, Nrf2, COX1, COX2, TNF- α , iNOS, and Bcl2, and a rise in the DNA damage and activation of the Bax gene.^{54,287,289,308-312} Recently, NF- κ B has been found in mitochondria, and suppression of NF- κ B and COX leads to the release of cytochrome c from mitochondria, leading to the activation of Bax and p53 and the inhibition of the release of Bcl2, stimulating the intrinsic pathway of apoptosis in cancer cells, as shown in Figure 7.³⁰⁸⁻³¹³ The attenuation of TNF- α by Jamun seems to trigger apoptosis by activating the extrinsic pathway of apoptosis, as indicated in Figure 7.

Conclusions

Various medicinal properties of Jamun are due to the presence of anthraquinones, alkaloids, flavonoids, glycosides, cardiac glycosides, catechins, quinones, phytosterols, tannins, saponins, phenols, resins, terpenoids, steroids, volatile oils, carbohydrates, proteins, and amino acids. Jamun acts by scavenging free radicals, suppressing ICAM1, CXCL2, iNOS, NF- κ B, COX1, COX2, TNF- α , Bcl2, cyclin D1, cyclin D3, Cdk4, and Fox1 expression, and arresting cells in the G₂+M phase of the cell cycle. The decline of AST, ALT, ALP, ACP, GGT, and bilirubin levels as well as elevation of Bcl2 expression by Jamun seem to protect various tissues in preclinical models. Furthermore, Jamun activates PPAR α , PPAR γ , p53, and

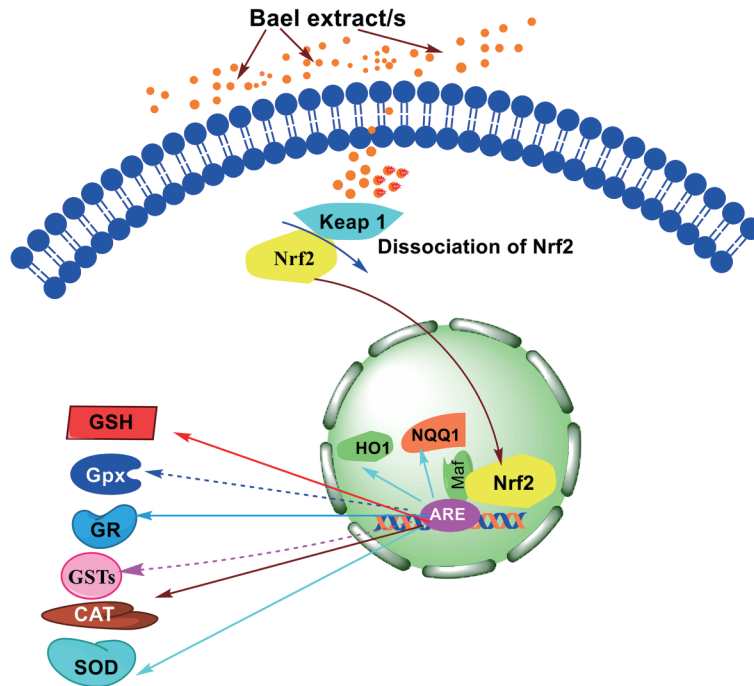


Fig. 4. The effects of Jamun on the antioxidant system. Activation of Nrf2 by different Jamun extracts and isolated active ingredients, such as quercetin, myricetin, ellagic acid, kaempferol, etc., activate the antioxidant system, including glutathione (GSH), glutathione peroxidase (Gpx), glutathione-s-transferase, glutathione reductase, and catalase as well as alleviate lipid peroxidation, thus protecting various organs as well as preventing diabetes and radiation-induced damage.

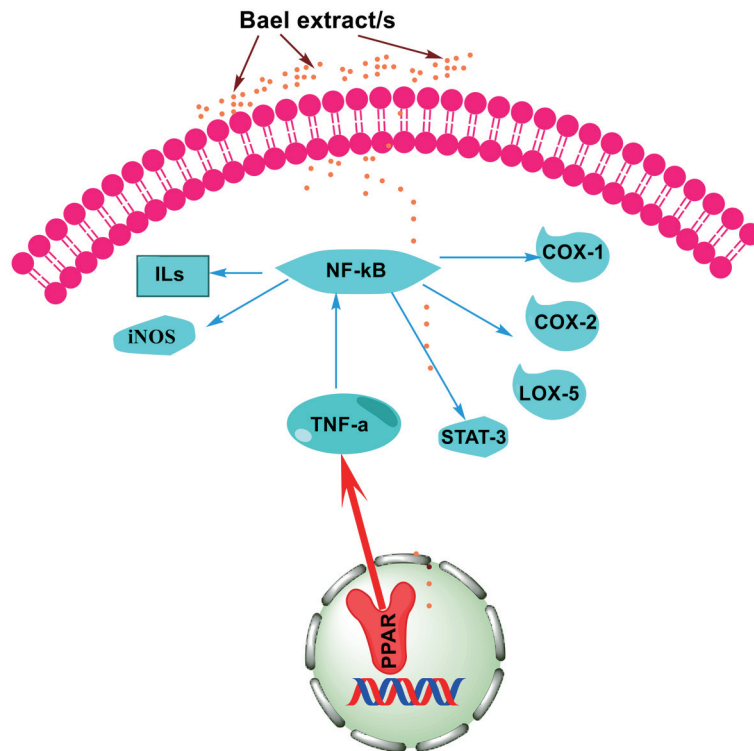


Fig. 5. The molecular pathways involved when cells are treated by Jamun extracts and its active phytochemicals myricetin, quercetin, and cyanidin. They activate peroxisome proliferator-activated receptor (PPAR), which inhibits the transactivation of nuclear factor kappa B (NF-kB), cyclooxygenase (COX) 1 and 2, interleukin (IL) 6, and inducible nitric oxide synthase (iNOS) to protect various organs as well as prevent inflammation and diabetes.

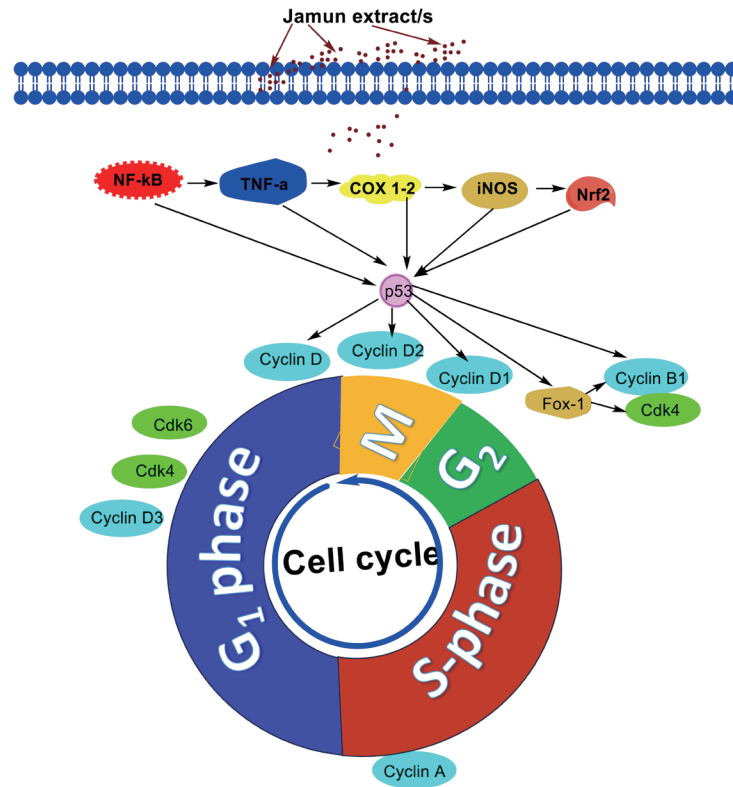


Fig. 6. The effects of Jamun on the cell cycle. Jamun and its active phytochemicals myricetin, quercetin, and cyanidin interfere with the cell cycle by suppressing cyclin D1, cyclin D3, and cyclin-dependent kinase 4 (Cdk4) and arresting cells in the G₂+M phase of the cell cycle. Inhibition of cyclin D3 and Cdk4 and elevation of p53 indicate that Jamun may also actively interfere with other cell cycle phases.

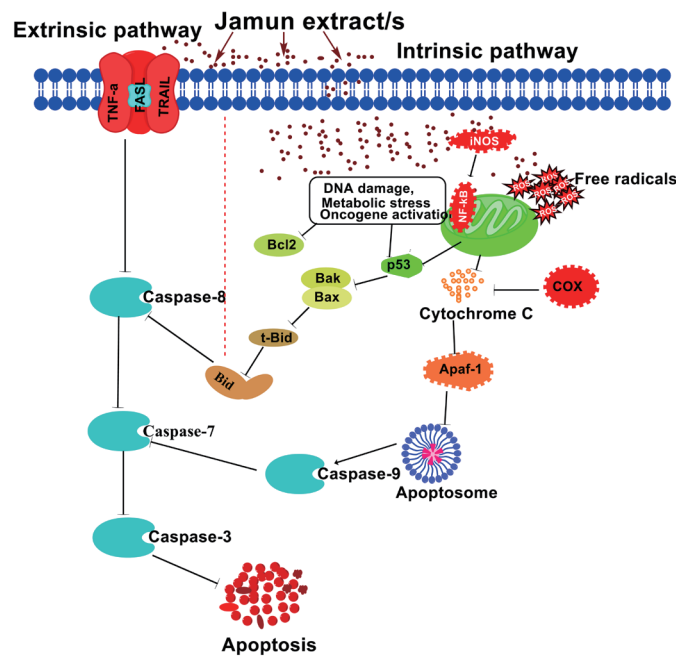


Fig. 7. The effects of Jamun on apoptosis. Jamun and its active phytochemicals myricetin, quercetin, and cyanidin increase reactive oxygen species formation and DNA damage as well as inhibit the transactivation of inducible nitric oxide synthase (iNOS), nuclear factor kappa B (NFκB), cyclooxygenase (COX) 1 and 2, Bcl2, and tumor necrosis factor (TNFα), leading to cytochrome c release from mitochondria and the activation of caspases and Bcl2-associated X (Bax), which subsequently triggers apoptosis to exert cytotoxic effects on neoplastic cells.

Bax and inhibits the expression of ACC1, stearoyl-CoA desaturase 1, CD36, FAS, p-IRS1, KDEL, GPR78, XBP1s, XBP1u, MTP, and PDI to exert its action on different tissues. All activities of Jamun are due to its ability to synthesize secondary metabolites, including myricetin, quercetin, cyanidin, imperatorin, ellagic acid, kaempferol, etc. Therefore, Jamun needs to be developed as the medicine of the future and tested clinically for the benefit of mankind.

The commercialization of Jamun becomes difficult as the phytochemicals in it may vary with the environment, soil composition of the region where it grows, and season of collection. This may compromise the medicinal activities of Jamun due to variations in the secondary metabolites synthesized by it. However, this problem can be solved by growing Jamun in tissue culture for medicinal purposes and incorporating quality control measures batchwise. Future studies need to be directed to unfurl the molecular mechanisms of action of the various medicinal activities of Jamun *in vitro* and *in vivo*.

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Conflict of interest

The author declares no conflict of interests.

Author contributions

GCJ is the sole author of the manuscript.

References

- [1] Crepet WL. Progress in understanding angiosperm history, success, and relationships: Darwin's abominably "perplexing phenomenon". *Proc Natl Acad Sci USA* 2000;97(24):12939–12941. doi:10.1073/pnas.97.24.12939, PMID:11087846.
- [2] Stevenson DW, Zimmermann MH, Stevens P, Dilcher DL, Cronquist A, Berry PE. *Angiosperm*. London: Encycle Britannica; 2023.
- [3] Chakraborty P. Herbal genomics as tools for dissecting new metabolic pathways of unexplored medicinal plants and drug discovery. *Biochim Open* 2018;6:9–16. doi:10.1016/j.biopen.2017.12.003, PMID:29892557.
- [4] Acharya SK. Ethnomedicinal Plants used in Vedic Medicine. *Int J Multidis Res Sci, Eng Technol (IJMRSET)* 2024;7(3):4719–4725. doi:10.15680/IJMRSET.2024.0703036.
- [5] Sen S, Chakraborty R. Revival, modernization and integration of Indian traditional herbal medicine in clinical practice: Importance, challenges and future. *J Tradit Complement Med* 2017;7(2):234–244. doi:10.1016/j.jtcme.2016.05.006, PMID:28417092.
- [6] de Paulo Farias D, Neri-Numa IA, de Araújo FF, Pastore GM. A critical review of some fruit trees from the Myrtaceae family as promising sources for food applications with functional claims. *Food Chem* 2020;306:125630. doi:10.1016/j.foodchem.2019.125630, PMID:31593892.
- [7] da Veiga Correia VT, da Silva PR, Ribeiro CMS, Ramos ALCC, Mazzinghy ACDC, Silva VDM, *et al.* An Integrative Review on the Main Flavonoids Found in Some Species of the Myrtaceae Family: Phytochemical Char-

- acterization, Health Benefits and Development of Products. *Plants (Basel)* 2022;11(20):2796. doi:10.3390/plants11202796, PMID:36297820.
- [8] Kumar M, Zhang B, Nishad J, Verma A, Sheri V, Dhuma S, *et al.* Jamun (*Syzygium cumini* (L.) Skeels) seed: A review on nutritional profile, functional food properties, health-promoting applications, and safety. *Processes* 2022;10:2169. doi:10.3390/PR10112169.
 - [9] Bajpai A, Singh AK, Ravishankar H. Reproductive phenology, flower biology and pollination in jamun (*Syzygium cumini*L.). *Indian J Hort* 2012;69:416–419.
 - [10] Alam M, Mani A, Mitra S, Bauri FK. Flowering, fruiting and physiochemical properties of Jamun (*Syzygium cumini* Skeels) grown in Nadia district of West Bengal. *Adv Biores* 2020;11:1–5. doi:10.15515/abr.0976-4585.11.5.15.
 - [11] Khadivi A, Mirheidari F, Saeidifar A, Moradi Y. Selection of the promising accessions of jamun (*Syzygium cumini* (L.) Skeels) based on pomological characterizations. *Food Sci Nutr* 2023;11(1):470–480. doi:10.1002/fsn3.3078, PMID:36655090.
 - [12] Newman DJ, Cragg GM. Natural Products as Sources of New Drugs over the Nearly Four Decades from 01/1981 to 09/2019. *J Nat Prod* 2020;83(3):770–803. doi:10.1021/acs.jnatprod.9b01285, PMID:32162523.
 - [13] Newman DJ. Natural products and drug discovery. *Natl Sci Rev* 2022;9(11):nwac206. doi:10.1093/nsr/nwac206, PMID:36404871.
 - [14] Shi Y, Zhang C, Li X. Traditional medicine in India. *J Tradit Chinese Med Sci* 2021;8:551–55. doi:10.1016/j.jtcms.2020.06.007.
 - [15] Jaiswal YS, Williams LL. A glimpse of Ayurveda-The forgotten history and principles of Indian traditional medicine. *J Tradit Complement Med* 2017;7(1):50–53. doi:10.1016/j.jtcme.2016.02.002, PMID:28053888.
 - [16] Sharma R, Jadhav M, Choudhary N, Kumar A, Rauf A, Gundamaraju R, *et al.* Deciphering the impact and mechanism of Trikatu, a spices-based formulation on alcoholic liver disease employing network pharmacology analysis and *in vivo* validation. *Front Nutr* 2022;9:1063118. doi:10.3389/fnut.2022.1063118, PMID:36466417.
 - [17] Atanasov AG, Zotchev SB, Dirsch VM, Supuran CT, International Natural Product Sciences Taskforce. Natural products in drug discovery: advances and opportunities. *Nat Rev Drug Discov* 2021;20(3):200–216. doi:10.1038/s41573-020-00114-z, PMID:33510482.
 - [18] Dastur JF. *Useful plants of India and Pakistan*. D. B. Taraporevala Sons & Co., Ltd; 1951.
 - [19] Steinmetz E. A botanical drug from the tropics used in the treatment of *Diabetes mellitus*. *Acta Phyther* 1960;7:23–25. doi:10.1016/S2222-1808(12)60054-1.
 - [20] Jagetia GC. Phytochemical composition and pleiotropic pharmacological properties of Jamun, *Syzygium cumini* Skeels. *J Explor Res Pharmacol* 2017;2:54–66. doi:10.14218/jerp.2016.00038.
 - [21] Morton JF. *Jambolan: In Fruits of warm climates*. London: J. Morton; 1987.
 - [22] Singh S, Singh A, Saroj P. Research status for technological development of jamun (*Syzygium cumini*) in India: A review. *Indian J Agric Sci* 2019;89:1991–1998. doi:10.56093/ijas.v89i12.96260.
 - [23] Singh S, Singh SP, Singh V, Shikha K. Studies on floral biology, fruit set and fruit drop of different genotypes of jamun (*Syzygium cumini* Skeels). *Pharma Innov J* 2019;8:558–561.
 - [24] Jagetia GC. A review on the role of jamun, *Syzygium cumini* Skeels in the treatment of diabetes. *Int J Complement Altern Med* 2018;11:91–95. doi:10.15406/ijcam.2018.11.00374.
 - [25] Gowri S, Vasanth K. Phytochemical screening and antibacterial activity of *Syzygium cumini* (L.) (Myrtaceae) leaves extracts. *Int J PharmTech Res* 2010;2:1569–1573.
 - [26] Rezende W, Borges L, Alves N, Ferri P, Paula J. Chemical variability in the essential oils from leaves of *Syzygium jambos*. *Braz J Pharmacogn* 2013;23:433–440. doi:10.1590/S0102-695X2013005000035.
 - [27] Satyavathi C, Bhavani NL. Evaluation of phytochemical constituents and antibacterial activity in leaf extracts of *Syzygium cumini* L. *World J Pharm Res* 2014;3:768–776.
 - [28] Subramanian R, Subbramanyan P, Raj V. Phytochemical screening, total phenolic contents and antioxidant activity of *Syzygium caryophyllatum* and *Syzygium densiflorum*. *J Biol Act Prod from Nat* 2014;4:224–235. doi:10.1080/22311866.2014.936900.

- [29] Kumar A, Kalakoti M. Phytochemical and antioxidant screening of leaf extract of *Syzygium cumini*. *Int J Adv Res* 2020;3:371–378.
- [30] Ramos IL, Bandiola T. Phytochemical screening of *Syzygium cumini* (Myrtaceae) leaf extracts using different solvents of extraction. *Der Pharm Lett* 2017;9:74–78.
- [31] Ahmed R, Tariq M, Hussain M, Andleeb A, Masoud MS, Ali I, et al. Phenolic contents-based assessment of therapeutic potential of *Syzygium cumini* leaves extract. *PLoS One* 2019;14:e0221318. doi:10.1371/journal.pone.0221318.
- [32] Nikhat F. Total synthetic development and pharmacological screening of bioactive isolated from *Syzygium cumini* (L) skeel. *Biointerface Res Appl Chem* 2020;10:6550–6564. doi:10.33263/BRIAC105.65506564.
- [33] Tewari BB. Basic phytochemical screening and antibacterial, anti-fungal and antioxidant properties of *Syzygium cumini*, a tree from Guyana. *Rev Bolív Química* 2020;37:132–141. doi:10.34098/2078-3949.37.3.1.
- [34] Hasanuzzaman M, Islam W, Islam M. Phytochemical screening of *Syzygium cumini* (L.) extracts in different solvents. *J Bio-Science* 2016;24:83. doi:10.3329/jbs.v24i0.37483.
- [35] Kadlag PR. Preliminary phytochemical evaluation of *Syzygium cumini* (L.) Skeels leaves. *Anveshana's Int J Res Eng App Sci* 2023;8(2):49–51.
- [36] Tewari BB. Profiling the Phytochemical and Pharmacological Properties of *Syzygium cumini*. *Novel Aspects Chem Biochem* 2024;9:97–114. doi:10.9734/bpi/nacb/v9/7296E16;24:11-8.
- [37] Gopinath SM, Rakesh CK, Ashwini Patil GM, Dayananda KS. Preliminary phytochemical evaluation of leaf extracts of *Euphorbia hirta*, *Syzygium cumini* of Siddarabetta, Tumkur district, Karnataka. *Int J Pharma Bio Sci* 2012;3:431–435.
- [38] More RN, Hingmire PM, Jadhav DM. Primary phytochemical and pharmacognostic studies on *Syzygium cumini* Linn. (Jambhul). *J Pharmacog Phytochem* 2024;13(1):299–305. doi:10.22271/phyto.2024.v13.i1d.14843.
- [39] Rauf A, Khan IA, Muhammad N, Al-Awthan YS, Bahattab O, Israr M, Mubarak MS. Phytochemical composition, in vitro urease, α -glucosidase and phosphodiesterase inhibitory potency of *Syzygium cumini* (Jamun) fruits. *South African J Botany* 2021;143:418–421. doi:10.1016/j.sajb.2021.04.006.
- [40] Murti K, Paliwal D, Madan S, Kundu R, Kaushik M. Exploration of preliminary phytochemical studies of seed of *Syzygium cumini*. *Am J Pharmacol Toxicol* 2012;7:12–4. doi:10.3844/ajptsp.2012.12.14.
- [41] Atale N, Jaiswal A, Chhabra A, Malhotra U, Kohli S, Mohanty S, et al. Phytochemical and antioxidant screening of *Syzygium cumini* seed extracts: a comparative study. *J Pharm Res* 2011;4:4530–4532.
- [42] Kamal A. Phytochemical screening of *Syzygium cumini* seeds. *Indian J Plant Sci* 2014;3:2319–3824.
- [43] Mubassara S, Biswas KK, Hasan MM, Hossain MI, Paul S. In vitro phytochemical, antibacterial and antioxidant analyses in different plant parts of *Syzygium cumini*. *Int J Pharmacogn Phytochem Res* 2015;7:150–155.
- [44] Das A, Bharath M, Jeevanantham M, Manoj Kumar S, Thanarithanika RP, Bindhu J. Phytochemical screening and antimicrobial activity of *Syzygium cumini* (Jamun) seed extract. *Res J Pharm Technol* 2018;11:4096–4100. doi:10.5958/0974-360X.2018.00752.7.
- [45] Pandhi S, Poonia A. Phytochemical screening of Jamun seeds using different extraction methods. *Pharma Innov J* 2019;8:226–231.
- [46] Monteiro FS, Carvalho AFS, Ribeiro RM, Borges ACR, Borges MOR. Phytochemical profile and investigation of the spasmolytic activity of hydroalcoholic extract of *Syzygium cumini* (L.) Skeels seeds. *European J Med Plants* 2020;31:27–38. doi:10.9734/ejmp/2020/v31i330220.
- [47] Bari T, Saeed S, Tayyab M. GC-MS Bioactives profiling, antibacterial and cytotoxic potential of jamun (*Syzygium cumini* L.) extracts against food-borne pathogen *Salmonella enteritidis*. *Pharm Chem J* 2024;57:1586–1592. doi:10.1007/s11094-024-03052-x.
- [48] Patel PR, Rao TVR. Antibacterial activity of underutilized fruits of jamun (*Syzygium cumini* L. Skeels). *Int J Curr Pharm Res* 2012;4(1):36–39.
- [49] Nikhat F, Satyanarayana D, Shastri C, Rajni S, Sheikh A. The phytochemicals explored from the roots of *Syzygium cumini* (L) skeel assessed for anti-hyperglycemic activity. *Asian J Res Chem* 2013;6:920–925.
- [50] Sengupta P, Das PB. Terpenoids and related compounds. Part IV. Triterpenoids from the stem bark of *Syzygium cumini*. *Ind Chem Soc* 1965;42:255–258.
- [51] Bhargava KK, Dayar R, Sheshardi TR. Chemical components of *Eugenia jambolana* stem barks. *Curr Sci* 1974;43:645–636.
- [52] Bhatia IS, Bajaj KL. Chemical constituents of the seeds and bark of *Syzygium cumini*. *Planta Med* 1975;28(4):346–352. doi:10.1055/s-0028-1097868, PMID:1208683.
- [53] Kopanski L, Schnelle G. Isolation of Bergein from Barks of *Syzygium cumini*. *Planta Med* 1988;54(6):572. doi:10.1055/s-2006-962577, PMID:17265363.
- [54] Siddika A, Das PK, Asha SY, Aktar S, Tareq ARM, Siddika A, et al. Antiproliferative activity and apoptotic efficiency of *Syzygium cumini* bark methanolic extract against EAC cells in vivo. *Anticancer Agents Med Chem* 2020;21:782–792. doi:10.2174/187152062066200811122137.
- [55] Gupta GS, Sharma DP. Triterpenoid and other constituents of *Eugenia jambolana* leaves. *Phytochemistry* 1974;13:2013–2014. doi:10.1016/0031-9422(74)85151-4.
- [56] Rastogi RP, Mehrotra BN. *Compendium of Indian medicinal plants*. New Delhi: Medicine; 1990.
- [57] Mahmoud II, Marzouk MSA, Moharram FA, El-Gindi MR, Hassan AMK. Acylated flavonol glycosides from *Eugenia jambolana* leaves. *Phytochemistry* 2001;58:1239–1244. doi:10.1016/S0031-9422(01)00365-X.
- [58] Kumar A, Jayachandran T, Aravindhan P, Deecaraman D, Ilavarasan R, Padmanabhan N. Neutral components in the leaves and seeds of *Syzygium cumini*. *African J Pharm Pharmacol* 2009;3:560–561.
- [59] Ghareeb MA, Hamed MM, Abdel-Aleem AAH, Saad AM, Abdel-Aziz MS, Hadad AH. Extraction, isolation, and characterization of bioactive compounds and essential oil from *Syzygium jambos*. *Asian J Pharm Clin Res* 2017;10:194–200. doi:10.22159/ajpcr.2017.v10i8.18849.
- [60] Shidiki A, Vyas A. Evaluation of antibacterial potential of *Syzygium cumini* against methicillin-resistant *Staphylococcus aureus* and macrolide-lincosamide-streptogramin B strains of *Staphylococcus aureus* and its liquid chromatography mass spectroscopy analysis. *Biomed Biotechnol Res J* 2022;6:66–72. doi:10.4103/BBRJ.BBRJ_139_21.
- [61] Shafi PM, Rosamma MK, Jamil K, Reddy PS. Antibacterial activity of *Syzygium cumini* and *Syzygium travancoricum* leaf essential oils. *Fitoterapia* 2002;73(5):414–416. doi:10.1016/S0367-326X(02)00131-4, PMID:12165339.
- [62] Elansary HO, Salem MZM, Ashmawy NA, Yacout MM. Chemical composition, antibacterial and antioxidant activities of leaves essential oils from *Syzygium cumini* L., *Cupressus sempervirens* L. and *Lantana camara* L. from Egypt. *J Agric Sci* 2012;4:4. doi:10.5539/jas.v4n10p144.
- [63] Nishandhini S, Sudha V, Mallavarapu GR, Murugan R. Chemical compositions, α -amylase inhibitory and antioxidant activities of the essential oils from unripe fruit pulp and leaves of *Syzygium cumini*. *Int J Pharm Pharm Sci* 2015;7:511–514.
- [64] Rezende WP, Borges LL, Alves NM, Ferri PH, Paula JR. Chemical variability in the essential oils from leaves of *Syzygium jambos*. *Rev Bras Farmacogn* 2013;23:433–440. doi:10.1590/S0102-695X2013005000035.
- [65] Sarma N, Begum T, Pandey SK, Gogoi R, Munda S, Lal M. Chemical composition of *Syzygium cumini* (L.) Skeels leaf essential oil with respect to its uses from North East region of India. *J Essent Oil-Bearing Plants* 2020;23:601–607. doi:10.1080/0972060X.2020.1796822.
- [66] Hanif MU, Hussain AI, Aslam N, Kamal GM, Chatha SAS, Shahida S, et al. Chemical composition and bioactivities of essential oil from leaves of *Syzygium cumini* (L.) Skeels native to Punjab, Pakistan. *Chem Biodivers* 2020;17:e1900733. doi:10.1002/cbdv.201900733.
- [67] Nair A, Subramanian S. Chemical examination of the flowers of *Eugenia jambolana*. *J Sci Ind Res* 1962;21B:457–458.
- [68] Veigas JM, Narayan MS, Laxman PM, Neelwarne B. Chemical nature, stability and bioefficacies of anthocyanins from fruit peel of *Syzygium cumini* Skeels. *Food Chem* 2007;105:619–627. doi:10.1016/j.foodchem.2007.04.022.
- [69] Li L, Zhang Y, Seeram NP. Structure of anthocyanins from *Eugenia jambolana* fruit. *Nat Prod Commun* 2009;4:217–219. doi:10.1177/1934578x0900400210.
- [70] Faria AF, Marques MC, Mercadante AZ. Identification of bioac-

- tive compounds from jambolão (*Syzygium cumini*) and antioxidant capacity evaluation in different pH conditions. *Food Chem* 2011; 126(4):1571–1578. doi:10.1016/j.foodchem.2010.12.007, PMID:25213929.
- [71] Aqil F, Gupta A, Munagala R, Jeyabalan J, Kausar H, Sharma RJ, et al. Antioxidant and antiproliferative activities of anthocyanin/ellagitanin-enriched extracts from *Syzygium cumini* L. (Jamun, the Indian Blackberry). *Nutr Cancer* 2012;64(3):428–438. doi:10.1080/01635581.2012.657766, PMID:22420901.
- [72] Santos DT, Cavalcanti RN, Rostagno MA, Queiroga CL, Eberlin MN, Meireles MAA. Extraction of polyphenols and anthocyanins from the jambul (*Syzygium cumini*) fruit peels. *Food Public Heal* 2013;3:12–20. doi:10.5923/j.fph.20130301.02.
- [73] Kaume L, Gilbert WC, Brownmiller C, Howard LR, Devareddy L. Cyanidin 3-O-β-d-glucoside-rich blackberries modulate hepatic gene expression, and anti-obesity effects in ovariectomized rats. *J Funct Foods* 2012;4:480–488. doi:10.1016/j.jff.2012.02.008.
- [74] Sharma RJ, Gupta RC, Singh S, Bansal AK, Singh IP. Stability of anthocyanin- and anthocyanidin-enriched extracts, and formulations of fruit pulp of *Eugenia jambolana* ('jamun'). *Food Chem* 2016;190:808–817. doi:10.1016/j.foodchem.2015.06.029, PMID:26213042.
- [75] Singh J, Sharma K, Walia S, Saha S. Anthocyanin profiling method based on isocratic elution for comparable speed, reproducibility, and quantitation with gradient elution. *Food Anal Methods* 2017;10:118–128. doi:10.1007/s12161-016-0561-z.
- [76] Qamar M, Akhtar S, Ismail T, Yuan Y, Ahmad N, Tawab A, et al. *Syzygium cumini* (L.) Skeels fruit extracts: In vitro and in vivo anti-inflammatory properties. *J Ethnopharmacol* 2021;271:113805. doi:10.1016/j.jep.2021.113805.
- [77] Qamar M, Akhtar S, Ismail T, Wahid M, Ali S, Nazir Y, et al. *Syzygium cumini* (L.) Skeels extracts; in vivo anti-nociceptive, anti-inflammatory, acute and subacute toxicity assessment. *J Ethnopharmacol* 2022;287:114919. doi:10.1016/j.jep.2021.114919, PMID:34995693.
- [78] Sudha K, Mathanghi SK, Nirmal RM. GC-MS analysis of bioactive components in Jamun, Amla and Kiwi fruits. *Pharma Innov J* 2023;12:892–899.
- [79] Koop BL, Knapp MA, Di Luccio M, Pinto VZ, Tormen L, Valencia GA, et al. Bioactive compounds from jambolan (*Syzygium cumini* (L.)) extract concentrated by ultra- and nanofiltration: a potential natural antioxidant for food. *Plant Foods Hum Nutr* 2021;76(1):90–97. doi:10.1007/s11130-021-00878-8, PMID:33517518.
- [80] Sapana SK, Jadhav VM, Kadam VI. Development and validation of HPTLC method for determination of 3-hydroxy androstane [16,17-C] (6'methyl, 2'-1-hydroxy-isopropene-1-yl) 4,5,6 H pyran in Jambul seed (*Syzygium cumini*). *Int J PharmTech Res* 2009;1:1129–1135.
- [81] Das G, Nath R, Das Talukdar A, Ağgündüz D, Yılmaz B, Capasso R, et al. Major bioactive compounds from java plum seeds: an investigation of its extraction procedures and clinical effects. *Plants (Basel)* 2023;12(6):1214. doi:10.3390/plants12061214, PMID:36986906.
- [82] Banerjee J, Narendhirakannan R. Phytochemical analyses, antibacterial, in vitro antioxidant and cytotoxic activities of ethanolic extract of *Syzygium cumini* (L.) seed extract. *Academia Edu* 2011;2:1799–1806.
- [83] Atale N, Rani V. GC-MS analysis of bioactive components in the ethanolic and methanolic extract of *Syzygium cumini*. *Int J Pharma Bio Sci* 2013;4:296–304.
- [84] Balyan U, Sarkar B. Aqueous extraction kinetics of phenolic compounds from jamun (*Syzygium cumini* L.) seeds. *Int J Food Prop* 2017;20:372–389. doi:10.1080/10942912.2016.1163266.
- [85] Khan MS, Abul Qais F, Ahmad I, Hussain A, Alajmi MF. Genotoxicity inhibition by *Syzygium cumini* (L.) seed fraction and rutin: understanding the underlying mechanism of DNA protection. *Toxicol Res (Camb)* 2018;7(2):156–171. doi:10.1039/c7tx00269f, PMID:30090571.
- [86] Kadri HS, Minocheherhomji FP. ADMET analysis of phyto-components of *Syzygium cumini* seeds and *Allium cepa* peels. *Futur J Pharm Sci* 2020;6:117. doi:10.1186/S43094-020-00136-9.
- [87] Sindhuja G, Vinolia R, Philomina A, Yasmin M, Agnes A. Phytochemical screening of ethanol extract of *Eugenia jambolana* Lam. *Ann Rom Soc Cell Biol* 2022;24:429–443.
- [88] Ghosh P, Pradhan RC, Mishra S, Patel AS, Kar A. Physicochemical and nutritional characterization of jamun (*Syzygium cumini*). *Curr Res Nutr Food Sci* 2017;5:25–35. doi:10.12944/CRNFSJ.5.1.04.
- [89] Gajera HP, Gevariya SN, Patel SV, Golakiya BA. Nutritional profile and molecular fingerprints of indigenous black jamun (*Syzygium cumini* L.) landraces. *J Food Sci Technol* 2018;55(2):730–739. doi:10.1007/s13197-017-2984-y, PMID:29391638.
- [90] Chiteva R, Onyari J, Njenga L, Madadi V. Physicochemical and nutritional properties of *Syzygium cumini* (L.) skeels fruits grown in varied microclimates in Kenya. *African J Pure Appl Chem* 2023;17:1–9.
- [91] Paul DK, Shaha RK. Nutrients, vitamins and minerals content in common citrus fruits in the Northern region of Bangladesh. *Pakistan J Biol Sci* 2004;7:238–242. doi:10.3923/PJBS.2004.238.242.
- [92] Sardar N, Akbari S, Bhatt H, Tagalpallewar G. Chemical and mineral composition of jamun fruit pulp (*Syzygium cumini* L.). *Pharma Innov J* 2022;11:1976–1979.
- [93] Reynertson KA, Basile MJ, Kennelly EJ. Antioxidant potential of seven Myrtaceae fruits. *Ethnobot Res Appl* 2005;3:25. doi:10.17348/era.3.0.25-36.
- [94] Charaka A. Acharya Charaka, Charaka SamhitaSutrasthana 27th chapter, Shloka No-140, Charaka Chandrika Hindi commentary of Agnivesha. Varanasi: Chaukhamba Surabharati Prakashana; 2002.
- [95] Jain S. Dictionary of Indian folk medicine and ethnobotany: a reference manual of man-plant relationships, ethnic groups & ethnobotanists in India. New Delhi: Deep Publications Paschimviha; 1991.
- [96] Swami SB, Thakor NSJ, Patil MM, Haldankar PM. Jamun (*Syzygium cumini* (L.)): A review of its food and medicinal uses. *Food Nutr Sci* 2012;03:1100–1117. doi:10.4236/fns.2012.38146.
- [97] Bhowmik D, Gopinath H, Kumar BP, Duraivel S, Aravind G, Kumar KPS. Traditional and medicinal uses of Indian black berry. *J Pharmacogn Phytochem* 2013;1:36–41.
- [98] Lal BN, Choudhuri KD. Observations on *Momordica charantia* Linn, and *Eugenia jambolana* Lam. as oral antidiabetic remedies. *Indian J Med Res* 1968;2:161.
- [99] Ayyanar M, Subash-Babu P, Ignacimuthu S. *Syzygium cumini* (L.) Skeels., a novel therapeutic agent for diabetes: folk medicinal and pharmacological evidences. *Complement Ther Med* 2013;21(3):232–243. doi:10.1016/j.ctim.2013.03.004, PMID:23642956.
- [100] Jagetia GC. Antidiabetogenic action of jamun *Syzygium cumini* Skeels: a review. *Int J Complement Altern Med* 2023;16:88–96. doi:10.15406/IJCAM.2023.16.00636.
- [101] Gordon A, Jungfer E, da Silva BA, Maia JG, Marx F. Phenolic constituents and antioxidant capacity of four underutilized fruits from the Amazon region. *J Agric Food Chem* 2011;59(14):7688–7699. doi:10.1021/jf201039r, PMID:21662239.
- [102] Burkhil IH. A dictionary of the economic products of the Malay Peninsula. *Nature* 1936;137:255–255. doi:10.1038/137255c0.
- [103] Quisumbing E. Medicinal Plants of the Philippines. Manila: Bureau of Printing; 1951.
- [104] Damasceno DC, Volpato GT, Calderon IDMP, Rudge MVC. Study of *Averrhoa carambola* and *Eugenia jambolana* extracts purchased from manipulation drugstore on the experimental diabetes. *Rev Bras Toxicol* 2002;15:9–14.
- [105] Ranjan A, Jaiswal A, Raja RB. Enhancement of *Syzygium cumini* (Indian jamun) active constituents by ultra-violet (UV) irradiation method. *Sci Res Essays* 2011;6:2457–2464. doi:10.5897/SRE10.977.
- [106] Jagetia GC, Baliga MS. Evaluation of the radioprotective effect of the leaf extract of *Syzygium cumini* (Jamun) in mice exposed to a lethal dose of gamma-irradiation. *Nahrung* 2003;47(3):181–185. doi:10.1002/food.200390042, PMID:12866620.
- [107] Jagetia GC, Baliga MS, Venkatesh P. Influence of seed extract of *Syzygium cumini* (Jamun) on mice exposed to different doses of gamma-radiation. *J Radiat Res* 2005;46(1):59–65. doi:10.1269/Jrr.46.59, PMID:15802860.
- [108] Silva S do N, Abreu IC, Silva GFC, Ribeiro RM, Lopes A de S, Cartágenes M do S de S, et al. The toxicity evaluation of *Syzygium cumini* leaves in rodents. *Rev Bras Farmacogn* 2011;22:102–108. doi:10.1590/S0102-695X2011005000181.
- [109] Yele SU, Veeranjaneyulu A. Toxicological assessments of aqueous extract of *Eugenia jambolana* stem bark. *Pharm Biol* 2010;48(8):849–854. doi:10.3109/13880200903300204, PMID:20673170.
- [110] Ugbabe GE, Ezeunala MN, Edmond IN, Apev J, Salawu OA. Preliminary phytochemical, antimicrobial and acute toxicity studies of the stem, bark and the leaves of a cultivated *Syzygium cumini* Linn. (fam-

- ily: Myrtaceae) in Nigeria. *African J Biotechnol* 2010;9:6943–6747. doi:10.4314/ajb.v9i41.
- [111] Ayyanna C, Sekar M, Niranjan R, Veera P. Nephrotoxic effect of ethanolic extract of *Syzygium cumini*. *Linn leaves on experimental animals*. *Int J Biol Pharm Res* 2015;6:678–683.
- [112] Bitencourt PER, Ferreira LM, Cargnelutti LO, Denardi L, Boligon A, Fleck M, *et al.* A new biodegradable polymeric nanoparticle formulation containing *Syzygium cumini*: Phytochemical profile, antioxidant and antifungal activity and in vivo toxicity. *Ind Crops Prod* 2016;83:400–407. doi:10.1016/j.indcrop.2016.01.007.
- [113] Prasad M, Venugopal SP, Alagarsamy V, Sridevi C. The preliminary phytochemical analysis and oral acute toxicity study of stem bark of *Syzygium cumini*. *Int J Pharm Pharm Sci* 2016;8:209–213.
- [114] Saleem U, Ahamad B, Shehzad A. Evaluation of mutagenic, acute and sub-acute toxicity potentials of *Syzygium cumini* seeds: A toxicological screening. *J Pharm Res Ther* 2020;1:20–30.
- [115] Jagetia GC, Baliga MS. The evaluation of nitric oxide scavenging activity of certain Indian medicinal plants in vitro: a preliminary study. *J Med Food* 2004;7(3):343–348. doi:10.1089/jmf.2004.7.343, PMID:15383230.
- [116] Ruan ZP, Zhang LL, Lin YM. Evaluation of the antioxidant activity of *Syzygium cumini* leaves. *Molecules* 2008;13(10):2545–2556. doi:10.3390/molecules13102545, PMID:18927517.
- [117] Islam MR, Parvin MS, Islam ME. Antioxidant and hepatoprotective activity of an ethanol extract of *Syzygium jambos* (L.) leaves. *Drug Discov Ther* 2012;6(4):205–211. PMID:23006991.
- [118] Jagetia GC, Shetty PC, Vidyasagar MS. Inhibition of radiation-induced DNA damage by jamun, *Syzygium cumini*, in the cultured splenocytes of mice exposed to different doses of γ -radiation. *Integr Cancer Ther* 2012;11(2):141–153. doi:10.1177/1534735411413261, PMID:21733986.
- [119] Mohamed AA, Ali SI, El-Baz FK. Antioxidant and antibacterial activities of crude extracts and essential oils of *Syzygium cumini* leaves. *PLoS One* 2013;8(4):e60269. doi:10.1371/journal.pone.0060269, PMID:23593183.
- [120] Eshwarappa RS, Iyer RS, Subbaramaiah SR, Richard SA, Dhananjaya BL. Antioxidant activity of *Syzygium cumini* leaf gall extracts. *Bioimpacts* 2014;4(2):101–107. doi:10.5681/bi.2014.018, PMID:25035854.
- [121] Chanudom L, Tangpong J. Anti-Inflammation Property of *Syzygium cumini* (L.) Skeels on Indomethacin-Induced Acute Gastric Ulceration. *Gastroenterol Res Pract* 2015;2015:343642. doi:10.1155/2015/343642, PMID:26633969.
- [122] Margaret E, Shailaja AM, Rao VV. Evaluation of antioxidant activity in different parts of *Syzygium cumini* (Linn.). *Int J Curr Microbiol App Sci* 2015;4:372–379.
- [123] Borges RM, Bitencourt PER, Stein CS, Bochi GV, Boligon A, Morresco RN, *et al.* Leaves and seeds of *Syzygium cumini* extracts produce significant attenuation of 2,2 azobis-2-amidinopropane dihydrochloride-induced toxicity via modulation of ectoenzymes and antioxidant activities. *J Appl Pharm Sci* 2017;7:37–48. doi:10.7324/JAPS.2017.70606.
- [124] Artanti N, Maryani F, Triana Dewi R, Handayani S, Dwiatmi Dewijanti I, Meilawati L, *et al.* In vitro antidiabetic, antioxidant and cytotoxic activities of *Syzygium cumini* fractions from leaves ethanol extract. *Indones J Cancer Chemoprevent* 2019;10:24–29. doi:10.14499/indonesianjcanchemoprev10iss1pp24-29.
- [125] Banerjee A, Dasgupta N, De B. In vitro study of antioxidant activity of *Syzygium cumini* fruit. *Food Chem* 2005;90:727–733. doi:10.1016/j.foodchem.2004.04.033.
- [126] Benherhal PS, Arumughan C. Chemical composition and in vitro antioxidant studies on *Syzygium cumini* fruit. *J Sci Food Agric* 2007;87(14):2560–2569. doi:10.1002/jsfa.2957, PMID:20836162.
- [127] Sudeep HV, Ramachandra YL, Rai SP. Investigation of in vitro, in vivo antioxidant and hepatoprotective activities of *Eugenia jambolana* Lam. stem bark. *J Pharm Res* 2011;4:4167–71.
- [128] Yadav N, Pal A, Sihag S, Nagesh C. Antioxidant activity profiling of acetonic extract of jamun (*Syzygium cumini* L.) seeds in different in-vitro models. *Open Food Sci J* 2020;12:3–8. doi:10.2174/187425640210210003.
- [129] De Oliveira GF, Furtado NAJC, Da Silva Filho AA, Martins CHG, Bastos JK, Cunha WR, *et al.* Antimicrobial activity of *Syzygium cumini* (Myrtaceae) leaves extract. *Brazilian J Microbiol* 2007;38:381–384. doi:10.1590/s1517-83822007000200035.
- [130] Ahsan N, Paul N, Islam N, Akhand AA. Leaf extract of *Syzygium cumini* shows anti-Vibrio activity involving DNA damage. *Dhaka Univ J Pharm Sci* 2012;11:25–28. doi:10.3329/dujps.v11i1.12483.
- [131] Migliato KF, Mello JCP, Higa OZ, Rodas ACD, Corrêa MA, Mendes-Giannini MJS, *et al.* Antimicrobial and cytotoxic activity of fruit extract from *Syzygium cumini* (L.) skeels. *Lat Am J Pharm* 2010;29:725–730.
- [132] Meshram GA, Yadav SS, Shinde D, Patil B, Singh D. Antibacterial study and effect of ethanolic extracts of *Syzygium cumini* seeds powder on glucoamylase invitro. *Biosci Biotechnol Res Asia* 2010;7(1):297–300.
- [133] Pareek A, Meena R, Yadav B.). Antimicrobial activity of *Syzygium cumini*. *Indian Journal of Applied Research* 2015;5(9):64–66.
- [134] Mueller M, Janneon K, Puttipan R, Unger F, Viernstein H, Okonogi S. Anti-inflammatory, antibacterial, and antioxidant activities of Thai medicinal plants. *Int J Pharm Pharm Sci* 2015;7:123–128.
- [135] Bitencourt PER, Cargnelutti LO, Stein CS, Lautenchleger R, Ferreira LM, Sangoi M, *et al.* Anti-inflammatory action of seed extract and polymeric nanoparticles of *Syzygium cumini* in diabetic rats infected with *Candida albicans*. *J Appl Pharm Sci* 2017;7:7–16. doi:10.7324/JAPS.2017.70102.
- [136] Haque R, Sumiya MK, Sakib N, Sarkar OS, Tarek T, Siddique I, Hosain S, Islam A, Khasru Parvez A, Talukder AA, Dey SK. (2017). Antimicrobial activity of jambul (*Syzygium cumini*) fruit extract on enteric pathogenic bacteria. *Adv Microbiol* 2017;7:195–204. doi:10.4236/aim.2017.73016.
- [137] Sharma Y, Mehrotra A, Kundu N, Srivastava NS. A study of antibacterial, antioxidant and neuroprotective effect of stem of *Syzygium cumini*. *Int J Green Pharm* 2017;11:236–243.
- [138] Sharma VK, Chitra D, Charumathy M, Gangadhar L, Anooj ES. Studies on antimicrobial activity of *Syzygium cumini* and *Syzygium alternifolium*. *Ann Trop Med Public Heal* 2020;23:1168–1173. doi:10.36295/ASRO.2020.23742.
- [139] Azzaz N, Hamed S, Mohamed A. Antimicrobial and anticancer activities of *Syzygium cumini* extracts. *J Agric Chem Biotechnol* 2022;13:35–38. doi:10.21608/jacb.2022.118330.1017.
- [140] Chaudhuri AKN, Pal S, Gomes A, Bhattacharya S. Anti-inflammatory and related actions of *Syzygium cumini* seed extract. *Phyther Res* 1990;4:5–10. doi:10.1002/ptr.2650040103.
- [141] Sayeed A, Heuertz R, Ezekiel UR. Curcumin, but not its degradation products, in combination with silibinin is primarily responsible for the inhibition of colon cancer cell proliferation. *MicroPubl Biol* 2022;2022. doi:10.17912/micropub.biology.000617, PMID:35966396.
- [142] Kumar A, Ilavarasan R, Jayachandran T, Deccaraman M, Mohan Kumar R, Aravindan P, *et al.* Anti-inflammatory activity of *Syzygium cumini* seed. *African J Biotechnol* 2008;7:941–943. doi:10.4314/ajb.v7i8.58581.
- [143] Muruganandan S, Srinivasan K, Chandra S, Tandan SK, Lal J, Raviprakash V. Anti-inflammatory activity of *Syzygium cumini* bark. *Fito-terapia* 2001;72(4):369–375. doi:10.1016/s0367-326x(00)00325-7, PMID:11395258.
- [144] Modi DC, Patel JK, Shah BN, Nayak BS. Antiinflammatory activity of seeds of *Syzygium cumini* Linn. *J Pharm Educ Res* 2010;1:68–70.
- [145] Jain A, Sharma S, Goyal M, Dubey S, Jain S, Sahu J, *et al.* Anti-inflammatory activity of *Syzygium cumini* leaves. *Int J Phytomedicine* 2010;2:124–126. doi:10.5138/ijpm.2010.0975.0185.02019.
- [146] Roy A, Bhattacharya S, Pandey JN, Biswas M. Anti-inflammatory activity of *Syzygium cumini* leaf against experimentally induced acute and chronic inflammations in rodents. *Altern Med Stud* 2011;1:6. doi:10.4081/ams.2011.e6.
- [147] Siani AC, Souza MC, Henriques MG, Ramos MF. Anti-inflammatory activity of essential oils from *Syzygium cumini* and *Psidium guajava*. *Pharm Biol* 2013;51(7):881–887. doi:10.3109/13880209.2013.768675, PMID:23577801.
- [148] Machado RRP, Jardim DF, Souza AR, Scio E, Fabri RL, Carpanez AG, *et al.* The effect of essential oil of *Syzygium cumini* on the development of granulomatous inflammation in mice. *Rev Bras Farmacogn* 2013;23:488–496. doi:10.1590/S0102-695X2013005000030.
- [149] Gupta A, Chaphalkar SR. Anti-inflammatory activity of flavonoids from medicinal plants against hepatitis B vaccine antigen on human peripheral blood mononuclear cells. *Asian J Med Pharm Sci*

- 2015;3:728–732.
- [150] Hidayah H, Gulo AF, Zulfa AA, Yunita D. Activity of the triterpenoid content of jamun as an anti-inflammatory compound. *Jurnal Multi-disiplin Indonesia* 2023;2(6):911–916.
- [151] Brito FA, Lima LA, Ramos MFS, Nakamura MJ, Cavalher-Machado SC, Siani AC, *et al.* Pharmacological study of anti-allergic activity of *Syzygium cumini* (L.) Skeels. *Brazilian J Med Biol Res* 2007;40:105–115. doi:10.1590/S0100-879X2007000100014.
- [152] Balakrishna G, Sowmya K, Bollapalli V, Rao M. Anti-allergic studies of *Albizia lebbek* and *Syzygium cumini* (L- *Syzygium gambolana*). *Open Access J Microbiol Biotechnol* 2016;1:000103. doi:10.23880/oajmb-16000103.
- [153] Moresco RN, Sperotto RL, Bernardi AS, Cardoso RF, Gomes P. Effect of the aqueous extract of *Syzygium cumini* on carbon tetrachloride-induced hepatotoxicity in rats. *Phytother Res* 2007;21(8):793–795. doi:10.1002/ptr.2158, PMID:17450508.
- [154] Veigas JM, Shrivastava R, Neelwarne B. Efficient amelioration of carbon tetrachloride induced toxicity in isolated rat hepatocytes by *Syzygium cumini* Skeels extract. *Toxicol In Vitro* 2008;22(6):1440–1446. doi:10.1016/j.tiv.2008.04.015, PMID:18538978.
- [155] Das S, Sarma G. Study of the hepatoprotective activity of the ethanolic extract of the pulp of *Eugenia jambolana* (Jamun) in albino rats. *J Clin Diagnostic Res* 2009;3:1466–1474.
- [156] Sisodia SS, Bhatnagar M. Hepatoprotective activity of *Eugenia jambolana* Lam. in carbon tetrachloride treated rats. *Indian J Pharmacol* 2009;41(1):23–27. doi:10.4103/0253-7613.48888, PMID:20177577.
- [157] Donepudi AC, Aleksunes LM, Driscoll MV, Seeram NP, Slitt AL. The traditional ayurvedic medicine, *Eugenia jambolana* (Jamun fruit), decreases liver inflammation, injury and fibrosis during cholestasis. *Liver Int* 2012;32(4):560–573. doi:10.1111/j.1478-3231.2011.02724.x, PMID:22212619.
- [158] Sharma B, Siddiqui MS, Kumar SS, Ram G, Chaudhary M. Liver protective effects of aqueous extract of *Syzygium cumini* in Swiss albino mice on alloxan induced *Diabetes mellitus*. *J Pharm Res* 2013;6:853–858. doi:10.1016/j.jopr.2013.07.020.
- [159] Behera SR, Sekkizhar M, K SB. Hepatoprotective activity of aqueous extract of *Syzygium cumini* seed on streptozotocin induced diabetes in rats. *Int J Ayurvedic Herb Med* 2014;2:1470–1477.
- [160] Islam M, Hussain K, Latif A, Hashmi FK, Saeed H, Bukhari NI, *et al.* Evaluation of extracts of seeds of *Syzygium cumini* L. for hepatoprotective activity using CCl₄-induced stressed rats. *Pak Vet J* 2015;35:197–200.
- [161] Sobeh M, Esmat A, Petruk G, Abdelfattah MAO, Dmirieh M, Monti DM, *et al.* Phenolic compounds from *Syzygium jambos* (Myrtaceae) exhibit distinct antioxidant and hepatoprotective activities in vivo. *J Funct Foods* 2018;41:223–231. doi:10.1016/j.jff.2017.12.055.
- [162] Srivastava BD, Srivastava M, Srivastav SK, Suzuki N, Srivastav AK. Cypermethrin-induced liver histopathology in rat: Protective role of jamun seed and orange peel extracts. *Iran J Toxicol* 2018;12:25–30. doi:10.32598/ijt.12.4.74.7.
- [163] Kumar M, Thakur R, Kumar S. Comparative efficacy of *Syzygium cumini* seed extracts in alleviating arsenic-induced hepatotoxicity and blood cell genotoxicity in Wistar albino rats. *Biomed Pharmacol J* 2019;12:1329–1238.
- [164] Abbas T, Ahmad KR, Akhtar HA, Fatima T. Ameliorative activities of morus and jamun against Cr induced andro-hepatic anomalies. *Eur J Nut Food Safety* 2024;16(1):66–78.
- [165] Ramirez RO, Roa CC. The gastroprotective effect of tannins extracted from duhat (*Syzygium cumini* Skeels) bark on HCl/ethanol induced gastric mucosal injury in Sprague-Dawley rats. *Clin Hemorheol Microcirc* 2003;29:253–261.
- [166] Riaz MN, Sajid MI, Jamshaid M, Khan G, Shahzad M, Shahzad M, *et al.* An in vitro study on *Eugenia jambolana* plant extract in isolated rabbit ileum showing spasmolytic effects. *Can J Appl Sci* 2014;3:1–16.
- [167] Chaturvedi A, Mohan Kumar M, Bhawani G, Chaturvedi H, Kumar M, Goel RK. Effect of ethanolic extract of *Eugenia jambolana* seeds on gastric ulceration and secretion in rats. *Indian J Physiol Pharmacol* 2007;51:131–140.
- [168] Chaturvedi A, Bhawani G, Agarwal PK, Goel S, Singh A, Goel RK. Antidiabetic and antiulcer effects of extract of *Eugenia jambolana* seed in mild diabetic rats: study on gastric mucosal offensive acid-pepsin secretion. *Indian J Physiol Pharmacol* 2009;53(2):137–146. PMID:20112817.
- [169] Sanchez JG, Rancu AL, Diatta FH, Jonnalagadda A, Dhodapkar MM, Knoedler L, *et al.* Increased risk of 90-day complications in patients with fibromyalgia undergoing total shoulder arthroplasty. *J Am Acad Orthop Surg Glob Res Rev* 2024;8(5):e24.00102. doi:10.5435/JAOS-Global-D-24-00102, PMID:38722914.
- [170] Mastan SK, Bhavya Latha T, Sri Latha T, Srikanth A, Chaitanya G, Eswar Kumar K. Influence of methanolic extract of *Syzygium cumini* seeds on the activity of gliclazide in normal and alloxan-induced diabetic rats. *Pharmacologyonline* 2009;3:845–850.
- [171] Soncharan P, Shanmugarajan TS, Somasundaram I, Niladri M. Protective effect of *Syzygium cumini* seeds against doxorubicin. *Citeseer* 2010;1:343–349.
- [172] Atale N, Chakraborty M, Mohanty S, Bhattacharya S, Nigam D, Sharma M, *et al.* Cardioprotective role of *Syzygium cumini* against glucose-induced oxidative stress in H9C2 cardiac myocytes. *Cardiovasc Toxicol* 2013;13:278–289. doi:10.1007/s12012-013-9207-1.
- [173] Atale N, Rani V. *Syzygium cumini*: An effective cardioprotective via its antiglycoxidation potential. *Int J Pharm Sci Rev Res* 2016;37:42–51.
- [174] Shukla SK, Sharma SB, Singh UR, Ahmad S, Maheshwari A, Misro M, *et al.* *Eugenia jambolana* pretreatment prevents isoproterenol-induced myocardial damage in rats: Evidence from biochemical, molecular, and histopathological studies. *J Med Food* 2014;17:244–253. doi:10.1089/jmf.2013.2795.
- [175] Ribeiro RM, Pinheiro Neto VF, Ribeiro KS, Vieira DA, Abreu IC, Silva SDN, *et al.* Antihypertensive effect of *Syzygium cumini* in spontaneously hypertensive Rats. *Evidence-Based Complement Altern Med* 2014;2014:605452. doi:10.1155/2014/605452.
- [176] De A Herculano E, Df Da Costa C, Rodrigues AK, Araújo-Júnior JX, Santana AE, Hb França P, *et al.* Evaluation of cardiovascular effects of edible fruits of *Syzygium cumini* Myrtaceae (L) Skeels in rats. *Trop J Pharm Res* 2014;13:1853–1861. doi:10.4314/tjpr.v13i11.12.
- [177] Sanwalka DN. Study of effect of jamun (*Syzygium cumini*) seed powder on glycaemic control and dyslipidemia in type 2 *Diabetes mellitus* a double blind randomized control trial. *J Med Sci Clin Res* 2019;7:753–755. doi:10.18535/jmscr/v7i9.71.
- [178] Assis KS, Araújo IGA, De Azevedo FDAA, Maciel PMP, Machado Calzerra NT, Da Silva TAF, *et al.* Potassium channel activation is involved in the cardiovascular effects induced by freeze dried *Syzygium jambolanum* (Lam.) DC fruit juice. *Biomed Res Int* 2018;2018:4827461. doi:10.1155/2018/4827461.
- [179] Atale N, Mishra CB, Kohli S, Mongre RK, Prakash A, Kumari S, *et al.* Anti-inflammatory effects of *S. cumini* seed extract on gelatinase-B (MMP-9) regulation against hyperglycemic cardiomyocyte stress. *Oxid Med Cell Longev* 2021;2021:8839479. doi:10.1155/2021/8839479.
- [180] Adikay S, Belide P, Koganti B. Protective effect of fruits of *Syzygium cumini* against cisplatin-induced acute renal failure in rats. *J Pharm Res* 2010;3:2756–2758.
- [181] Mahalakshmi A, Prasanna G. Nephroprotective effect of *Syzygium cumini* Linn. against paracetamol induced toxicity in albino rats. *Adv Pharmacol Toxicol* 2011;12:25–32.
- [182] Behera SR, Sekkizhar M, Babu SK. Nephro-protective effect of aqueous extract of *Syzygium cumini* seed on streptozotocin induced diabetes in rats. *Int J Chem Lifesciences* 2014;3:1285–1288.
- [183] Sathish Babu P, Krishnan G, Anand Babu K, Chitra K. In silico and in vitro evaluation of anti-urolithiatic activity of ethanolic extract of *Syzygium cumini* stem bark. *Res J Pharm Technol* 2017;10:1317–1321. doi:10.5958/0974-360X.2017.00233.5.
- [184] Sutariya B, Taneja N, Saraf M. Betulinic acid, isolated from the leaves of *Syzygium cumini* (L.) Skeels, ameliorates the proteinuria in experimental membranous nephropathy through regulating Nrf2/NF-κB pathways. *Chem Biol Interact* 2017;274:124–137. doi:10.1016/j.cbi.2017.07.011, PMID:28711658.
- [185] Xie J, Wang M, Long Z, Ning H, Li J, Cao Y, *et al.* Global burden of type 2 diabetes in adolescents and young adults, 1990–2019: systematic analysis of the Global Burden of Disease Study 2019. *BMJ* 2022;379:e072385. doi:10.1136/bmj-2022-072385, PMID:36740855.
- [186] Jagetia GC. Jamun *Syzygium cumini* Skeels in the treatment of dia-

- betes. Clin J Diabetes Care Control 2022;5:180042.
- [187] Helmstädtter A. *Syzygium cumini* (L.) SKEELS (Myrtaceae) against diabetes—125 years of research. Pharmazie 2008;63(2):91–101. PMID:18380393.
- [188] WASTL H, BOERICKE GW, FOSTER WC. Studies of effects of *Syzygium jambolanum* on alloxan-diabetic rats. Arch Int Pharmacodyn Ther 1947;75(1):33–50. PMID:18905801.
- [189] Pepato MT, Mori DM, Baviera AM, Harami JB, Vendramini RC, Brunetti IL. Fruit of the jambolan tree (*Eugenia jambolana* Lam.) and experimental diabetes. J Ethnopharmacol 2005;96(1-2):43–48. doi:10.1016/j.jep.2004.07.029, PMID:15588649.
- [190] Brahmachari HD, Augusti KT. Hypoglycaemic agents from Indian indigenous plant. J Pharm Pharmacol 1961;13:381–382. doi:10.1111/j.2042-7158.1961.tb11839.x.
- [191] Kedar P, Chakrabarti CH. Effects of jambolan seed treatment on blood sugar, lipids and urea in streptozotocin induced diabetes in rabbits. Indian J Physiol Pharmacol 1983;27(2):135–140. PMID:6885126.
- [192] Chirvan-Nia P, Ratsimamanga AR. Regression of cataract and hyperglycemia in diabetic sand rats (*Psammomys obesus*) having received an extract of *Eugenia Jambolana* (Lamarck). C R Acad Hebd Seances Acad Sci D 1972;274(2):254–257. PMID:4622072.
- [193] Ratsimamanga A, Loiseau A, Ratsimamanga-Urveg S, Bibal-Prot P. Nouvelle contribution à l'étude de l'action d'un principe hypoglycémiant mis en évidence dans d'écorce jeune de *Eugenia jambolana* (Myrtacées) sur l'hyperglycémie provoquée du lapin normal et poursuite de sa purification. Comptes Rendus Hebd Des Seances l'Academie Des Sci Ser D Sci Nat 1973;277:2219–2222.
- [194] Nair RB, Santhakumari G. Anti-diabetic activity of the seed kernel of *Syzygium cumini* linn. Anc Sci Life 1986;6(2):80–84. PMID:22557552.
- [195] Prince PSM, Menon VP, Pari L. Hypoglycaemic activity of *Syzygium cumini* seeds: Effect on lipid peroxidation in alloxan diabetic rats. J Ethnopharmacol 1998;61:1–7. doi:10.1016/S0378-8741(98)00002-6.
- [196] Achrekar S, Kaklij G, Pote MS, Kelkar SM. Hypoglycemic activity of *Eugenia jambolana* and *Ficus bengalensis*: mechanism of action. In Vivo (Brooklyn) 1991;5:143–147.
- [197] Vikrant V, Grover JK, Tandon N, Rathi SS, Gupta N. Treatment with extracts of *Momordica charantia* and *Eugenia jambolana* prevents hyperglycemia and hyperinsulinemia in fructose fed rats. J Ethnopharmacol 2001;76(2):139–143. doi:10.1016/S0378-8741(01)00218-5, PMID:11390126.
- [198] Grover JK, Vats V, Rathi SS, Dawar R. Traditional Indian anti-diabetic plants attenuate progression of renal damage in streptozotocin induced diabetic mice. J Ethnopharmacol 2001;76(3):233–238. doi:10.1016/S0378-8741(01)00246-x, PMID:11448544.
- [199] Pandey M, Khan A. Hypoglycaemic effect of defatted seeds and water soluble fibre from the seeds of *Syzygium cumini* (Linn.) skeels in alloxan diabetic rats. Indian J Exp Biol 2002;40(10):1178–1182. PMID:12693701.
- [200] Sridhar SB, Sheetal UD, Pai MR, Shastri MS. Preclinical evaluation of the antidiabetic effect of *Eugenia jambolana* seed powder in streptozotocin-diabetic rats. Braz J Med Biol Res 2005;38(3):463–468. doi:10.1590/S0100-879X2005000300018, PMID:15761627.
- [201] Sharma SB, Nasir A, Prabhu KM, Murthy PS, Dev G. Hypoglycaemic and hypolipidemic effect of ethanolic extract of seeds of *Eugenia jambolana* in alloxan-induced diabetic rabbits. J Ethnopharmacol 2003;85(2-3):201–206. doi:10.1016/S0378-8741(02)00366-5, PMID:12639741.
- [202] Ravi K, Ramachandran B, Subramanian S. Effect of *Eugenia jambolana* seed kernel on antioxidant defense system in streptozotocin-induced diabetes in rats. Life Sci 2004;75(22):2717–2731. doi:10.1016/j.lfs.2004.08.005, PMID:15369706.
- [203] Ravi K, Ramachandran B, Subramanian S. Protective effect of *Eugenia jambolana* seed kernel on tissue antioxidants in streptozotocin-induced diabetic rats. Biol Pharm Bull 2004;27(8):1212–1217. doi:10.1248/bpb.27.1212, PMID:15305024.
- [204] Ravi K, Rajasekaran S, Subramanian S. Antihyperlipidemic effect of *Eugenia jambolana* seed kernel on streptozotocin-induced diabetes in rats. Food Chem Toxicol 2005;43(9):1433–1439. doi:10.1016/j.fct.2005.04.004, PMID:15964674.
- [205] Mulkalwar S, Kulkarni V, Deshpande T, Bhide H, Patel A, Tilak AV. Antihyperglycemic activity of *Syzygium cumini* (jamun) in diabetic rats. J Pharma Res Int 2021;33:12–19.
- [206] Kumar A, Ilavarasan R, Jayachandran T, Deecaraman M, Aravindan P, Padmanabhan N, et al. Anti-diabetic activity of *Syzygium cumini* and its isolated compound against streptozotocin-induced diabetic rats. J Med Plants Res 2008;2:246–249.
- [207] Bhuyan ZA, Rokeya B, Masum N, Hossain S, Mahmud I. Antidiabetic effect of *Syzygium cumini* l. Seed on type 2 diabetic rats. Dhaka Univ J Biol Sci 2010;19:157–164. doi:10.3329/dujbs.v19i2.8959.
- [208] Sharma SB, Rajpoot R, Nasir A, Prabhu KM, Murthy PS. Ameliorative Effect of active principle isolated from seeds of *Eugenia jambolana* on carbohydrate metabolism in experimental diabetes. Evid Based Complement Alternat Med 2011;2011:789871. doi:10.1093/ecam/nep233, PMID:21811514.
- [209] Sharma SB, Tanwar RS, Nasir A, Prabhu KM. Antihyperlipidemic effect of active principle isolated from seed of *Eugenia jambolana* on alloxan-induced diabetic rabbits. J Med Food 2011;14(4):353–359. doi:10.1089/jmf.2010.1227, PMID:21370965.
- [210] Sharma AK, Bharti S, Kumar R, Krishnamurthy B, Bhatia J, Kumari S, et al. *Syzygium cumini* ameliorates insulin resistance and β -cell dysfunction via modulation of PPAR γ , dyslipidemia, oxidative stress, and TNF- α in type 2 diabetic rats. J Pharmacol Sci 2012;119:203–213. doi:10.1254/jphs.11184FP, PMID:22786584.
- [211] Sharma SB, Nasir A, Prabhu KM, Murthy PS. Antihyperglycemic effect of the fruit-pulp of *Eugenia jambolana* in experimental *Diabetes mellitus*. J Ethnopharmacol 2006;104(3):367–373. doi:10.1016/j.jep.2005.10.033, PMID:16386863.
- [212] Rekha N, Balaji R, Deecaraman M. Antihyperglycemic and antihyperlipidemic effects of extracts of the pulp of *Syzygium cumini* and bark of *Cinnamon zeylanicum* in streptozotocin-induced diabetic rats. J Appl Biosci 2010;28:1718–1730.
- [213] Deb L, Bhattacharjee C, Sherty S, Dutta A. Evaluation of anti-diabetic potential of the *Syzygium cumini* (Linn) Skeels by reverse pharmacological approaches. Bull Pharm Res 2013;3:135–145.
- [214] Siddiqui M, Sharma B, Ram G. Anti-hyperglycemic and anti-hyperlipemia effects of *Syzygium cumini* seed in alloxan induced *Diabetes mellitus* in Swiss albino mice (*Mus musculus*). Med Aromat Plants 2014;3:166.
- [215] Gajera HP, Gevariya SN, Hirpara DG, Patel SV, Golakiya BA. Antidiabetic and antioxidant functionality associated with phenolic constituents from fruit parts of indigenous black jamun (*Syzygium cumini* L.) landraces. J Food Sci Technol 2017;54(10):3180–3191. doi:10.1007/s13197-017-2756-8, PMID:28974803.
- [216] Raza A, Butt MS, Ishaq-Ul-Haq, Suleria HAR. Jamun (*Syzygium cumini*) seed and fruit extract attenuate hyperglycemia in diabetic rats. Asian Pac J Trop Biomed 2017;7:750–754. doi:10.1016/j.apjtb.2017.07.006.
- [217] Nahid S, Mazumder K, Rahman Z, Islam S, Rashid MH, Kerr PG. Cardio- and hepato-protective potential of methanolic extract of *Syzygium cumini* (L.) Skeels seeds: A diabetic rat model study. Asian Pac J Trop Biomed 2017;7:126–133. doi:10.1016/j.apjtb.2016.11.025.
- [218] Sharma S, Pathak S, Gupta G, Sharma SK, Singh L, Sharma RK, et al. Pharmacological evaluation of aqueous extract of *Syzygium cumini* for its antihyperglycemic and antidyslipidemic properties in diabetic rats fed a high cholesterol diet—Role of PPAR γ and PPAR α . Biomed Pharmacother 2017;89:447–453. doi:10.1016/j.biopha.2017.02.048, PMID:28249245.
- [219] Xu J, Liu T, Li Y, Yuan C, Ma H, Seeram NP, et al. Hypoglycemic and hypolipidemic effects of triterpenoid-enriched Jamun (*Eugenia jambolana* Lam.) fruit extract in streptozotocin-induced type 1 diabetic mice. Food Funct 2018;9:3330–3337. doi:10.1039/c8fo00095f, PMID:29808185.
- [220] Ishartati E, Sufianto, Rohman S, Sukardi. Bioactive properties and anti-diabetic potential of black jamun (*Syzygium cumini* (L.) Skeels) pulp and seed extracts. Med Plants 2022;14:421–428. doi:10.5958/0975-6892.2022.00045.4.
- [221] Ashfaq M, Imran M, Tufail T, Aslam M, Shahid M, Fatima A, Inayat S. Effect of different concentrations of jamun and amla extracts to combat *Diabetes mellitus*. Pakistan BioMedical Journal 2022;5(1):276–281. doi:10.54393/pbmj.v5i1.200.
- [222] Sudha K, Baskaran D, Narayanan R. Evaluation of In vitro antioxidant, antidiabetic and anti-lipase activities of selected fruits (Amla,

- Grape, Jamun and Kiwi). *Pharma Innovation J* 2023;12(7):114–121.
- [223] Bopp A, De Bona KS, Bellé LP, Moresco RN, Moretto MB. *Syzygium cumini* inhibits adenosine deaminase activity and reduces glucose levels in hyperglycemic patients. *Fundam Clin Pharmacol* 2009;23(4):501–507. doi:10.1111/j.1472-8206.2009.00700.x, PMID: 19709327.
- [224] De Bona KS, Bellé LP, Sari MH, Thomé G, Schetinger MRC, Morsch VM, *et al.* *Syzygium cumini* extract decrease adenosine deaminase, 5'nucleotidase activities and oxidative damage in platelets of diabetic patients. *Cell Physiol Biochem* 2010;26:729–738. doi:10.1159/000322340, PMID:21063110.
- [225] Ayya N, Nalwade V, Khan TN. Effect of jamun (*Syzygium cumini* L.) seed powder supplementation on blood glucose level of type-II diabetic subject. *Food Sci Res J* 2015;6:353–356. doi:10.15740/has/fsrj/6.2/353-356.
- [226] Sidana S, Singh V, Meena B, Beniwal S, Chandra S, Singh K, *et al.* Effect of *Syzygium cumini* (jamun) seed powder on dyslipidemia: A double blind randomized control trial. *Int J Res Med Sci* 2016;4:2603–2610. doi:10.18203/2320-6012.ijrms20161917.
- [227] Banu H, Jyothi A. Hypoglycemic and hypo cholesterolemic effect of *Eugenia jambolana* (Kala Jamun) spicy mix on Type II diabetic subjects. *Imp J Interdiscip Res* 2016;2:850–857.
- [228] Nayak MR, Mishra R. Effectiveness of fenugreek seeds versus jamun seeds on blood glucose level among pre-diabetic clients in a selected urban community. *Odisha Res J Berhampur Uni* 2022;4:86–96.
- [229] Raza A, Usman Ali M, Akram MN, Wazir I, Nawaz Sharif M. Anti-hypercholesterolemic role of ethanolic extract of jamun (*Syzygium cumini*) fruit and seed in hypercholesterolemic rats. *Am J Agric Environ Sci* 2015;15:1012–1018.
- [230] Parveen S, Khan AA, Khan QA. Antihyperlipidemic effect of seeds of jamun (*Eugenia jambolana*) in subjects of intermediate hyperglycemia: A pilot study. *Tradit Integr Med* 2020;5:191–197. doi:10.18502/tim.v5i4.5164.
- [231] Ulla A, Alam MA, Sikder B, Sumi FA, Rahman MM, Habib ZF, *et al.* Supplementation of *Syzygium cumini* seed powder prevented obesity, glucose intolerance, hyperlipidemia and oxidative stress in high carbohydrate high fat diet induced obese rats. *BMC Complement Altern Med* 2017;17:289. doi:10.1186/s12906-017-1799-8, PMID:28578702.
- [232] Xu J, Liu T, Li Y, Liu W, Ding Z, Ma H, *et al.* Jamun (*Eugenia jambolana* Lam.) fruit extract prevents obesity by modulating the gut microbiome in high-fat-diet-fed mice. *Mol Nutr Food Res* 2019;63:e1801307. doi:10.1002/mnfr.201801307, PMID:30762938.
- [233] França LM, Ferreira Coelho CF, Costa Freitas LN, Santos Souza IL, Chagas VT, Debbas V, *et al.* *Syzygium cumini* leaf extract reverts hypertriglyceridemia via downregulation of the hepatic XBP-1s/PDI/MTP axis in monosodium l-glutamate-induced obese rats. *Oxid Med Cell Longev* 2019;2019:9417498. doi:10.1155/2019/9417498, PMID:30762938.
- [234] Mahapatra PK, Chakraborty D, Chaudhuri AK. Anti-inflammatory and antipyretic activities of *Syzygium cumini*. *Planta Med* 1986;52(5):540. doi:10.1055/s-2007-969339, PMID:17345495.
- [235] Alyas S, Zahra N, Zahid N, Nisar A, Ahmad M. Anti-inflammatory, antipyretic and analgesic activities of ethanol extract of *Eugenia jambolana* Lam. *Int J Biosci* 2020;16:506–511. doi:10.12692/ijb/16.3.493-498.
- [236] Shamkuwar PB, Pawar DP, Chauhan SS. Antidiarrhoeal activity of seeds of *Syzygium cumini* L. *J Pharm Res* 2012;9:5537–5539.
- [237] Mukherjee PK, Saha K, Murugesan T, Mandal SC, Pal M, Saha BP. Screening of anti-diarrhoeal profile of some plant extracts of a specific region of West Bengal, India. *J Ethnopharmacol* 1998;60(1):85–89. doi:10.1016/s0378-8741(97)00130-x, PMID:9533436.
- [238] Abedin F, Hussain MS, Islam A, Sen N, Das A, Kar A, *et al.* Thrombolytic, CNS depressant and anti-diarrhoeal activities of ethanolic extract of bark of *Syzygium cumini* L. Skeels: An in-vivo and in-vitro study. *J Pharm Nutr Sci* 2018;8:129–136. doi:10.6000/1927-5951.2018.08.03.7.
- [239] Monteiro F de S, Carvalho AFS, Marques E de C, Ribeiro RM, Borges ACR, Borges MO da R. Antidiarrhoeal and antispasmodic activity of leaves of *Syzygium cumini* L. (Myrtaceae) mediated through calcium channel blockage. *African J Pharm Pharmacol* 2018;12:11–18. doi:10.5897/ajpp2017.4868.
- [240] Kayser MS, Nath R, Khatun H, Rashid MA. Peripheral analgesic and anti-diarrheal activities of leaf of *Syzygium cumini* (L.) Skeel. *Bangladesh Pharm J* 2019;22:13–17. doi:10.3329/bpj.v22i1.40020.
- [241] Dias CN, Rodrigues KAF, Carvalho FAA, Carneiro SMP, Maia JGS, Andrade EHA, *et al.* Molluscicidal and leishmanicidal activity of the leaf essential oil of *Syzygium cumini* (L.) skeels from Brazil. *Chem Biodivers* 2013;10:1133–41. doi:10.1002/cbdv.201200292.
- [242] Rodrigues KADF, Amorim LV, Dias CN, Moraes DFC, Carneiro SMP, Carvalho FADA. *Syzygium cumini* (L.) Skeels essential oil and its major constituent α -pinene exhibit anti-Leishmania activity through immunomodulation in vitro. *J Ethnopharmacol* 2015;160:32–40. doi:10.1016/j.jep.2014.11.024.
- [243] Archana N, Ramasamy M, David Raj C. Pharmacological screening of ethanolic extract of *Syzygium cumini* seed on isolated smooth muscle strip and heart. *Int J Pharm Pharm Sci* 2012;4:108–110.
- [244] Boskabady MH, Jandaghi P. Relaxant effects of carvedilol on guinea pig tracheal chains and its possible mechanisms. *Pharmazie* 2003;58(9):661–663. PMID:14531466.
- [245] Riaz MN. An invitro study on *Eugenia jambolana* plant extract in isolated rabbit ilium showing spasmolytic effects. *Can J Appl Sci* 2014;4:1. doi:10.21065/19257430.4.1.
- [246] Monteiro FS, Carvalho AF, Ribeiro RM, Borges AC, Borges MO. Phytochemical profile and investigation of the spasmolytic activity of hydroalcoholic extract of *Syzygium cumini* (L.) Skeels Seeds. *Eur J Med Plants* 2020;31(3):27–38. doi:10.9734/EJMP/2020/v31i330220.
- [247] Priya S, Devi P, Madeswaran A. In silico docking studies of RP2 (X-Linked retinitis pigmentosa) protein using anthocyanins as potential inhibitors. *Bangladesh J Pharmacol* 2013;8:292–299.
- [248] Alikatte KL, Akondi BR, Yerragunta VG, Veerareddy PR, Palle S. Antiamnesic activity of *Syzygium cumini* against scopolamine induced spatial memory impairments in rats. *Brain Dev* 2012;34(10):844–851. doi:10.1016/j.braindev.2012.02.008, PMID:22475379.
- [249] Hossain S, Islam J, Bhowmick S, Haque M, Rahaman A. Effects of *Syzygium cumini* seed extract on the memory loss of Alzheimer's disease model rats. *Adv Alzheimer's Dis* 2017;06:53–73. doi:10.4236/aad.2017.63005.
- [250] Ajiboye BO, Ojo OA, Akuboh OS, Okesola MA, Idowu OT, Oyinloye BE, *et al.* The protective effect of polyphenol-rich extract of *Syzygium cumini* leaves on cholinesterase and brain antioxidant status in alloxan-induced diabetic rats. *Jordan J Biol Sci* 2018;11:163–169.
- [251] Mahapatra PK, Chakraborty D, Chaudhuri AK. Anti-inflammatory and antipyretic activities of *Syzygium cumini*. *Planta Med* 1986;52(6):540. doi:10.1055/s-2007-969339, PMID:17345495.
- [252] Kumar A, Padmanabhan N, Krishnan MRV. Central nervous system activity of *Syzygium cumini* seed. *Pakistan J Nutr* 2007;6:698–700.
- [253] Rahaman R, Hossain S, Rahman M, Ibrahim H, Taslima N, Borhan U, *et al.* *Syzygium cumini* (L.) seed extract improves memory related learning ability of old rats in eight arm radial maze. *J Pharmacogn Phytochem* 2013;1:85–94.
- [254] Cargnelutti LO, Bitencourt PER, Bochi G, Duarte T, Boligon A, Pigatto AS, *et al.* *Syzygium cumini* leaf extract protects against ethanol-induced acute injury in rats by inhibiting adenosine deaminase activity and proinflammatory cytokine production. *Res J Phytochem* 2015;9:56–67.
- [255] Avila-Peña D, Peña N, Quintero L, Suárez-Roca H. Antinociceptive activity of *Syzygium jambos* leaves extract on rats. *J Ethnopharmacol* 2007;112(2):380–385. doi:10.1016/j.jep.2007.03.027, PMID:17478066.
- [256] Singh H, Sharma A, Bhardwaj A, Kaur B, Badial K, Kaur S, *et al.* Evaluation of antinociceptive effect of seed extracts of *Eugenia jambolana* Linn. *Plant Arch* 2019;19:3355–3361.
- [257] Nikhat F, Satyanarayana D, Suresh D, Purohit M, Raza H, Hamza S. Analgesic activity of the isolated constituent *Syzygium cumini*(L) skeel. *Res J Pharmacogn Phytochem* 2011;3:178–9.
- [258] Saha SS, Du Y, Sandha SS, Garcia LA, Jawed MK, Srivastava M. Inertial navigation on extremely resource-constrained platforms: methods, opportunities and challenges. *IEEE ION Position Locat Navig Symp* 2023;2023:708–723. doi:10.1109/plans53410.2023.10139997, PMID:37736264.
- [259] Kumar EK, Mastan SK, Reddy KR, Reddy GA, Raghunandan N. Chait-

- anya G. Anti-arthritis property of the methanolic extract of *Syzygium cumini* seeds. *Int J Integr Biol* 2008;4:55–62.
- [260] Venkataramanan RV, Raju CD, Sumithra M, Prakash MP, Goswami H. Evaluation of antirheumatic activity of petroleum ether extract of *Syzygium cumini* stem bark in rats. *Artic Biomed Pharmacol J* 2016;9:639–642. doi:10.13005/bpj/984, PMID:1356737.
- [261] Chandravarkar S, Desai SN. Diuretic activity of different extracts of bark of *Syzygium cumini* (Linn.) skeels. *Int J Res Ayurveda Pharm* 2014;5:102–4. doi:10.7897/2277-4343.05121.
- [262] Venkateshwarlu E, Bhava BSS, Kumar RS, Venkateshwar RJ, Gouthami E, Umasankar K. Evaluation of diuretic activity of *Syzygium cumini* and its effect on prostaglandin system. *Orient Pharm Exp Med* 2015;15:45–51. doi:10.1007/s13596-015-0179-5.
- [263] Sujitha K, Phani SA, Mohan RPM, Mohammed L, Srinivasarao K, Karuna SV. Preliminary screening of *Syzygium cumini* and *Achyranthes aspera* for their anthelmintic activity. *Res J Pharmacogn Phytochem* 2010;2:445–9.
- [264] Kavitha K, Murali M, Jayachandra K. Preliminary phytochemical screening, anthelmintic activity of methanolic and aqueous extract of *Syzygium cumini* Linn. bark (Myrtaceae). *J Pharm Sci Res* 2011;3:1460–5.
- [265] Jayamohan N, Kumar P, Jayachandra K. Surveillance of invitro antioxidant and anthelmintic activity of methanolic extract of *Syzygium cumini* bark (Myrtaceae). *Int J Phytopharm* 2013;3:56–62. doi:10.7439/ijpp.
- [266] Carolin N, Ajith Kumar B, Sudha N, Priya P, Sambath Kumar R. Invitro anthelmintic activity of *Mollugo nudicaulis* Lam, *Syzygium cumini* Linn and *Hibiscus vitifolius* Linn on Pheretima posthuma. *Am J Pharmtech Res* 2020;10(6)..
- [267] Oliveira L, Miranda J, Curado G, Costa Neto J, Santos B, Barros E, et al. In vivo anthelmintic activity of *Syzygium cumini* leaves against *Haemonchus contortus* in sheep. *Planta Med* 2012;78:P1413. doi:10.1055/s-0032-1321100.
- [268] Azam A, Saeed I, Saleem M, Azam T. Evaluation of anthelmintic activity of different fractions of *Syzygium cumini* L. leaves. *Fuust J Biol* 2020;10:51–5.
- [269] Mastan SK, Saraseeruha A, Gourishankar V, Chaitanya G, Raghunandan N, Reddy GA, et al. Immunomodulatory activity of methanolic extract of *Syzygium cumini* seeds. *Pharmacologyonline* 2008;3:895–903.
- [270] Kazmi SAJ, Riaz A, Akhter N, Khan RA. Evaluation of wound healing effects of *Syzygium cumini* and laser treatment in diabetic rats. *Pak J Pharm Sci* 2020;33:779–786. doi:10.36721/PJPS.2020.33.2.S UP.779-786.1.
- [271] Chaudhary A, Bag S, Banerjee P, Chatterjee J. Wound healing efficacy of Jamun honey in diabetic mice model through reepithelialization, collagen deposition and angiogenesis. *J Trad, Complementary Med* 2020;10(6):529–543. doi:10.1016/j.jtcme.2019.10.002.
- [272] Abdalla FH, Belle LP, Bitencourt PER, De Bona KS, Zanette RA, Boligon AA, et al. Protective effects of *Syzygium cumini* seed extract against methylmercury-induced systemic toxicity in neonatal rats. *BioMetals* 2011;24:349–56. doi:10.1007/s10534-010-9402-5.
- [273] Barai M, Ahsan N, Paul N, Hossain K, Rashid MA, Kato M, et al. Amelioration of arsenic-induced toxic effects in mice by dietary supplementation of *Syzygium cumini* leaf extract. *Nagoya J Med Sci* 2017;79:167–77. doi:10.18999/nagjms.79.2.167.
- [274] Prakash V, Adhikari A, Punetha H. Biochemical investigation on *Eugenia jambolana* (Jamun) bark extract in arsenic induced toxicity in rats. *Biotech Today* 2020;10(2):45–49. doi:10.5958/2322-0996.2020.00026.5.
- [275] Gautam N, Sehgal S, Gupta V, Gupta R. Hair growth activity of seeds and fruit pulp of *Eugenia jambolana* (Jamun). *Pharm Pharmacol Int J* 2015;2:184–187.
- [276] Arun R, Prakash MV, Abraham SK, Premkumar K. Role of *Syzygium cumini* seed extract in the chemoprevention of in vivo genomic damage and oxidative stress. *J Ethnopharmacol* 2011;134(2):329–333. doi:10.1016/j.jep.2010.12.014, PMID:21182920.
- [277] Jagetiya GC, Baliga MS. *Syzygium cumini* (jamun) reduces the radiation-induced DNA damage in the cultured human peripheral blood lymphocytes: a preliminary study. *Toxicol Lett* 2002;132(1):19–25.
- [278] Jagetiya GC, Shetty PC, Vidyasagar MS. Treatment of mice with leaf extract of jamun (*Syzygium cumini* Linn. Skeels) protects against the radiation-induced damage in the intestinal mucosa of mice exposed to different doses of γ -radiation. *Pharmacologyonline* 2008;1:169–195.
- [279] Jagetiya GC, Shetty PC. Augmentation of antioxidant status in the liver of swiss albino mice treated with jamun *Syzygium Cumini*, Skeels extract before whole body exposure to different doses of γ -radiation. *J Adv Res Biotech* 2016;1(1):1–13. doi:10.15226/2475-4714/1/1/00110.
- [280] Lichota A, Gwozdziński K. Anticancer Activity of Natural Compounds from Plant and Marine Environment. *Int J Mol Sci* 2018;19(11):3533. doi:10.3390/ijms19113533, PMID:30423952.
- [281] Barh D, Viswanathan G. *Syzygium cumini* inhibits growth and induces apoptosis in cervical cancer cell lines: a primary study. *Ecancermedicallscience* 2008;2:83. doi:10.3332/ecancer.2008.83, PMID:22275971.
- [282] Li L, Adams LS, Chen S, Killian C, Ahmed A, Seeram NP. *Eugenia jambolana* Lam. berry extract inhibits growth and induces apoptosis of human breast cancer but not non-tumorigenic breast cells. *J Agric Food Chem* 2009;57(3):826–831. doi:10.1021/jf803407q, PMID:19166352.
- [283] Afify AE-MR, Fayed SA, Shalaby EA, El-Shemy HA. *Syzygium cumini* (pomposia) active principles exhibit potent anticancer and antioxidant activities. *African J Pharm Pharmacol* 2011;5:948–956. doi:10.5897/AJPP10.420.
- [284] Tripathy G, Pradhan D. In-vitro anti breast cancer activity of *Syzygium cumini* against MCF-7 cell line. *J Innov Pharm Biol Sci* 2015;2:119–24.
- [285] Charepalli V, Reddivari L, Vadde R, Walia S, Radhakrishnan S, Vanamala JK. *Eugenia jambolana* (Java Plum) fruit extract exhibits anti-cancer activity against early stage human HCT-116 colon cancer cells and colon cancer stem cells. *Cancers (Basel)* 2016;8(3):29. doi:10.3390/cancers8030029, PMID:26927179.
- [286] Tripathy G, Pradhan D, Pradhan S, Dasmohapatra T. Evaluation of plant extracts against lung cancer using H460 cell line. *Asian J Pharm Clin Res* 2016;9:227–9.
- [287] Frauches NS, Montenegro J, Amaral T, Abreu JP, Laiber G, Junior J, et al. Antiproliferative activity on human colon adenocarcinoma cells and in vitro antioxidant effect of anthocyanin-rich extracts from peels of species of the myrtaceae family. *Molecules* 2021;26:564. doi:10.3390/molecules26030564.
- [288] Gibbert L, Sereno AB, Andrade MTP de, Silva MAB da, Miguel MD, Montruchio DP, et al. Nutritional composition, antioxidant activity and anticancer potential of *Syzygium cumini* (L.) and *Syzygium malaccense* (L.) fruits. *Res Soc Dev* 2021;10:e5210413743. doi:10.33448/rsd-v10i4.13743.
- [289] Khodavirdipour A, Zarean R, Safaralizadeh R. Evaluation of the anti-cancer effect of *Syzygium cumini* ethanolic extract on HT-29 colorectal cell line. *J Gastrointest Cancer* 2021;52(2):575–581. doi:10.1007/s12029-020-00439-3, PMID:32506290.
- [290] Ruthurusamy SK, Dheeba B, Hameed SS, Palanisamy S. Anti-cancer and anti-oxidative potential of *Syzygium cumini* against breast cancer cell lines. *J Chem Pharm Res* 2015;7:P449–460.
- [291] Ezhilarasan D, Apoorva VS, Ashok Vardhan N. *Syzygium cumini* extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells. *J Oral Pathol Med* 2019;48(2):115–121. doi:10.1111/jop.12806, PMID:30451321.
- [292] Debnath A, Salim M, Miah F, Karim R, Alam J, Medicine I. Evaluation of invertase inhibition activity and cytotoxicity of ethanol and acetone extracts of *Swietenia macrophylla* leaves, *Syzygium cumini* and *Trigonella foenum-graecum* seeds. *Tradit Integr Med* 2020;5:59–69.
- [293] Yadav SS, Meshram GA, Shinde D, Patil RC, Manohar SM, Upadhye M V. Antibacterial and anticancer activity of bioactive fraction of *Syzygium cumini* L. seeds. *HAYATI J Biosci* 2011;18:118–22. doi:10.4308/hjb.18.3.118.
- [294] Ogato MD, Mauti EM, Mauti OG, Ambrose B, Kowanga KD. Anticancer activity of *Eugenia jambolana* seeds against Hep2 cell lines. *J Phytopharm* 2015;4:295–8.
- [295] Sharma V, Heer A, Kour N, Sharma A, Singh SK. Karonda and Jamun seeds' in vitro anticancer efficacy. *Indian J Tradit Knowl* 2019;18:573–8.
- [296] Singh P, Bast F, Bhushan S, Mehra R, Kamboj P. Molecular docking and in vitro study of *Syzygium cumini*-derived natural compounds on receptor tyrosine kinases pathway components. *Int J Bioinform Res*

- Appl 2019;15:144–58. doi:10.1504/IJBRA.2019.099576.
- [297] Nawadkar AD, Jain BU. In-vitro anticancer activity of ethanolic extract of *Syzygium cumini* leaves. World J Pharm Res 2021;10:1252–7.
- [298] Mauti GO. Extracts of Jamun seeds inhibited the growth of human (Hep-2) cancer cells. J Cancer Res Ther 2024;20(1):189–192. doi:10.4103/jcrt.jcrt_638_22, PMID:38554319.
- [299] Eldin Elhawary SS, Elmotyam A, kamal E., Alsayed D, kamel, Zahran EM, Fouad MA, Sleem AA, et al. Cytotoxic and anti-diabetic potential, metabolic profiling and insilico studies of *Syzygium cumini* (L.) Skeels belonging to family Myrtaceae. Nat Prod Res 2022;36:1026–30. doi:10.1080/14786419.2020.1843032.
- [300] Iswariya G, Sankari Samk, Karpagam S. Efficiency of *Syzygium cumini* in the control of colon cancer using HT 29 colon cancer cells. Int J Curr Sci 2023;13:22–30.
- [301] Parmar J, Sharma P, Verma P, Goyal PK. Chemopreventive action of *Syzygium cumini* on DMBA-induced skin papillomagenesis in mice. Asian Pac J Cancer Prev 2010;11:261–265.
- [302] Parmar J, Sharma P, Verma P, Sharma P, Goyal PK. Elimination of deleterious effects of DMBA-induced skin carcinogenesis in mice by *Syzygium cumini* seed extract. Integr Cancer Ther 2011;10(3):289–297. doi:10.1177/1534735410385112, PMID:21147816.
- [303] Aqil F, Jeyabalan J, Munagala R, Singh IP, Gupta RC. Prevention of hormonal breast cancer by dietary jamun. Mol Nutr Food Res 2016;60(6):1470–1481. doi:10.1002/mnfr.201600013, PMID:27030099.
- [304] Rehman AA, Riaz A, Asghar MA, Raza ML, Ahmed S, Khan K. In vivo assessment of anticoagulant and antiplatelet effects of *Syzygium cumini* leaves extract in rabbits. BMC Complement Altern Med 2019;19:236. doi:10.1186/s12906-019-2661-y.
- [305] Ima RM, Polonini HC, Brandão MA, Raposo FJ, Dutra RC, Raposo NR. In vitro assessment of anti-aging properties of *Syzygium cumini* (L.) leaves extract. Biomed Ical J Ournal Sci Rnce Tech Res 2019;13:10185–91. doi:10.26717/BJSTR.2019.13.002447.
- [306] Sachdeva C, Mohanakrishnan D, Kumar S, Kaushik NK. Assessment of in vitro and in vivo antimalarial efficacy and GC-fingerprints of selected medicinal plant extracts. Exp Parasitol 2020;219:108011. doi:10.1016/j.exppara.2020.108011, PMID:33010286.
- [307] Bhong PN, Nilofar NS, Pratibha MR, Madhavi BS. In-vitro and in-vivo evaluation of anti-asthmatic activity of *Eugenia jambolana* bark. Res J Pharm Technol 2021;14(6):3337–3342. doi:10.52711/0974-360X.2021.00580.
- [308] Reyes-Farias M, Carrasco-Pozo C. The Anti-Cancer Effect of Quercetin: Molecular Implications in Cancer Metabolism. Int J Mol Sci 2019;20(13):3177. doi:10.3390/ijms20133177, PMID:31261749.
- [309] Jia Y, Wu C, Rivera-Piza A, Kim YJ, Lee JH, Lee SJ. Mechanism of action of cyanidin 3-O-glucoside in gluconeogenesis and oxidative stress-induced cancer cell senescence. Antioxidants (Basel, Switzerland) 2022;11:749. doi:10.3390/ANTIOX11040749.
- [310] Imran M, Saeed F, Hussain G, Imran A, Mehmood Z, Gondal TA, et al. Myricetin: A comprehensive review on its biological potentials. Food Sci Nutr 2021;9:5854–68. doi:10.1002/FSN3.2513.
- [311] Afroze N, Pramodh S, Hussain A, Waleed M, Vakharia K. A review on myricetin as a potential therapeutic candidate for cancer prevention. 3 Biotech 2020;10(5):211. doi:10.1007/s13205-020-02207-3, PMID:32351869.
- [312] Ballav S, Ranjan A, Basu S. Partial activation of PPAR- γ by synthesized quercetin derivatives modulates TGF- β 1-induced EMT in lung cancer cells. Adv Biol (Weinh) 2023;7(10):e2300037. doi:10.1002/adbi.202300037, PMID:37042092.
- [313] Albensi BC. What is nuclear factor kappa B (NF- κ B) doing in and to the mitochondrion? Front Cell Dev Biol 2019;7:154. doi:10.3389/fcell.2019.00154, PMID:31448275.